

Physicochemical Studies and Biological Activity of Mixed Ligand Complexes Involving Bivalent Transition Metals, 2-Aminomethylthiophenyl-4-Bromosalicylaldehyde Schiff Base and Glycine

A.A. El-Sherif^{1,2*} (Corresponding author)

1, Department of Chemistry, Faculty of Arts and Science, Northern Borders University, Rafha, Kingdom of Saudi Arabia

2, Department of chemistry, Faculty of Science, Cairo University, Cairo, Egypt.

Mutlaq S. Aljhdali

Department of Chemistry, Faculty of Science, King Abdulaziz University, Jeddah 21589, Kingdom of Saudi Arabia

Abeer T. AbdelKarim

Department of chemistry, Faculty of Science, Cairo University, Cairo, Egypt.

Mahmoud M. Ahmed

Department of Sciences and Mathematics, New-Valley Faculty of Education, Assiut University, Assiut, Egypt

Abstract

The Mixed-ligand complexes of Co(II), Ni(II), Mn(II) and Zn(II) with 2-aminomethylthiophenyl-4-bromosalicylaldehyde Schiff base (ATS) and glycine as a representative example of amino acids have been achieved. These complexes namely [Ni(ATS)(Gly)] (1), [Co(ATS)(Gly)].H₂O (3) [Mn(ATS)(Gly)].2H₂O (2) and [Zn(ATS)(Gly)] (4) were characterized by elemental, molar conductance, infrared, magnetic moment, and electronic spectra. ATS behaves as mononegatively charged bidentate ligand with coordination through azomethine nitrogen and phenolate oxygen groups while glycine acts as monobasic bidentate ligand is coordinated via amino and ionized carboxylate group after deprotonation. The low molar conductance values suggest the non-electrolytic nature of these complexes. The magnetic and spectral data indicates a square planar geometry for Ni²⁺ complex, tetrahedral geometry for Zn²⁺ and octahedral geometry for both Mn²⁺ and Co²⁺ complexes. The isolated chelates have been screened for their antimicrobial activity.

Keywords: Schiff base, Nickel(II); Glycine; Electronic Spectra; antibacterial activity.

1. Introduction

Amino acids constitute the building blocks of proteins and act as indispensable chemical species for performing a huge number of biological functions, as exemplified by the role of enzymes [1]. Also, amino acids have special importance among the other chemical groups since they are the foundation stones of the living organisms. The environment around the metal center 'as coordination geometry, number of coordinated ligands and their donor group' is the key factor for metalloprotein to carry out a specific physiological function [2]. About 20 zinc enzymes are known in which zinc is generally tetrahedrally four coordinate and bonded to hard donor atoms such as nitrogen or oxygen [3]. Manganese plays an important role in several biological redox-active systems [4,5]. The synthesis and reactivity of cobalt complexes of Schiff base ligands have always attracted the attention of inorganic chemists [6,7]. For instance, the cobalt complexes of tetradentate Schiff base ligands have been extensively used to mimic cobalamin (B₁₂) coenzymes [8-10], dioxygen carriers and oxygen activators [11-13]. The preparation of model complexes having similar spectroscopic features is perhaps the most important step to understand the structure and behavior of these biological systems. Schiff base metal complexes attract considerable interest and occupy an important role in the development of the chemistry of chelate systems [14,15] due to the fact that especially these with N₂O₂ tetradentate ligands, such systems closely resemble metallo-proteins. Survey of the literature reveals an excellent work devoted to synthesis and characterization of many metal complexes of Schiff base [16-20],

Additionally, it has been reported that Schiff bases and their first-row transition metal complexes can exhibit fungicidal, bactericidal, antiviral and antitumor activities in addition to their important roles in catalysis and organic synthesis [21-23]. Metal complexes of Schiff bases derived from substituted salicylaldehydes and various amines have been widely investigated because of their wide applicability [24-26] especially metal complexes of Schiff bases with heterocyclic compounds find applications as potential drugs [27]. Therefore, the synthesis of new Schiff bases and their complexes have been being a popular theme. The ternary complexes of the Schiff bases and amino acids with metal ions bear the importance of both Schiff bases and amino acids. In

continuation of our earlier work [28-33] and to wide the scope of investigation on the coordination behavior of Schiff base ligand and amino acids, we report here synthesis, characterization and biological activity of the ternary system including bivalent transition metal ions M(II), Schiff-base ligand (ATS) and glycine as a representative example of amino acids (Scheme 1). The complexes were characterized based on elemental analyses, IR, UV-vis, magnetic moment and molar conductance measurements. The isolated metal chelates were screened for their antibacterial and antifungal activities and the results are reported and discussed.



Scheme 1. Structural formulae of ATS-Schiff base and glycine amino acid.

2. Experimental

2.1. Materials

All chemicals used in this investigation were laboratory pure including $\text{CoCl}_2 \cdot 6\text{H}_2\text{O}$, $\text{NiCl}_2 \cdot 6\text{H}_2\text{O}$, $\text{MnCl}_2 \cdot 4\text{H}_2\text{O}$, $\text{ZnCl}_2 \cdot 6\text{H}_2\text{O}$, $\text{C}_2\text{H}_5\text{OH}$, DMSO, 5-bromosalicylaldehyde, glycine, and 2-aminomethylthiophene. All reagents were provided from Aldrich and Sigma chemicals companies.

