

Effect of *Chlorella vulgaris* Alcoholic Extract to Limit the Growth of (*Staphylococcus aureus*) That Cause Food Poison

Ibtisam Fareed Ali

Market research center and consumer protection/University of Baghdad/Iraq

Abstract

The present study was planned to investigate and conducted to evaluate the antibacterial activities of the green algae crud extract of *Chlorella vulgaris* Staphylococcal food poisoning is a common cause of foodborne illness worldwide. The growth phases of algae were determined and the cultures were harvested at the end of logarithmic phase. The cultures of green algae gave 0.2gm/L. The lipid content of *C. vulgaris* cell was measured and it was 6.3% of dry weight then the fatty acids were analyzed using the high perform liquid chromatography HPLC technique system. The pathogenic bacteria was isolated and identified from different parts of body including swab from skin, expectorant of lung and from sputum of gum, then 5 different concentration of crude extract of alga *C. vulgaris*, 100%, 50%, 25%, 12.5% and 6.3% were tested against pathogenic bacteria *Staphylococcus aureus*. The resistance of bacterial isolates was examined against all algal extract concentrations. The results showed that *Staphylococcus aureus* was sensitive to 6.3% concentration of the crude extract of alga *C. vulgaris* which is the lowest concentration of the algal crude extract. The study concluded that microalgae especially fresh water algae represent a very important source for many active compounds and can be used as antimicrobial agents.

Introduction

Algae have a significant attraction as natural source of bioactive molecules with a broad range of biological activities, including antimicrobial, anticancer, antioxidant, and anti-inflammatory effects (1, 2). Microalgae are microscopic algae, typically found in freshwater and marine systems. They are unicellular species which exist in dividedly, or in chains or groups. depending on the species, their sizes can range from a few micrometers (μm) to a few hundreds of micrometers. Unlike higher plants, microalgae do not have roots, stems and leaves. Microalgae, capable of performing photosynthesis which is very important for life on earth; they produce approximately half of the atmospheric oxygen and use simultaneously the greenhouse gas carbon dioxide to grow photoautotrophically.(3)

Most efforts were devoted to the study of antibiotic resistance in bacteria for several reasons:

- (i) Bacterial infections are responsible for most community-acquired and nosocomial infections
- (ii) The large and expanding number of antibacterial classes offers more diverse range of resistance mechanisms.
- (iii) The ability to move bacterial resistance determinants into standard, well-characterized bacterial strains facilitates more detailed studies of the underlying molecular mechanisms(4)

The growing resistance of some bacterial strains arising from the widespread and essentially unrestricted use of antibiotics in cattle handling, and by domestic consumers use via self-prescription (5) However, a key factor for their eventual economic feasibility, which are able to produce biomass and metabolites to sufficiently high level (6),(7). Some microalgae species such as *Chlorella*, *pirulina* and *Dunaliella* species have been used in several areas in nutraceutical, pharmaceutical, cosmetics, nutrition and functional quality of foods(8). Cyanobacteria like *Anabaena* and *Spirulina* and green microalgae like *Chlorella* and *Scenedesmus* were tested in compliance with the agar well diffusion method for their antibacterial and antifungal agent production on various organisms that incite diseases of humans and plants(9). Fatty acids are widely occurring in natural fats and dietary oils and they play an important role as nutritious substances and metabolites in living organisms (10). Many fatty acids are known to have antibacterial and antifungal properties (11).

Aim and study

Collect, isolate, optimize the growth and Lipid analysis of fresh water sample of *Chlorella* sp. Also study the bacterial susceptibility testing using algal extracts by MHA assay.

Materials and methods

Sampling of algae

Fresh water Algal samples were collected from Tigris River at Baghdad-Iraq, The algal sample was cleaned and necrotic parts were removed. Then the sample was rinsed with sterile water to remove any associated debris. The cleaned culture of the sample was incubated in sterile container. Modified Chu-10 table (1) was used for the algal growth.(12), (13). Isolation of algae Streaking on plate agar techniques were used for algae isolation and purification in this study the samples were incubated under 200 $\mu\text{E}/\text{m}^2/\text{s}$, 16:8 light: dark and $25 \pm 2 \text{ }^\circ\text{C}$, with a PH of 8.2 ± 1 .for 14 days. (14), (15).

