

Phytochemical analysis of the selected five plant extracts

Naomi Waiganjo*^{1, 2, 3}, Horace Ochanda², Dorcas Yole^{1, 3}

1. Technical University of Kenya, P.O BOX 52428-00200, Nairobi, Kenya
 2. University of Nairobi, P.O BOX 30197-00100, Nairobi, Kenya
 3. Institute of Primate Research, P.O BOX 24481-00502, Karen, Nairobi, Kenya
- * E-mail of the corresponding author: nwaiganjo@yahoo.com

Abstract

Herbal medicine is still the mainstay of about 75 - 80% of the whole population, and the major part of traditional therapy involves the use of plant extracts and their active constituents. Plants were collected, identified, dried then extracted using hexane, Dichloromethane/methanol and water. Identification assays to test the presence of various chemical constituents were carried out. The five plants were: *Sonchus luxurians*, *Ocimum americanum*, *Bridelia micrantha*, *Croton megalocarpus* and *Aloe secundiflora*. The Phytochemical screening of the compounds present in the plant extracts were; alkaloid, glycosides, Saponins, reducing sugar, Steroid, Flavones and Catecholics. The most common compound in all the plant extracts was Catecholics. Steroids are used in medicine to treat many diseases. The Plant extracts can be possible candidates for drug development.

Keywords: Herbal medicine, Phytochemical compounds, Traditional therapy, Plant extracts

1. Introduction

The use of medicinal plants has gained much attention in the last decade, and among those plants commonly used as medication in folk medicine, various extracts have been the subject of many pharmacological studies (Artuso, 1997). Plants are the richest resource of materials used in modern medicines, food supplements, folk medicines, pharmaceutical intermediates and chemical entities for synthetic drugs (Hammer *et al.*, 1999; Taylor, 2001 and Molgaard *et al.*, 2001). Whereas medicinal plants have produced some very effective treatment for malaria as in case of artemisinin (Frederich *et al.*, 2002; Togola *et al.*, 2008), few attempts have been made to evaluate antischistosomal activity of such natural plants. The importance of plants as sources of natural product bioactive molecules lies not only in their pharmacological or chemotherapeutic effect but also in their role as template molecules for the production of new drugs (Phillipson, 1994, WHO, 2002). The Phytochemical research based on ethno-pharmacological information is generally considered an effective approach in the discovery of new effective agents from plants.

2. Materials and methods

2.1 Plant Extracts

Sonchus luxurians (whole plant), *Ocimum americanum* (whole plant), *Bridelia micrantha* (bark), *Croton megalocarpus*(bark) and *Aloe secundiflora*(whole plant) were collected and placed in plastic bags. The plants were dried at room temperature for 2 months and crushed into tiny particles using Mekon Micro miller Single Phase and passed through a 0.5 mm mesh to standardize the particles. The ground plant material for each plant was divided into equal portions and separately placed in clean large container.

2.1.1 Hexane Extraction

The powdered materials (2kg) from roots, stems, leaves and were first subjected to extraction by soaking in hexane for 72 hours. The process was repeated three times in order to make sure that all non-polar materials are eluted. The contents were filtered using Whatman filter paper (No. 1, medium crystalline). The filtrates were concentrated using Rotary evaporator (RE-100 Bibby, made in Japan) at 70°C. The filtrates from hexane were subjected to drying in fume cupboards at 25°C for 1-2 weeks to remove most of the solvents from the extracts.

2.1.2 Dichloromethane / Methanol Extraction

The residues from n-hexane filtration were soaked for 72 hours in Dichloromethane (DCM) / Methanol in ratio of 1:1 to elute the medium polar materials. The process was repeated 3 times as described for n-hexane extraction.

2.1.3 Aqueous Extraction

The residues from Dichloromethane/ methanol filtrations were further re-soaked using distilled water for 72 hours in order to extract any remaining polar materials. The solutions were then filtered and subjected to freeze drying. The freeze dried materials constitute the aqueous Extract.