2.2. Preparation of Schiff base

The Schiff base was prepared as reported in literature [35] by mixing an ethanolic solution (20 ml) of 2.01g (0.01 mol) of 5-bromosalicylaldehyde with 1.13g (0.01 mol) of 2-aminomethylthiophene in the same volume of ethanol. The mixture then refluxed with stirring for 1 hour. The precipitate was collected by filtration through buchner funnel, recrystallized from ethanol and dried at room temperature with 89 % yield.

2.3. Preparation of the solid complexes

All complexes were prepared by refluxing an ethanolic solution of the Schiff base ligand (1 mmol, 0.295 g) and glycine (1 mmol, 0.075 g Gly) 1with an ethanolic solution of metal salt (1 mmol, $\text{CoCl}_2 \cdot 6\text{H}_2\text{O}$, 0.238 g; $\text{NiCl}_2 \cdot 6\text{H}_2\text{O}$, 0.237 g; $\text{MnCl}_2 \cdot 4\text{H}_2\text{O}$, 0.1979g, $\text{ZnCl}_2 \cdot 6\text{H}_2\text{O}$, 0.2429g) in the molar ratio 1:1:1 (w:w:w). An equivalent amount of NaHCO_3 was added to neutralize the released protons. The reaction mixtures were refluxed on a water bath for 4 h and allowed to cool to room temperature overnight. The precipitated complexes were then filtered off, washed with petroleum ether and dried overnight in a vacuum desiccator. The dried complexes were subjected to elemental and spectroscopic analysis.

2.4. Biological activity

The antimicrobial bioassay was performed according to protocols described previously using a modified Kirby-Bauer disc diffusion method [35-37]. The antimicrobial activities of metal complexes were studied against Gram (+) bacteria as (*Staphylococcus epidermidis* and *Bacillus cereus*); Gram (-) bacteria as (*Pseudomonas aeruginosa*, *Escherichia coli*) and fungi as *Aspergillus fumigatus* and *Aspergillus niger*. Standard discs of *Ciprofloxacin* (antibacterial agent), *Ketoconazole* (antifungal agent) served as positive controls for antimicrobial activity but filter discs impregnated with 10 μl of solvent (DMSO) were used as a negative control.

2.5. Instruments

The microchemical analysis of the separated solid chelates for C, H and N were performed in the microanalytical Center, Cairo University. The analyses were performed twice to check the accuracy of the analyses data. Infrared spectra were recorded on a 8001-PC FTIR Shimadzu spectrophotometer using KBr pellets. The solid reflectance spectra were measured on a Shimadzu 3101 pc spectrophotometer. The molar conductance of the complexes was measured for 1.00×10^{-3} M DMSO solutions at $25 \pm 1^\circ\text{C}$ using a systronic conductivity bridge type 305. The room temperature magnetic susceptibility measurements for the complexes were determined by the Gouy balance using $\text{Hg}[\text{Co}(\text{SCN})_4]$ as a calibrant.

3. Results and Discussion

The condensation of 5-bromosalicylaldehyde with 2-aminomethylthiophene in boiling ethanol yields a Schiff base compound (ATS) as reported in literature [34]. The elemental analyses for the interaction of Schiff base ligand and amino acids with metal (II) chloride in EtOH agree well with a 1:1:1 M(II):ATS:Gly stoichiometry for all the complexes. The results of elemental analysis (C, H, N and S), along with molecular formulae and percent yield of the complexes are presented in Table 1. The complexes are air stable, insoluble in the most

organic solvents and water but freely soluble in DMF and DMSO. The complexes have higher melting points than their corresponding ligands indicating that they are thermally stable. This could be attributed to the formation of chelate rings.

3.1. IR Spectra and mode of bonding

The results of IR measurements were listed in Table 2 with assignments for the most of the major peaks. The IR spectrum of the free Schiff base ligand (ATS) reveals a band at 3458 cm^{-1} due to $\nu(\text{OH})$. This band is absent in the spectra of metal(II) complexes, indicating coordination through the deprotonated phenolic OH group [38]. Moreover, it was evidenced from the shift in the position of ν_{CO} (aromatic carbon and phenolic oxygen) at 1319 cm^{-1} [39] to the lower frequency region in the spectra of complexes [40]. However, the strong band observed at 1675 cm^{-1} in the free ATS ligand due to azomethine group vibration is shifted towards lower frequencies in the complexes by $(28\text{-}37\text{ cm}^{-1})$, suggesting that azomethine group is involved in coordination [41-43]. In the free Schiff base ligand, the sharp band observed at 825 cm^{-1} due to $\nu(\text{C-S-C})$ stretching frequency of thiophene ring remained unchanged in all complexes confirming the noninvolvement of the thiophene sulfur in complex formation [44]. The appearance of new bands in the low frequency ranges at $501\text{-}525$ and $445\text{-}412\text{ cm}^{-1}$ due to $\nu(\text{M-O})$ and $\nu(\text{M-N})$ vibrations [45] respectively, support the participation of the nitrogen atom of the azomethine group and oxygen of the OH group of the Schiff-base ligand in the complexation with metal(II) ion [45]. The amino acid was coordinated as bidentate and bound to the central metal ion through the carboxylate and the amino groups. The δNH_3^+ band, which is characteristic of zwitter ion, disappears in the spectra of complexes. This indicates that the NH_2 group must be involved in coordination. This is supported by the appearance of stretching vibration of amino group as split bands in the regions $3424\text{-}3410$ and $3202\text{-}3190\text{ cm}^{-1}$ [46] assigned for the coordinated amino group [46,47]. One particularly interesting aspect to be analyzed in complexes of this type is the displacement of the characteristic carboxylic bands after coordination. This peculiar aspect has been summarized in Table 2. The "free" amino acid exists as zwitter ion in the crystalline state; thus, one expects two stretching vibrations for the COO^- moiety present in these systems, namely $\nu_{\text{s}}(\text{COO}^-)$ and $\nu_{\text{as}}(\text{COO}^-)$. The first one is usually of medium intensity in the IR spectrum, whereas the second is strong and broad. The bands in the regions 1608 and 1412 cm^{-1} , due to $\nu_{\text{asym}}(\text{COO}^-)$ and $\nu_{\text{sym}}(\text{COO}^-)$ of the amino acids appear in the complexes at $1587\text{-}1591$ and $1379\text{-}1382\text{ cm}^{-1}$. The frequency difference between asymmetric and symmetric vibration of carboxylate is found to be $> 200\text{ cm}^{-1}$ reflecting the monodentate coordination of carboxylate group of amino acid with the metal ion in the synthesized complexes [48]. The shift of these two bands suggests the involvement of the carboxylic group of the amino acids in the complex formation.

