

# Effect of Traditional Fermentation Time on Total Phenolic Content and In vitro Antioxidant Activity of Kocho: Ethiopian Fermented Food from Enset (*Ensete ventricosum* welw. Chessman)

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## Abstract

*Ensete ventricosum*, locally known as Enset, is the main crop of a sustainable food source that ensures food security in Central, South and South West Ethiopia. It has been used as a food and local medicine for thousands of years. Kocho is a product of matured Enset prepared traditionally by fermenting (for few weeks to some months) a mixture of pulverized pseudostem and corm in pit. The objective of this study was to evaluate the effect of traditional fermentation period on the total phenolic content and antioxidant activity of Kocho. The sample was fermented in the pit for 15, 30, 45, 60, 75, 90, 105 and 120 days. Oven dried Kocho powder was extracted with methanol followed by water. Total phenolic content of the extract was determined by the Folin-Ciocalteu method. Antioxidant activities of the extracts were evaluated with 2, 2-diphenyl-1-picrylhydrazyl (DPPH) scavenging activity and total antioxidant activity using phosphomolybdenum assay. The results revealed that Kocho showed greater total phenolic content and antioxidant activities than unfermented product. Methanol extract of Kocho fermented for 45 days showed the highest amount of total phenolic content ( $9.27 \pm 0.44$  milligram of gallic acid equivalent per gram of dried extract). In comparison of DPPH scavenging and total antioxidant, methanol extracts were also superior to aqueous extracts. Methanol extract of Kocho fermented for 60 days exhibited the strongest DPPH scavenging activity ( $IC_{50} = 0.15 \pm 0.02$  mg/mL) and total antioxidant activity ( $0.43 \pm 0.04$  milligram of ascorbic acid equivalent per gram of dried extract). Thus fermentation offers a tool to further increase the bioactive compounds and antioxidant potential of Kocho.

**Keywords:** antioxidant activity, *Ensete ventricosum*, fermentation, Kocho, total phenolic

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## 1. Introduction

Enset is one of the potential indigenous crops for food, and the Enset cultivation system is economically viable, and is one of the few successful indigenous and sustainable agricultural systems (Yemataw *et al.*, 2018). It is sustainable because it has been providing food for humans for generations from the same plot, and maintains the quality of life of the people (Negash and Anke 2004). It grows in a wide range of environmental conditions. Even though it is grown in many wide areas of the world, the people of the central and southern parts of Ethiopia are the only people that use Enset as a staple and co-staple crop.

Enset (*Ensete ventricosum*) products are one of the main energy sources of foods for about 20 million people in south and south western Ethiopia (Brandt *et al.*, 1997). It is related to and resembles banana plant produced primarily for the large quantity of carbohydrate-rich food. The major food types obtained from Enset are Kocho, Bulla and Amicho. Kocho is the bulk of the fermented food obtained from the mixture of the decorticated (scraped) leaf sheaths and grated corm (Tiruha *et al.*, 2014) (underground stem base). Bulla is obtained by squeezing out the liquid containing starch from scraped leaf sheaths and grated corm and allowing the resultant starch to concentrate into white powder. Amicho is boiled Enset corm pieces, mainly obtained from young Enset plants that are prepared and consumed in a similar manner to other root and tuber crops (Zerihun *et al.*, 2014). Starch is the major component of fermented Enset comprising approximately 99.24% of the dry weight and is in the form of granule size of 46  $\mu$ m (Gebre-Mariam & Schmidt, 1996). The recent study (Zenebe *et al.*, 2018) on fibers, obtained from the scraping of the pseudo stem, as well as the inflorescence stalk remaining after leaves sheath removal, showed the presence of lignin, polysaccharides and extractives.

Besides the macronutrient, study was conducted on micronutrient from ten Enset varieties collected from Sidama zone of southern Ethiopia. Samples of leaf lamina, leaf midrib, pseudostem and corm were taken at the age of 5 to 6 years during the main rainy season. Most Enset fractions were rich sources of major minerals such as phosphorus, potassium, calcium (except corm) and magnesium. Sodium content was very low (Ajebu *et al.*, 2008). According to the study conducted by Foesido *et al.* (2013), Enset products had higher fiber, calcium, potassium, magnesium and manganese content as compared to wheat and corn.

Besides its use as a source of food, Enset is also utilized for animal forage, fiber production, construction material, as ornament, and for its medicinal values (Samuel M. 2014; Melese *et al.*, 2015). Among the Gurage people, South Ethiopia, different varieties of Enset have played a considerable role in traditional medicine which

demonstrated antimicrobial activity against viral, bacteria, fungal and nematodal diseases of humans (Holscher D. and Schneider B. 1998). Some of the Enset varieties are used as medicines by local people for both humans and livestock to cure bone fractures, joint displacement, broken bones, and childbirth problems ie, assisting to discharge the placenta (Balcha, 1990). So far, Biruk *et al.* (2012) studied Enset starch for industrial uses and found a characteristics of high gelatinization property whereas Gebre-Mariam and Schmidt (1996) obtained its use as a binder and disintegrant for tablets.