Algae identification and cultivation

Chlorella vulgaris Beijerinck Based on the morphological identification the culture was identified under the microscope according to algal classification reference (16). For algae cultivation, 25 ml of isolated culture was added to a flask containing 250 ml of Chu-10 media and incubated under same factors for 14 days, then transported to 1000 ml of media and incubated for 14 days.

Table (1) The components concentration of modified Chu-10 medium and the concentration of each component

Number of stock solution	Chemical formula of each salt	Concentration g/l
1	MgSO ₄ .7H ₂ O	10
2	K ₂ HPO ₄	4
3	NaNO ₃	8
	CaCl ₂	16
4	FeCl ₃	0.32
5	EDTA-Na ₂	4
6	NaCl	30
7	Na ₂ CO ₃	8
8	MnCl ₂ .4H ₂ O	0.02
	(NH ₄) ₆ Mo ₇ O ₂₄ .4H ₂ O	0.028
	ZnSO ₄ .7H ₂ O	0.224
	CuSO ₄ .5H ₂ O	0.08
	COCl ₂ .6H ₂ O	0.004
	H ₃ BO ₃	0.288

Determination of Growth Curve:

The growth curve was determined for the alga *Chlorella vulgaris*. Microalgae concentration was determined daily by optical density (OD) measurements at 540 nm by a UV-Vis spectrophotometer (17).

Harvesting of Algae for Lipid Extraction:

Microalgae culture was harvested at the beginning of the stationary phase., *C. vulgaris* harvested in the tenth day (18) , The cells were harvested by centrifugation at 3000 rpm for 5min., then the algal pellet was collected and dried at 50°C for 48hrs.(19)

Preparation of crude extract

A volume of 2 g from algal has been put on filter paper (Whatman No.1) and extracted in a soxhlet apparatus with 200 ml of solvent (mixture from methanol and hexane 1:1) had been used and the process taken three-four hours. The solvent was evaporated under vacuum at room temperature.(20), (21).

Antibacterial assay

The crude extract was tested for antibacterial activity by agar diffusion assay (22). Wells of 6mm were made in agar plates, and about 0.1 ml of bacteria was spreading on agar surface. . Indicator microbe was spread on muller-hinton agar plates. This was allowed at least 5 minutes to dry before cutting the wells of 6 mm. The crude extract was tested against different concentration of algal extract samples of pathogenic bacteria *Staphylococcus aureus* taken by swab from different parts of human body, Staph 1 swab from skin, staph 2 from expectorant of lung, Staph 3 from sputum of gum , then 5 different concentration of crude extract 100%,50%, 25%, 12.5% and 6.3% and about 50µl of each extract was pipetted into the wells. The control run using the same volume solvent only (methanol and hexane 1:1) The plates were incubated at 37°C for 24hr, then the inhibition zone was measured (19). (23), (24).

Lipid Analysis:

High Performance Liquid Chromatography (HPLC) system, model Shemadzo was used for lipid analysis. HPLC consists from a mobile phase which is polar mixture of solvents such as water: methane (60:40), while the stationary phase is a column stainless steel s, the column is discovery HSC: 18, dimension (25cm × 4.6mm × 5µm), injection flow is 1 ml/min, the absorption at UV 210 nm, and 30 C° temperature.

The results

Isolation and Identification of micro algae *Chlorella* sp. is an important microalgae which involved in the aquatic food chain, and rich in protein content it can be easily grown in chemically defined media and in sewage. (25). *Chlorella* sp that isolated from the collected freshwater was spherical, chloroplast a parietal cup which nearly fills the cell , sometimes without a paranoid ; cells 2-4.5 µm in diameter.(26).