This conclusion is supported further by the absence of the band at 1705 cm^{-1} in the IR spectra of the metal complexes, which is due to the $\nu_{\text{asym}}(\text{C=O})$ mode associated with the bidentate coordinated $-\text{COOH}$ groups of the amino acid ligand. Hence, the amino acid moiety is chelated to the metal ion forming five-membered chelate ring. The OH stretching frequency appears in the spectra of M(II) complexes around $3518\text{-}3530\text{ cm}^{-1}$ is attributed the presence of water of hydration. Also, according to Stefov et al. [49], coordinated water should exhibit frequencies at 825 , 575 and 500 cm^{-1} . The absence of spectral bands in these regions in the spectra of M(II) complexes indicates that the water molecules in these complexes are not coordinated but is present as lattice water.

As a general conclusion, the Schiff base participated in bonding to metal(II) ion as monobasic bidentate (N,O donor) ligand and the amino acids also as monobasic bidentate ligands with a total of four coordination sites around the M(II) ion whose charge is neutralized by the deprotonation of the Schiff base phenolic OH and the amino acid COOH. The nonelectrolytic nature of the complexes was evidenced from the low values of the molar conductance of the complexes measured in DMSO [50] (Table 3).

3.2. Electronic spectra and magnetic moment measurements

3.2.1. Electronic spectra

As the result of failure to obtain a single crystal for X-ray analyses to confirm the geometric structure for these complexes, solid reflectance spectra and magnetic moment measurements are used for this purpose. The solid reflectance spectra of metal complexes show different bands at different wavelengths, each one is corresponding to certain transition which suggests the geometry of the complex compounds. The complexes show two bands in the $43450\text{-}44230$ and $32294\text{-}36712\text{ cm}^{-1}$ ranges which are assigned to intraligand transition [51].

The Ni(II) complex (1) has a diamagnetic character (no unpaired electrons). This suggests a square-planar geometry around nickel(II) [52]. The electronic spectrum of the Ni(II) complex (1) showed two bands for complex (1). The two bands in the spectrum of the nickel(II) complex observed at $14,122$ and $24,165\text{ cm}^{-1}$ which may be assigned to the $^1\text{A}_{1g} \rightarrow ^1\text{B}_{2g}$ and $^1\text{A}_{1g} \rightarrow ^1\text{B}_{1g}$ transitions, respectively, for a square-planar geometry around Ni(II) [53].

The electronic spectrum of the mononuclear Co(II) complex (2) showed a strong band at 15826 cm^{-1} which is assigned to the $^4\text{A}_2(\text{F}) \rightarrow ^4\text{T}_1(\text{P})$ transition, suggesting tetrahedral geometry around Co(II) ion as reported

in literature [54,55]. The measured magnetic moment of Co(II) complex (2) is given in Table 3. The theory of magnetic susceptibility of Co(II) ion was given by Schlapp and Penney [56] and the best summary of results on the magnetism behavior of cobalt(II) compounds is that of Figgis and Nyhlom [57]. The observed values of magnetic moments for cobalt (II) complexes are generally diagnostic of the coordination geometry about the metal ion. The effective magnetic moment value (μ_{eff}) for the mononuclear Co(II) complex (2) is 4.11 B.M. (Table 3), which is lower than the expected range for tetrahedral cobalt(II) complexes (4.2-4.7 B.M.) according to the literature cited values [58-60]. The lower magnetic moment value of the [Co(ATS)(Gly)] complex would point to a not purely tetrahedral geometry of cobalt(II) and a tendency to be distorted [61].

The magnetic moment value of Mn^{2+} complexes under study is found to be 5.62 B.M. The solid reflectance data of Mn^{2+} complex show the observed bands at 12210, 17627 and 19190 cm^{-1} attributable to ${}^6\text{A}_1 \rightarrow {}^4\text{T}_1$, ${}^6\text{A}_1 \rightarrow {}^4\text{T}_2$ (G) and ${}^6\text{A}_1 \rightarrow {}^4\text{E}$ transitions, respectively suggest a tetrahedral stereochemistry for these complexes [62].