The plant extracts obtained were as follows; OAH- *Ocimum americanum* hexane extract, OAD- *Ocimum americanum* Dichloromethane /methanol extract DCM, OAW- *Ocimum americanum* water extract, OAC- *Ocimum americanum* crude, BMH- *Bridelia micrantha* hexane extract, BMD- *Bridelia micrantha* Dcm /Methanol extract, BMW - *Bridelia micrantha* water extract, BMC- *Bridelia micrantha* crude, SLD- *Sonchus luxurians*, Dcm /methanol extract, SLW- *Sonchus luxurians* water extract, SLC- *Sonchus luxurians* crude. CMW- *Croton megalocarpus* water extract, CMC- *Croton megalocarpus* crude, ASW- *Aloe secundiflora* water extract and ASC - *Aloe secundiflora* crude.

2.2 Identification tests to test the presence of various chemical constituents

2.2.1 Alkaloid

0.5 ml of the plant extract solution was evaporated to dryness and 2% hydrochloric acid added in the residue heated on a boiling water bath. After cooling, the mixture was filtered and treated with a few drops of Mayer's reagent. The samples were then observed for the presence of turbidity or yellow precipitation (Siddiqui and Ali, 1997).

2.2.2 Glycoside

To 0.5 ml of the extract solution in glacial acetic acid, few drops of ferric chloride and concentrated sulphuric acid were added. A reddish brown coloration at the junction of two layers and bluish green color in the upper layer was observed (Siddiqui and Ali, 1997).

2.2.3 Terpenoid and Steroid

Four mg of extract was treated with 0.5 ml of acetic anhydride and 0.5 ml of chloroform. Concentrated solution of sulphuric acid was added slowly and red violet color was observed for terpenoid and green bluish color for steroids (Siddiqui and Ali, 1997).

2.2.4 Flavonoid

Four milliliters of extract solution was treated with 1.5 ml of 50% methanol solution. The solution was warmed and metal magnesium was added. To this solution, 5-6 drops of concentrated hydrochloric acid was added and red color was observed for flavonoids and orange color for flavones (Siddiqui and Ali, 1997).

2.2.5 Tannins

One ml of water and 1-2 drops of ferric chloride solution was added to 0.5 ml of extract solution. Blue color was observed for Gallic tannins and green black for Catecholics tannins (Iyengar, 1995).

2.2.6 Reducing Sugar

One ml of water and 5-8 drops of Fehling's solution was added to 0.5 ml extract solution, heated and observed for brick red precipitate (Siddiqui and Ali, 1997; Harbone, 1998).

2.2.7 Saponins

The extract was diluted with 20 ml of distilled water and then agitated in a graduated cylinder for 15 minutes. The formation of 1 cm layer of foam showed presence of Saponins. The frothing was mixed with 3 drops of olive oil and shaken vigorously. Presence of Saponins results in formation of an emulsion (Siddiqui and Ali, 1997).

3. Results

The composition of five plant extracts is shown on the Table 1. The Hexane and Dichloromethane/Methanol extracts of the five plants were semi- solid or sticky composition while water extracts were powder, pellets and semi- solid or sticky form respectively.

The presence of the chemical constituents on five extracts was observed. *Ocimum americanum* plant extracts shown on Table 2 indicated the presence of Steroid, flavones and Catecholics (Tannins). In Table 3 *Bridelia micrantha* plant extract showed; Alkaloid, Catecholics, Glycosides, Saponins and reducing sugar were present. *Sonchus luxurians* plant extracts shown in Table 4 Glycosides, steroid and Catecholics were present. *Croton megalocarpus* plant extracts shown in Table 5 Terpenoid, Flavonoid and reducing sugar were present. *Aloe secundiflora* plant extract shown in Table 6 Catecholics was present.

4. Discussion

Presence of Phytochemical compounds in these extracts illustrates the bioactive compounds in the extracts. Steroid, flavones and Catecholics were common in the *Ocimum americanum* plant extracts while Flavonoid, Gallic and Saponins were absent. Alkaloid, Catecholics, Glycosides, Saponins and reducing sugar were present in all the *Bridelia micrantha* plant extracts. The above compounds able to bind physically to cell walls thereby preventing the adhesion of pathogens to human cell walls hence have antimicrobial activity (Nostro *et al.*, 2000). Glycosides, steroid and Catecholics were present in all *Sonchus luxurians* plant extracts. Terpenoid, Flavonoid and reducing sugar were present in all *Croton megalocarpus* plant extracts.