Division :Chlorophyta

Class: Chlorophyceae

Order: Chlorococcales

Family :Chlorococcaceae
 Genus: Chlorella Beyerinck 1890
 Species: Chlorella vulgaris Beyerinck

Growth Curve

The growth density was measured to identify the growth curve and the phases that the alga culture were going through. *C. vulgaris* spent in lag phase four days and seven days in log phase and entered the stationary phase at the day 12, and the decline phase was begun from the day 20. (27)

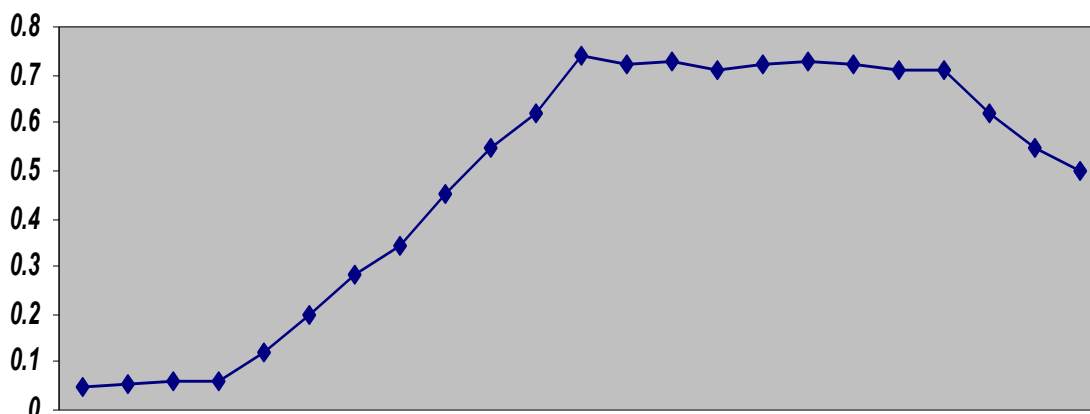


Figure (1) Growth curve of *C. vulgaris*

Lipid and Fatty Acids

The lipid content for *C. vulgaris* was measured and it was 6.3% of dry weight then the fatty acids were analyzed using the HPLC, Stearic acid (18:0) and Oleic acid(18:1). Stearic acid for *C. vulgaris* was 0.81% of total lipid and Oleic acid 0.048 % of the total lipid. (28)

Microorganisms Growth and algal extract activity

Staphylococcus aureus bacteria were employed as test organism which is Gram positive bacteria.

Antibacterial activity of five different concentration extract was assayed using the diffusion method (Bauer et al., 1996). sub cultured and routinely maintained on Muller-hinton agar MHA. Antimicrobial activity was evaluated using the agar diffusion technique in petri dishes.

Different concentration of algal extract was added to the respective wells on the MHA plates. After incubation for 24 hours at 30°C, a clear zone round a wells were evidence of antimicrobial activity. The indication whether test organisms is resistant (No inhibition zone) or sensitive (clear or inhibition zone) to the algal extract was observed. (30).

Effect of algal extraction as Antibacterial activity

The activity of crude extract of algae against gram positive bacteria agreed with most studies used algae extract as source for active compounds against bacterial growth(19), (31). Different concentration of the extract of *C. vulgaris* against the bacteria *Staphylococcus aureus* shows different inhibition zone as is shown in table (2).

The highest effect of extract was recorded in staph1 at the concentration 6.3% the inhibition zone was 9.0 mm, while the lowest inhibition zone was at concentration 50%, 100%. It is observed that the lowest extract of the algal had the maximum activity against *staphylococcus aureus* with an inhibition zone compared to other concentration of algal extract. (32). It is clear that using organic solvent always provides a higher efficiency in extracting compounds for antibacterial activities. (23).

Table (2): The inhibition zone diameter (mm) of different concentration of crude algal extract against samples of *Staphylococcus aureus* bacteria

Bacteria	6.3%	12%	25%	50%	100%
Staph 1	9.0	5.0	3.0	0.0	0.0
Staph 2	6.0	7.0	3.0	0.0	0.0
Staph 3	5.0	6.0	5.0	3.0	2.0

In this study *C. vulgaris* was rich in fatty acid, protein, and pigment concentration the lipid showed that the extracts were rich in fatty acids, in particular saturated fatty acids (SFA), which may indicate their probable role in the antimicrobial activity (33), (34). Fatty acids are potent inhibitors of diverse enzymes and unsaturated fatty acids USFA usually have greater inhibitory activity than (SFA).(35), (36).