Due to the d^{10} electronic configuration of the metallic Zn(II) ion, electronic spectra of the Zn(II)-complex do not allow suggestion concerning its stereochemistry. The electrolytic nature of the complex and the number of donor atoms of the ligands and in analogy with those described for Zn(II) complexes containing N-O donors [63,64], it is likely to be tetrahedral in accordance with the preference of Zn(II) for tetrahedral coordination [65].

3.3. Conductivity measurements

The chelates were dissolved in DMSO and the molar conductivities of 10^{-3} M of their solutions at 25 ± 1 °C were measured. As seen from Table 3, the molar conductivity values for M(II)-chelates are 11.9-13.8 $\Omega^{-1} \text{cm}^2 \text{mol}^{-1}$ indicating nonelectrolytic nature of the complexes.

3.4. Structure of the complexes

It is concluded that from elemental analysis, IR, conductance, electronic spectra, magnetic and conductance measurements, the ATS-Schiff base behaves as a monobasic bidentate ligand coordinated to the M(II) ion through the deprotonated hydroxo group and the azomethine-N atom while amino acid acts as a monobasic bidentate ligand coordinated through the amino and carboxylate groups. On the basis of the elemental analysis and spectral data square planar geometry is suggested for Ni(II) and Zn(II) complexes in addition to tetrahedral geometry for both Co(II) and Mn(II) complexes as shown in scheme 2.

3.5. Biological activity

3.5.1. Antibacterial activity

To assess the biological potential of the synthesized compounds, the metal complexes were tested against different species of bacteria. In testing the antimicrobial activity of these compounds, we used more than one test organism to increase the chance of detecting antibiotic principles in tested materials. The organisms used in the present investigations included two Gram positive (*Staphylococcus epidermidis* and *Bacillus cereus*) and two Gram negative (*Escherichia coli*, *Pseudomonas aeruginosa*). The disc diffusion method was used to evaluate the antibacterial activity of the synthesized mixed ligand complexes. The medium used for growing the culture was nutrient agar. The results of the antimicrobial activities of the synthesized compounds are recorded in Table 4. The biological activity of complexes may be arising from the hydroxyl groups which may play an important role in the antibacterial activity [66]. The mode of action of the compounds may involve formation of a hydrogen bond through the azomethine group ($>\text{C}=\text{N}-$) with the active centers of various cellular constituents, resulting in interference with normal cellular processes [67]. It has been suggested that the ligands with nitrogen and oxygen donor systems inhibit enzyme activity, since the enzymes which require these groups of their activity appear to be especially more susceptible to deactivation by metal ions on coordination. Moreover, coordination reduces the polarity of the metal ion mainly because of the partial sharing of its positive charge with the donor groups [68] within the chelate ring system formed during coordination. This process, in turn, increases the lipophilic nature of the central metal atom, which favors its permeation more efficiently through the lipid layer of the microorganism [69], thus destroying them more aggressively. The antibacterial activity can be ordered as $[\text{Zn}(\text{ATS})(\text{Gly})] > [\text{Ni}(\text{ATS})(\text{Gly})] > [\text{Co}(\text{ATS})(\text{Gly})] > [\text{Mn}(\text{ATS})(\text{Gly})]$ (Fig. 1), suggesting that the lipophilic behaviour increases in the same order. This result is in accordance with the stabilities of these complexes.

3.5.2. Antifungal activity

The antifungal activities of the synthesized complexes were tested against *Aspergillus niger* and *Aspergillus fumigatus* test organisms in vitro using disc-diffusion method. The Ketoconazole was used as standards drug. The susceptibility of the selected strains of fungi towards the synthesized complexes was judged by measuring the diameter of the formed zone of inhibition. The results are given in Table 4. Metal ions are adsorbed on the cell walls of the microorganisms, disturbing the respiration processes of the cells and thus blocking the protein synthesis that is required for further growth of the organisms. Hence, metal ions are essential for the growth-

inhibitory effects [70]. According to Overtone's concept of cell permeability, the lipid membrane that surrounds the cell favors the passage of only lipid-soluble materials, so lipophilicity is an important factor controlling the antifungal activity. Upon chelation, the polarity of the metal ion will be reduced due to the overlap of the ligand orbitals and partial sharing of the positive charge of the metal ion with donor groups. This increased lipophilicity facilitates the penetration of the complexes into lipid membranes, further restricting proliferation of the microorganisms. In general, the [Mn(ATS)(Gly)] compound was found to be less effective against the selected fungal strains and [Zn(ATS)(Gly)] complex is the most active one among the synthesized bivalent metal(II)-complexes. Thus, the antifungal activity can be ordered as [Zn(ATS)(Gly)] > [Ni(ATS)(Gly)] > [Co(ATS)(Gly)] > [Mn(ATS)(Gly)] (Fig. 2). The order of antifungal activity is in accordance with the stability order of metal ions with chloro and bromo amino acid salicylaldehyde Schiff bases [71].