Though *Enset* is widely used as food source and traditional medicine, scarce information is available on the antioxidant activities of food products of this plant (Forsido *et al.*, 2013). To my knowledge no study has been reported on the effect of fermentation period on total phenolic contents and antioxidant potential of Kocho. Therefore, the objective of the present study was to evaluate the effect of traditional fermentation period on total phenolic content and *in vitro* antioxidant activities of aqueous and methanol extracts of Kocho. In addition, the correlation between total phenolic content and antioxidant activities was also evaluated.

## 2. Methods and Materials

### 2.1. Chemicals

Gallic acid, butylated hydroxyl toluene (BHT), Folin–Ciocalteu reagent, 2,2-diphenyl-1-picrylhydrazyl (DPPH), sodium phosphate, ammonium molybdate, and ascorbic acid were purchased from sigma-aldrich. The other chemicals and solvents used in this experiment were of analytical grade.

### 2.2. Sample Collection and Preparation

Enset local variety known as “Adew” was used for this experiment. The Enset plant was 4 years old grown in 5 Km South of Hawassa town, Sidama zone, South Ethiopia. Woman having traditional experience about the preparation of Kocho was selected to carry out the processing following the traditional farmers’ practices. The scraped and pulverized masses (Figure 1A) were thoroughly mixed with small amount of previously fermented Kocho (as starter of fermentation) and placed in the pit (Figure 1B), lined with Enset leaves and left for fermentation at ambient temperature. A portion of fermented sample (Kocho) was removed from the pit and the liquid was squeezed out of it, resulting into a moist fibrous Kocho (Figure 1C). Sample was collected every 15 days for analysis, up to four months from the date of processing until final fermentation stage and the samples were tested for the total phenolic content and antioxidant activities.

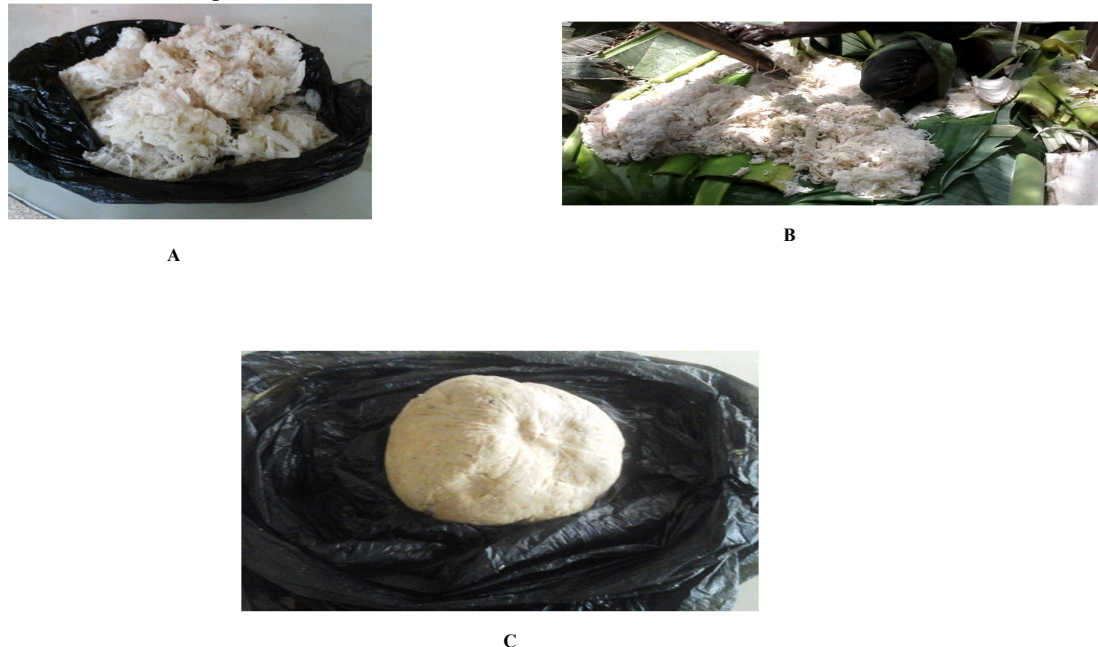


Figure 1: Mixture of decorticated leaf sheaths, the grated and pulverized corm and pseudostem (A), mixture of decorticated leaf sheaths and the grated and pulverized corm and pseudostem in the pit (B), fermented Kocho (C).

### 2.3. Preparation of Kocho Extracts

After oven dried (at 50°C) sample was ground to fine powder using electric grinder (FM100 model, China). The aqueous and methanol extracts of all were prepared by dissolving 10 g of the samples fine powder separately in 100 mL of each solvent. The contents were kept in orbital shaker for 6 h at room temperature. Thereafter, each extract was filtered using Whatman no.1 filter paper and evaporated to dryness under vacuum at 40°C by using a rotary evaporator (Buchi, 3000 series, Switzerland). The extraction was done in triplicate for each solvent and

the resulting extracts were stored in a sealed plastic container at 4°C until further investigation.