Reference

- 1- Tuney I., Cadirci B.H., Unal D., Sukatar A. Antimicrobial activities of the extracts of marine algae from the coast of Urla (Izmir, Turkey). *Turkish Journal of Biology*. 2006; 30: 171-175.
- 2- Patra J.K., Rath S.K., Jena K. Rathod V.K., Thatoi H.N. Evaluating of antioxidant and antimicrobial activity of sea weed (*Sargassum* sp.) extract: A study on inhibition of Glutathione-S transferase activity. *Turkish Journal of Biology* 2008; 32, 119-125.
- 3- Khan S. A, Rashmi Mir. Z. Husain, Prasad S. and Banerjee U. C. 2009. Prospects of biodiesel production from microalgae in India. *Renew. Sustain. Energy Rev.* 13, 2361-2372.
- 4- Ghannoum MA, Rice LB. Antifungal agents: mode of action, mechanisms of resistance, and correlation of these mechanisms with bacterial resistance. *Clinical Microbiology Reviews*. 1999; 12:501-517.
- 5- Guedes AC, Barbosa CR, Amaro HM, Pereira CI, Malcata FX. Microalgal and cyanobacterial cell extracts for use as natural antibacterial additives against food pathogens. *International Journal of Food Science and Technology*. 2011; 46:862-870.
- 6- Kim MK, Park JW, Park CS, Kim SJ, Jeune KH, Chang MU, Acreman J. Enhanced production of *Scenedesmus* spp. (green microalgae) using a new medium containing fermented swine wastewater. *Bioresource Technology*. 2007; 98:2220-2228.
- 7- Sánchez F, Fernández JM., Acien FG, Rueda A, Perez-Parra J, Molina E. Influence of culture conditions on the productivity and lutein content of the new strain *Scenedesmus almeriensis*. *Process Biochemistry*. 2008; 43:398-405.
- 8- Abd El Baky HH and El Baroky G.S. 2013. Health benefits of microalgal bioactive compounds.
- 9- Ramkumar. K. Mandalam. Bernhard Palsson. 1998. Elemental balancing of Biomass and medium composition enhances growth capacity in high-density *Chlorella vulgaris* cultures.
- 10- Cakir, A. (2004). Essential oil and fatty acid composition of the fruits of *Hippophae hamnoides* L. (Sea Buckthorn) and *Myrtus communis* L. from Turkey. *Biochem. System. Ecol.*, 32, 809-816.
- 11- Russel, A.D. (1991). Mechanisms of bacterial resistance to nonantibiotics: food additives and food pharmaceutical preservatives. *J. Appl. Bacteriol.*, 71, 191-201.
- 12- Kassim, T.I.; Al-Saadi, H.; and Salman, N.A. 1999. Production of some phyto- and zooplankton and their use as live food for fish larva. *Iraqi J. Agric. Proc. of 2nd Sci. Confer. Nov. 4(5):188-201*.
- 13- Bischoff, H.W. and Bold, H.C. *Phycological Studies IV. Some soil algae from Enchanted Rock and related algal species* [P]. Univ. Texas Publ, 1963, 6318: 1-95.
- 14- Sinigalliano, C.D.; Winshell, J.; Guerrero, M.A.; Scorzetti, G.; Fell, J.W.; Eaton, R.W.; Brand, L. and Rein, K.S. (2009). Viable cell sorting of dinoflagellates by multiparametric flow cytometry. *Phycologia*. 48:249-257
- 15- Stein, J.R. and Borden, C.A. (1982). *Algae in medicine: Introduction and Bibliography*. In: selected paper in phycology 2nd (eds. Rosowski, J.R. and Parker, B.C.). *Phycol. Soc. of Am. Inc. PP. 788-792*.
- 16- Prescott, G.W. (1982). *Algae of the Western Great Lakes Areas*. Willam, C.; Brown, C.O. Pub. Dubuque. I. Iowa, 16th printing.
- 17- Huang, X.H.; Li, C.L.; Liu, C.W.; Wang, Z.D. and Chen, J.J. (2002). Studies on the N and P nutrient demand of *Nannochloris oculata*. *J. Marine Sciences (China)*. 26(8): 13-17.
- 18- Jawad, A.M. (1982). Interaction between cyanobacteria and other micro-organisms. Ph.D. Thesis. Liverpool University, England.
- 19- Binea, H. K. (2001). The inhibitory effect of the ethanol extract of the local diatom *Nitzschia paleacea* on bacterial stains. M.Sc. thesis, College of Science, University of Baghdad.
- 20- Fajardo, A.R.; Cerdan, L.E.; Medina, A.R.; Fernandez, F.G.A.; Moreno, P.A.G. and Grima, E.M. (2007). Lipid extraction from the microalgae *Pheodactylum tricorutum*. *Eur. J. Lipid Sci. Technol.* 109:120-126
- 21- AOAC (Association of Official Analytical Chemists). (1995). *Official Methods of Analysis*, 16th Edition. AOAC International, Gaithersburg, MD.
- 22- Kellam, S. J. and Walker, J. M. (1989). Antibacterial activity from marine microalgae in laboratory culture. *Br. Phycol. J.*, 24: 191-194.
- 23- Inci Tuney, Bilge Hilal Cadirci, Dilek Unal, Atakan Sukatar, 2006. Antimicrobial activities of the extracts of marine algae from the coast of Urla (Izmir, Turkey). *Turk J Biol*, 30, 171-175.
- 24- Dhanalakshmi M and Angayarkanni, J. (2013). Phytochemistry And Antibacterial Activity Of *Chlorosarcinopsis* Species international journal of scientific and technology research , volume 2, issue 10, October 2013 issn 2277-8616.
- 25- Iwamoto, H., 2004. Industrial production of microalgal cell mass and secondary products major industrial species *Chlorella*. In: Richmond A (ed) *Handbook of microalgal culture*. Blackwell. Oxford, pp 255-263.
- 26- Bellinger, E.G. and Sigeo, D.C. (2010) *Freshwater Algae Identification and use as Bioindicators*. John Wiley and Sons, Ltd pp. 271.
- 27- Al-Hassany, J.S. (2003). Study of Suitable Conditions to the Diatom Algae *Nitzschia paleacea* (Kuetz.) W. Smith. M.Sc. Thesis, College of Science for Women, Univ. Baghdad. 91pp.