4. Conclusions

The present paper reports on the synthesis, characterization and electronic absorption spectra of mixed-ligand complexes of M(II) with Schiff base ligand (ATS) and glycine as a representative example of amino acids. The synthetic procedure in this work resulted in the formation of complexes in the molar ratio (1:1:1) (M^{II}:ATS:Gly). The newly synthesized Schiff base participated in bonding to metal as monobasic bidentate ligand through the azomethine nitrogen and phenolic oxygen atom via deprotonation forming stable six membered heterocyclic ring. The glycine amino acid also acts as monobasic bidentate ligands via amino and ionized carboxylate groups. Thus, the charge of metal ion was neutralized by the deprotonation of the Schiff base phenolic-OH group and the amino acid carboxylate group with a total of four coordination sites around the metal. From conductance measurements all complexes are non-electrolytes. The antimicrobial study reveals that some of the synthesized complexes show better activity or comparable activity to the standard drug antibiotic.

References

- [1] K.H. Zimmermann, An Introduction to Protein Informatics, Kluwer Academic Publishers, London, 2003.
- [2] R. Klement, F. Stock, H. Elias, H. Paulus, P. Pelikan, M. Valko, M. Mazur, (1999) *Polyhedron* 18, 3617.
- [3] F. Marchetti, C. Pettinari, R. Pettinari, A. Cingolani, D. Leanesi, A. Lorenotti, (1999) *Polyhedron* 18, 3041.
- [4] C. Zhacg, J. Sun, X. Kong, C. Zhao, (2000) *J. Coord. Chem.* 50, 353.
- [5] P.G. Ramappa, (1999) *J. Indian Chem. Soc.* 76, 235.
- [6] S. Yamada, (1999) *J. Coord. Chem. Rev.* 190–192, 537.
- [7] M. Amirnasr, K.J. Schenk, A. Gorji, R. Vafazadeh, (2001) *Polyhedron* 20, 695.
- [8] R. Cmi, S.J. Moore, L.G. Marzilli, (1998) *Inorg. Chem.* 37, 6890.
- [9] S.M. Polson, R. Cini, C. Pifferi, L.G. Marzilli, (1997) *Inorg. Chem.* 36, 314.
- [10] S. Hirota, E. Kosugi, L.G. Marzilli, O. Yamauchi, (1998) *Inorg. Chim. Acta* 275, 90.
- [11] A.E. Martell, D.T. Sawyer, Oxygen Complexes and Oxygen Activation by Transition Metals, Plenum Press, New York, 1988.
- [12] N.J. Henson, P.J. Hay, A. Redondo, (1999) *Inorg. Chem.* 38, 1618.
- [13] C. Benchini, R.W. Zoeliner, (1997) *Adv. Inorg. Chem.* 44, 263.
- [14] P.L. Gurian, L.K. Cheatham, J.W. Ziller, A.R. Barron, (1991) *J. Chem. Soc. Dalton Trans* 1449.
- [15] E. Labisbal, J.A. Garcia-Vazquez, J. Romero, S. Picos, A. Sousa, (1995) *Polyhedron* 14, 663.
- [16] T. Gupta, M.K. Saha, S. Sen, S. Mitra, A.J. Edwards, W. Clegg, (1999) *Polyhedron* 18, 197.
- [17] S. Zolezzi, Adecinti, E. Spodine, (1999) *Polyhedron* 18, 897.
- [18] A. Pasini, F. Demartin, O. Piovesana, B. Chiari, A. Cinti, O. Crispu, (2000) *J. Chem. Soc. Dalton Trans.* 3467.
- [19] N. Rajaiiah Sangeetha, S. Pal, (2000) *Polyhedron* 19, 1593–1600.
- [20] E.I. Jouad, A. Riou, M. Allain, M.A. Khan, G.M. Bouet, (2001) *Polyhedron* 20, 67.
- [21] M. Nath, S. Pokharia, R. Yadav, (2001) *Coord. Chem. Rev.* 215, 99.
- [22] H. Sharghi, M. A. Nasser, (2003) *Bull. Chem. Soc. (Jpn)* 76, 137.
- [23] W. T. Gao, Z. Zheng, (2002) *Molecules* 7, 511.
- [24] E. Canpolat, M. Kaya, (2004) *J. Coord. Chem.* 57, 127.
- [25] Y. Fan, C. Bi, J. Li, (2003) *Synth. React. Inorg. Met.-Org. Chem.* 33, 137.
- [26] E. Canpolat, M. Kaya, A. Yazici, (2005) *Spectrosc. Lett.* 38, 35.
- [27] S.S. Konstantinovi, B.C. Radovanovi, Z. Caki, V. Vasic, (2003) *J. Ser. Chem. Soc.* 68, 641.
- [28] B. Jeragh, D. Al-Wahaib, A.A. El-Sherif, A. El-Dissouky, (2007) *J. Chem. Eng. Data*, 52, 1609-1614.
- [29] A. A. El-Sherif, (2006) *J. Solution Chem.* 35, 1287-1301.
- [30] M.M.A. Mohamed, A.A. El-Sherif, (2010) *J. Solution Chem* 39, 639-653.