Catecholics was present in two of *Aloe secundiflora* plant extract however, Terpenoid, Saponins and Gallic were absent. The most common compound in all the plant extracts was Catecholics. Phytochemical studies of the plant preparations are necessary for standardization, which helps in understanding the significance of phytoconstituents in terms of their observed activities.

5. Conclusions

Catecholics is tannin and tannins compounds are widely distributed in many species of plants, where they play a role in protection from predation, and perhaps also as pesticides. Of particular interest is the use of Saponins against the vector of the disease schistosomiasis. Steroids are used in medicine to treat many diseases. The five plant extracts can be possible candidates for drug development.

6. Acknowledgement

We are grateful to: Phytochemistry Laboratory staff at Museums of Kenya for allowing me to carry out plant extractions and also Technical University of Kenya.

References:

- Artuso A, (1997). Drugs of natural origin: Economic and policy aspects of discovery, development and marketing. Pharmaceutical products press, New York.
- Frederich M, Dogne JM, Angenot L and De Mol P, (2002). New trends in anti- malarial agents. *Current Medicine Chemistry*; 9: 1435-1456.
- Hammer, K. A., Carson CF and Riley TV (1999). Antimicrobial activity of essential oils and other plants extracts. *Journal of Applied Microbiology*; 86 (6): 985.
- Harbone, JB, (1998). Phytochemistry methods: A guide to modern Techniques of plant analysis. 3rd edition. Chapman and Hall Ltd, London: 279.
- Iyengar M A, (1995). Study of crude drugs. 8th ed., Manipal Power Press, Manipal, India: 2-5.
- Molgaard P, Nielsen SB, Rasmussen DE, Drummond RB, Makaza N and Andreassen J, (2001) Anthelmintic screening of Zimbabwean plants traditionally used against schistosomiasis. *Journal of Ethnopharmacology*; 74: 257-264.
- Phillipson J.D (1994). Natural products as drugs. *Transactions of the Royal Society of Tropical Medicine and Hygiene*; 88: 17-19.

Siddiqui, A.A and Ali, M (1997). Practical pharmaceutical chemistry.1st Ed., CBS Publishers and Distributors, New Delhi: 126-131.

Taylor, JLS, Rabe T, McGaw LJ, Jäger AK and van Staden J (2001). Towards the scientific validation of traditional medicinal plants. *Plant Growth Regulation*; 34: 23–37.

Togola, A., Astarheim, I M., Theis, A., Diallo D and Paulsen BS, (2008). Ethnopharmacological uses of *Erythrina senegalensis*; A comparison of three areas in Mali, and a link between traditional knowledge and modern biological science. *Journal of Ethnobiology and Ethnomedicine*; 4: 4:6.

World Health Organization (2002) . Traditional Medicine Strategy 2002-2005. World Health Organization , Geneva.

Table 1: The state of the five plant extracts

EXTRACT	STATE		
	Hexane	Dichloromethane/methanol	Water
<i>Ocimum americanum</i>	Semi- solid and sticky	Semi- solid and sticky	powder
<i>Bridelia micrantha</i>	Semi- solid	Semi- solid	pellet
<i>Sonchus luxurians</i>	Semi- solid	Semi-solid	powder
<i>Croton megalocarpus</i>	Semi- solid	Semi-solid	powder
<i>Aloe secundiflora</i>	Semi- solid and sticky	Semi-solid and sticky	Semi- solid and sticky

Table 2: Presence of the active chemical constituents in *Ocimum americanum* plant extracts

	Alkaloid	Glycosides	Terpenoid	Steroid	Flavonoid	Tannins		Reducing sugar	saponins
					Flavonoid	flavones	Gallic catecholic		
OAH	-	-	-	+	-	-	-	-	-
OAD	-	-	-	+	-	+	-	+	Trace
OAW	+	+	+	-	-	+	-	+	-