- 28- Christie, W.W. and Han, X. *Lipid Analysis - Isolation, Separation, Identification and Lipidomic Analysis* (4th edition), 446 pages (Oily Press, Bridgwater, U.K.) (2010).
- 29- Bauer AW, Kirby WMM, Sherris JC, Turck M (1966). Antibiotic susceptibility testing by a standardized single disk method. *Am. J. Clin. Pathol.* 36: 493-496.
- 30- Vishnu. N. and Sumathi. R. (2014). Isolation of fresh water microalgae *Chlorella* sp and its antimicrobial activity on selected pathogens. *Int.J.Adv. Res.Biol.Sci.* 2014; 1(3): 36-43
- 31- Azza M. Abd El-Aty*1, Amal A. Mohamed2, Farag A. Samhan1. (2014). In vitro antioxidant and antibacterial activities of two fresh water Cyanobacterial species, *Oscillatoria agardhii* and *Anabaena sphaerica*. *Journal of Applied Pharmaceutical Science* Vol. 4 (07), pp. 069-075
- 32- Rajasulochana, P, Dhamotharan, R. Krishnamoorthy, P. Murugesan, S.(2009). Antibacterial Activity of the Extracts of Marine Red and Brown Algae. *Journal of American Science* 2009;5(3)20-25.
- 33- Salem WM, GalalH, Nasr El-deenF. Screening for antibacterial activities in some marine algae from the Red Sea (Hurghada, Egypt). *Afr J Microbiol Res* 2011; 5(15): 2160-2167.
- 34- Plaza M, SantoyoS, Jaime L, García-BlairsyReina G, HerreroM, SeñoránsFJ, et al. Screening for bioactive compounds from algae. *J Pharm Biomed Anal* 2010; 51: 450-455
- 35- Nair MK, Joy J, VasudevanP, Hinckley L, Hoagland TA, VenkitanarayananKS.(2005). Antibacterial effect of caprylic acid and monocaprylin on major bacterial mastitis pathogens. *J Dairy Sci* 2005; 88: 3488-3495.
- 36- HornitzkyMA. *Fatty acids-an alternative control strategy for honeybee diseases: a report for the Rural Industries Research and Development Corporation*. Barton, Australia: Rural Industries Research and Development Corporation; 2003