- [31] A.A. El-Sherif, M.M. Shoukry, (2005) *J. Coord. Chem.* 58(16), 1401-1415.
- [32] A.A. El-Sherif, M.M. Shoukry, (2007) *J. Spectrochim. Acta (A)* 66, 691-700.
- [33] A.A. El-Sherif, M.M. Shoukry, (2007) *Inorg. Chim. Acta* 360, 473-487.
- [34] A.A. El-Sherif, T.M. Eldebss, (2011) *J. Spectrochim. Acta (A)* 79, 1803.
- [35] Pfaller, M.A.; Burmeister, L.; Bartlett, M.A.; Rinaldi, M.G. (1988) *J. Clin. Microbiol.*, 26 , 1437-1441.
- [36] L.D. Liebowitz, H.R. Ashbee, E.G.V. Evans, Y. Chong, N. Mallatova, M. Zaidi, D. Gibbs, (2001) *Microbiol. Infet. Dis.* 24 , 27-33.
- [37] National Committee for Clinical Laboratory Standards, Methods for dilution antimicrobial susceptibility tests for bacteria that grow aerobically. Approved standard M7-A3, Villanova, Pa (1993).
- [38] Z. M. Zaki, S. S. Haggag, A. A. Soayed, (1983) *Spectroscopy Letters* 31(4) ,757.
- [39] A. Kulkarni, S. A. Patil, P.S. Badami, (2009) *Eur. J. Med. Chem.* 44, 2904-2912.
- [40] S. Sarkar, K. Dey, (2005) *J. Spectrochimica Acta A* 62 ,383-393.
- [41] E. Campolat, (2005) *Polish J. Chem.* 79, 619.
- [42] D.X. West, A.A. Nassar, (1998) *Trans. Met. Chem.* 23, 321.
- [43] G. B. Bagihalli, P.G. Avaji, S.A. Patil and P.S. Badami, (2008) *Eur. J. Medicinal Chem.*, 43 , 2639-2649.
- [44] S. Gronowitz, A.R. Katritsky, R.E. Reavill, (1963) *J. Chem. Soc.* 3881.
- [45] K.C. Satpathy, A.K. Panda, R. Mishra, A.P. Chapdar, S.K. Pradhan, (1994) *J. Indian Chem. Soc.* 71 , 593.
- [46] K. Nakamoto, Infrared Raman Spectra of Inorganic and Coordination Compounds, Third ed., John Wiley and Sons, New York, 1992.
- [47] V. Aletras, N. Hadjiliadis, B. Lippert, (1992) *Polyhedron* 11 , 1359.
- [48] G. B. Decon and R. J. Philips, (1980) *Coord. Rev.*, 33, 227.
- [49] V. Stefov, V.M. Petrusovski, B. Soptrajanov, (1993) *J. Mol. Struct.* 293, 97.
- [50] M. Aljahdali, A.A. EL-Sherif, (2013) *Inorg. Chim. Acta* 407 ,58-68.
- [51] A.A. El-Sherif, M.M. Shoukry, M.M.A. Abd-Elgawad, (2012) *J. Spectrochim. Acta (A)* 98, 307–321.
- [52] B. Prabbakar, K. Reddy, P. Lingaiat, (1988) *Indian, J. Chem.* 27A ,217.
- [53] D.N. Sathyanarayana, Electronic Absorption Spectroscopy and Related Techniques, Orient Longman Limited, Universities Press (India) Limited, 2001.
- [54] A. Shrivastav, N. Kishore, S.M. Singh, (2003) *Biometals* 16 (2), 311–320.
- [55] S.M.E. Khalil, H.F.O. El-Shafiy, (2000) *Synth. React. Inorg. Met. – Org. Chem.* 30 ,1817–1826.
- [56] R. Schlapp, W.G. Penney, (1932) *Phys. Rev.* 42 , 666–686.
- [57] B.N. Figgis, R.S. Nyhlom, (1958) *J. Chem. Soc.* 4190–4200.
- [58] K.A.R. Salib, A.A. Saleh, S. Abu El-Wafa, H.F.O. El-Shafiy, (2003) *J. Coord. Chem.* 56 (4) , 283–291.
- [59] A.A. El-Sherif, (2010) *J. Solution Chem.* 39 ,1562–1581.
- [60] A. A. El-Sherif, (2010) *J. Solution Chem* 39, 1562–1581
- [61] Bailar, J.C., Emeleus, H.J., Nyhlom, R., Trotmon-Dickenson, A.F.: *Comprehensive Inorg. Chem.*, vol. 3. Pergamon Press, New York (1975). pp. 1048, 1049, 1089, 1093, 1153
- [62] R.C. Mishra, B.K. Mohapatra, D. Panda, (1983) *J. Indian Chem. Soc.* 60, 782.
- [63] Sanmartin, J.; Bermejo, M.R.; Deibe, A.M.G.; Maneiro, M. ; Lage, C.; Filho, A.J.C. (2000) *Polyhedron*, 19, 185.
- [64] Krzyminiewska, V.P.; Litkowska, H.; Paryzek, W.R. (1999) *Monatsh, Chem.*, 130, 243-249.
- [65] Melnik, M.; Gyoryova, K.; Skorsep, J.; Holloway, C.E.(1995) *J. Coord. Chem.*, 35, 179-185.
- [66] S. Çakir, E. Bicer, M. Odabaşoğlu, Ç.Albayrak, (2005). *Braz. Chem. Soc.* 4, 711.
- [67] A. L. Koch, *Clin, Microbiol. Rev.* 16 (2003) 673.
- [68] Z.H. Chohan, A. Scozzafava, C.T. Supran, (2003) *J. Enzyme Inhib. Med. Chem.* 17 , 261.
- [69] Z.H. Chohan, K.M. Khan, C.T. Supuran, (2004) *Appl. Organomet. Chem.* 18 , 305.
- [70] I. Pal, F. Basuli, and S. Bhattacharya, Proceedings of the Indian Academy of Sciences: (2002) *Chemical Sciences*, 114(4) ,255–268.
- [71] M. Robert, R.J. Motekaitis, A.E. Martell. (1985) *Inorg. Chim. Acta*, 103 73.