Table 3: Presence of the active chemical constituents in *Bridelia micrantha* plant extracts

	Alkaloid	Glycosides	Terpenoid	Steroid	Flavonoid		Tannins		Reducing sugar	saponins
					Flavonoid	flavones	Gallic	catecholic		
BMH	+	+	-	+	-	+	-	+	-	-
BMD	+	+	+	-	-	-	-	+	+	+
BMW	+	Trace	+	-	+	-	-	+	+	+

Table 4: Presence of the active chemical constituents in *Sonchus luxurians* plant extracts

	Alkaloid	Glycosides	Terpenoid	Steroid	Flavonoid		Tannins		Reducing sugar	Saponins
					Flavonoid	flavones	Gallic	catecholic		
SLH	-	+	-	+	-	-	-	-	-	-
SLD	-	+	-	+	+	-	-	+	-	Trace
SLW	+	+	+	-	-	-	-	+	-	Trace

Key:
OAH- Ocimum americanum hexane extract, **OAD-** Ocimum americanum, dichloromethane/methanol extract, **OAW-** Ocimum americanum water extract; **BMH-** *Bridelia micrantha* hexane extract, **BMD-** *Bridelia micrantha* dichloromethane/methanol extract, **BMW-** *Bridelia micrantha* water extract; **SLH-** *Sonchus luxurians* hexane extract, **SLD-** *Sonchus luxurians* dichloromethane/methanol extract, **SLW-** *Sonchus luxurians* water extract.

Table 5: Presence of the active chemical constituents in *Croton megalocarpus* plant extracts

	Alkaloid	Glycosides	Terpenoid	Steroid	Flavonoid		Tannins		Reducing sugar	Saponins
					Flavonoid	flavones	Gallic	catecholic		
CMH	-	-	+	-	+	-	-	-	-	-
CMD	-	-	+	-	+	-	-	-	+	-
CMW	+	+	+	-	-	+	-	-	+	+

Table 6: Presence of the active chemical constituents in *Aloe secundiflora* Plant extracts

	Alkaloid	Glycosides	Terpenoid	Steroid	Flavonoid	Flavonoid flavones	Tannins Gallic catecholic	Reducing sugar	Saponins
ASH	-	-	-	-	-	-	-	-	-
ASD	-	-	-	-	-	+	-	+	-
ASW	+	+	-	-	+	-	-	+	traces

Key:

CMH- Croton megalocarpus hexane, **CMD-***Croton megalocarpus* Dichloromethane/ methanol extract, **CMW-***Croton megalocarpus* water extracts; **ASH-***Aloe secundiflora* hexane extract, **ASD-** *Aloe secundiflora* Dichloromethane / methanol, **ASW-***Aloe secundiflora* water extract.

This academic article was published by The International Institute for Science, Technology and Education (IISTE). The IISTE is a pioneer in the Open Access Publishing service based in the U.S. and Europe. The aim of the institute is Accelerating Global Knowledge Sharing.

More information about the publisher can be found in the IISTE's homepage:

<http://www.iiste.org>

CALL FOR JOURNAL PAPERS

The IISTE is currently hosting more than 30 peer-reviewed academic journals and collaborating with academic institutions around the world. There's no deadline for submission. **Prospective authors of IISTE journals can find the submission instruction on the following page:** <http://www.iiste.org/journals/> The IISTE editorial team promises to review and publish all the qualified submissions in a **fast** manner. All the journals articles are available online to the readers all over the world without financial, legal, or technical barriers other than those inseparable from gaining access to the internet itself. Printed version of the journals is also available upon request of readers and authors.

MORE RESOURCES

Book publication information: <http://www.iiste.org/book/>

Recent conferences: <http://www.iiste.org/conference/>

IISTE Knowledge Sharing Partners

EBSCO, Index Copernicus, Ulrich's Periodicals Directory, JournalTOCS, PKP Open Archives Harvester, Bielefeld Academic Search Engine, Elektronische Zeitschriftenbibliothek EZB, Open J-Gate, OCLC WorldCat, Universe Digital Library, NewJour, Google Scholar