Table 1. Analytical and physical data of the synthesized compounds

Compound	Mwt	% Yield	% Found (Calc.)			
			C	H	N	S
[Ni(ATS)(Gly)](1)	425.92	75 %	39.21(39.29)	3.01(3.06)	6.48(6.55)	7.47(7.52)
[Co(ATS)(Gly)].H ₂ O	444.94	77 %	37.71(37.75)	2.42(2.47)	6.22 (6.29)	7.16(7.19)
(2)						
[Mn(ATS)(Gly)].2H ₂ O	460.20	74 %	36.51(36.54)	3.69(3.72)	6.07(6.09)	6.91(6.95)
(3)						
[Zn(ATS)(Gly)] (4)	431.85	62 %	39.21(38.69)	2.98(3.01)	6.42(6.45)	7.35(7.38)

Table 2. Tentative assignment of the important infrared bands of the synthesized complexes.

Ligand/ complex	$\nu(\text{cm}^{-1})$										
	ν_{OH}	$\nu_{(\text{C}=\text{N})}$	$\delta_{\text{NH}_3^+}$ (Free AA)	$\nu_{(\text{C}-\text{O})}$ phenolic	ν_{asym} (COO)	ν_{sym} (COO)	$\Delta\nu$	ν_{NH_2} (Coord. AA)	$\nu_{(\text{C}-\text{S})}$	$\nu_{\text{M}-\text{N}}$	$\nu_{\text{M}-\text{O}}$
ATS	3458	1675	-	1319	-	-	-	-	825	-	-
Gly	-	-	1585	-	1608	1412	-	-	-	-	-
(1)	-	1638	-	1282	1590	1379	211	3420; 3190	824	445	525
(2)	3518	1635	-	1280	1591	1382	209	3410; 3202	826	428	512
(3)	3530	1640	-	1285	1589	1381	208	3415; 3198	825	412	505
(4)	-	1647	-	1270	1587	1380	207	3414; 3200	826	418	501

ATS = 2-aminomethylthiophenyl-4-bromosalicylaldehyde; Gly = glycine.

Table 3. Molar conductance, magnetic moment, electronic spectra and electron spin resonance data of [M(ATS)(Gly)] complexes.

Complex	Λ_M^a	$M_{\text{eff.}}$ (B.M.)	$\lambda_{\text{max}} (\text{cm}^{-1})$			Assignment
			$n \rightarrow \pi^*$	$\pi \rightarrow \pi^*$	d-d transition	
ATS	-	-	28409	40816	-	-
(1)	13.8	Diamag.	32294	43450	14122	$^1A_{1g} \rightarrow ^1B_{2g}$ $^1A_{1g} \rightarrow ^1B_{1g}$
(2)	12.2	5.11	34524	43780	15826	$^4A_2(\text{F}) \rightarrow ^4T_1(\text{P})$
(3)	11.9	5.62	36712	44230	12210	$^6A_1 \rightarrow ^4T_1$ $^6A_1 \rightarrow ^4T_2 (\text{G})$ $^6A_1 \rightarrow ^4E$
(4)	12.8	Diamag.	35860	44010	-	-

^aMolar conductance measured for 10^{-3} M DMSO solution, $\Omega^{-1} \text{cm}^2 \text{mol}^{-1}$; Gly= Glycine and ATS = 2-aminomethylthiophenyl-4-bromosalicylaldehyde.

Table 4. Antibacterial and antifungal activities of [M(ATS)(Gly)]^a complexes.

Compounds	Diameter of inhibition zone (in mm) ^b					
	Bacteria				Fungi	
	(G ⁻)		(G ⁺)		Aspergillus niger	Aspergillus fumigatus
	Pseudomonas aeruginosa	Escherichia Coli	Bacillus cereus	Staphylococcus epidermidis		
Conc. (µg/ml)	100	100	100	100	100	100
(1)	13.8± 0.11	11.5± 0.31	19.3 ± 0.17	20.5 ± 0.54	23.6 ± 0.15	23.2 ± 0.22
(2)	12.7± 0.49	NA	18.4 ± 0.13	19.1 ± 0.75	19.6 ± 0.23	20.2 ± 0.32
(3)	11.3± 0.63	NA ^c	15.1 ± 0.47	16.3 ± 0.34	16.7± 0.52	18.3 ± 0.12
(4)	19.9± 0.24	17.2 ± 0.15	23.5 ± 0.21	27.9 ± 0.67	25.7± 0.35	26.3 ± 0.44
Standard ^d	22± 0.15	27± 0.18	24± 0.28	28± 0.14	28 ± 0.12	30 ± 0.38

^aM = Ni(II), Co(II), Mn(II) and Zn(II), ^aATS = 2-aminomethylthiophenyl-4-bromosalicylaldehyde, Gly = glycine, ^bMean zone of inhibition in mm ± standard deviation beyond well diameter (6 mm) produced on a range of environmental and clinically pathogenic microorganisms. ^cNA: not detected, ^dCiprofloxacin is used as standard for antibacterial activity and Ketoconazole is used as standard for antifungal activity.

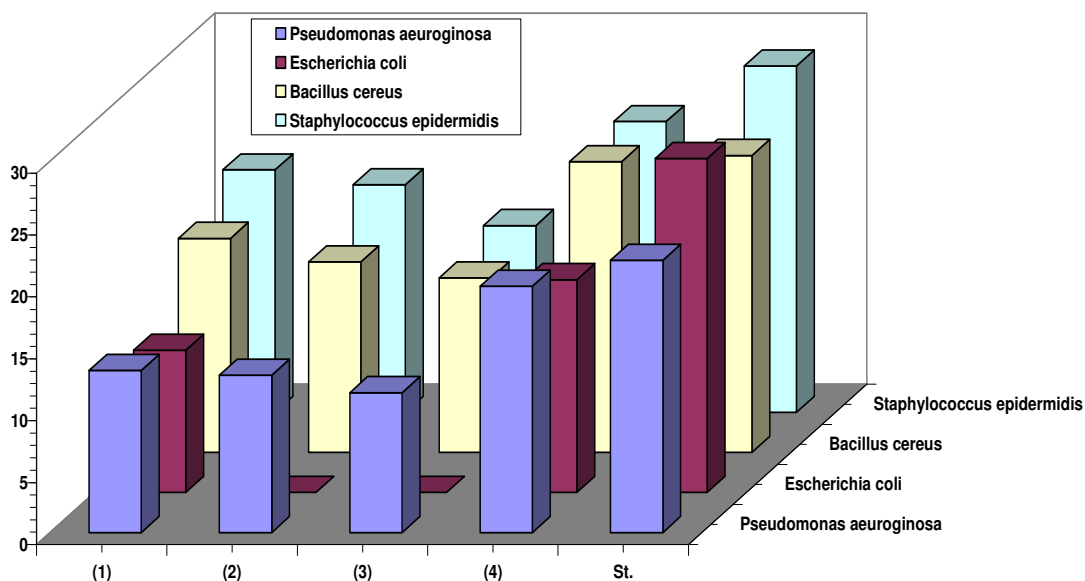


Fig.1. Antibacterial activity of M(II)-complexes towards different types of bacterial strains.

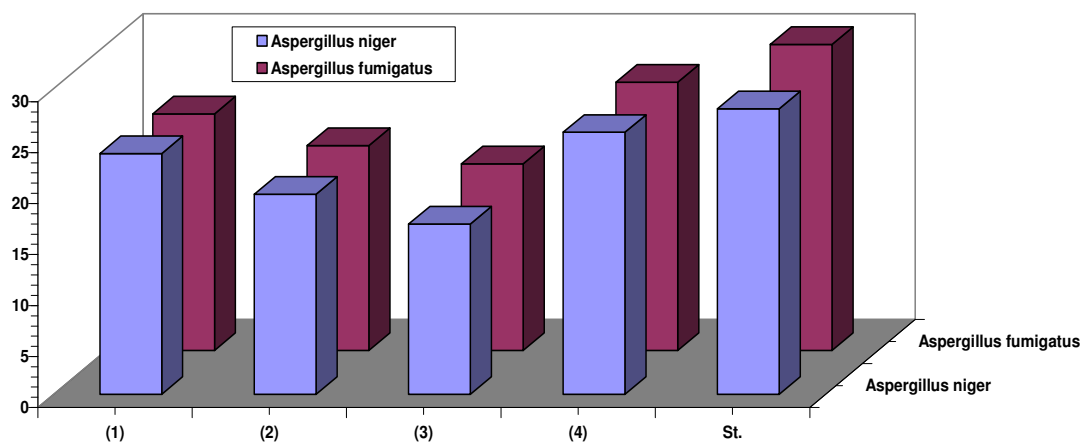


Fig.2. Antifungal activity of M(II)-complexes towards different types of fungal strains

The IISTE is a pioneer in the Open-Access hosting service and academic event management. The aim of the firm is Accelerating Global Knowledge Sharing.

More information about the firm can be found on the homepage:

<http://www.iiste.org>

CALL FOR JOURNAL PAPERS

There are more than 30 peer-reviewed academic journals hosted under the hosting platform.

Prospective authors of journals can find the submission instruction on the following page: <http://www.iiste.org/journals/> All the journals articles are available online to the readers all over the world without financial, legal, or technical barriers other than those inseparable from gaining access to the internet itself. Paper version of the journals is also available upon request of readers and authors.

MORE RESOURCES

Book publication information: <http://www.iiste.org/book/>

Academic conference: <http://www.iiste.org/conference/upcoming-conferences-call-for-paper/>

IISTE Knowledge Sharing Partners

EBSCO, Index Copernicus, Ulrich's Periodicals Directory, JournalTOCS, PKP Open Archives Harvester, Bielefeld Academic Search Engine, Elektronische Zeitschriftenbibliothek EZB, Open J-Gate, OCLC WorldCat, Universe Digital Library, NewJour, Google Scholar