#### 2.4. Total phenolic Contents (TPC)

Total phenolic content was estimated by Folin-Ciocalteu method as described in Shan *et al.* (2005) with slight modification using gallic acid as standard. To 0.5mL of the extract (5mg/mL), 2 mL Folin-Ciocalteu reagent (diluted ten times) was added and the mixture was left for 5 min and then 1mL (75% w/w) of sodium carbonate was added. The absorbance of the resulting blue color was measured at 765 nm with a UV- visible spectrophotometer (JENWAY, 96500, UK) after incubation for 90 min at room temperature. The total phenolic content was estimated from gallic acid (1-100 µg/mL) calibration curve ( $y = 0.02x + 0.09$ ,  $R^2 = 0.99$ ) and results were expressed as milligram gallic acid equivalent/gram of dry extract (mgGAE/g).

#### 2.5. Determination of Antioxidant Activity

##### 2.5.1. DPPH method

The capacity of kocho samples to scavenge free radical was measured with the stable radical of DPPH using method as described by Iqbal *et al.* (2005), with minor modification. Two mL of freshly prepared DPPH solution (0.06%, w/v) in methanol was mixed with different concentrations ((0.1up to 5 mg/mL) of 1 mL of extract and reference standards (BHT) dissolved in methanol. The mixtures were vortexed and kept in a dark room for 30 min at room temperature. The absorbance of the resulting solution was then taken at 520 nm. Methanol was used as blank. The ability to scavenge the DPPH radical was calculated using the following equation:

$$DPPH\ scavenged\ (\%) = \frac{(Ac - As)}{Ac} \times 100$$

Where  $Ac$  is the absorbance of the control and  $As$  is the absorbance in presence of the sample of the extracts. The antioxidant activity of the extract was expressed as  $IC_{50}$ . The  $IC_{50}$  value was defined as the concentration of extracts that scavenges the DPPH radical by 50%. The value (in mg/mL) was extrapolated by plotting percentage inhibition of sample against the corresponding sample concentration.

##### 2.5.2. Determination total antioxidant activity by phosphomolybdenum method

The total antioxidant activity of the crude extracts was evaluated by the phosphomolybdenum method (Prieto *et al.*, 1999) with slight modification. The method is based on the reduction of Mo (VI) to Mo (V) by the antioxidant compounds or crude extract and subsequent formation of green Mo (V) complexes with a maximal absorption at 695 nm at acidic medium (Mohamed *et al.*, 2011). Plant extract (0.3 mL) was mixed with 3 mL of reagent solution (0.6 M sulphuric acid, 28 mM sodium phosphate and 4 mM ammonium molybdate). The samples were incubated at 95 °C for 90 min, cooled to room temperature and absorbance was measured at 695 nm and methanol (3 mL) was used as blank. The total antioxidant activity was expressed as milligram butylated hydroxytoluene equivalent/gram of dried extract (mgAAE/g) based on the calibration curve;  $y = 0.301x + 0.002$ ,  $R^2 = 0.99$ .

#### 2.6. Statistical Analysis

The data were subjected to analysis of variance (ANOVA) and Duncan's multiple range tests were used for mean separation at  $p < 0.05$ . Linear regression analysis was used to calculate  $IC_{50}$  value. Pearson correlations between antioxidant activities, total phenolic contents were considered at  $p < 0.05$ .

### 3. Result and Discussions

#### 3.1. Total Phenolic Content of Kocho

The total phenolic content of Kocho was expressed as milligram of gallic acid equivalent per gram of dried extract (mgGAE/g) as shown in Figure 1. In all samples methanol extracts had higher total phenolic content than that of aqueous extracts. Phenolic content of methanol Kocho extracts analyzed in the study ranged from  $1.32 \pm 0.02$  mgGAE/100 g for unfermented sample to  $9.27 \pm 0.9$  mgGAE/100 g for Kocho fermented for 45 day. Among the aqueous extracts, Kocho fermented for 60 days had the highest ( $p < 0.05$ ) phenolic content, while unfermented Kocho sample had the lowest phenolic content. Similarly the total phenolic content of methanol extracts of fermented Kocho increased with increasing the period of fermentation. The total phenolic content of Kocho fermented for 45 days was the highest ( $p < 0.05$ ). Whereas, the unfermented sample had the lowest total phenolic content ( $p < 0.5$ ). The result increased up to 45 days. After 60 days of fermentation the total phenolic content decreased gradually but no significance level ( $p > 0.05$ ) was observed. But the total phenolic content all fermented samples was significantly higher than that of unfermented sample. These results were consistent with findings on fermented Okra seeds (Adetuyi and Ibrahim 2014), Cabbage (Sun *et al.*, 2009), and Jaruk tigarun (Nazarni *et al.*, 2016), traditional fermented food from South Borneo Indonesia, where fermentation caused increase in the total phenolic contents. In natural form, phenolic compounds are combined or bound with sugar

which reduces their availability to organism. Many studies indicated that deglycosilation of phenolic compounds could be performed through microbial fermentation due to glycosyl hydrolase family activities (Huynh *et al.*, 2014). These enzymes are well known for their deglycosylation capability by hydrolyzing the different glycosidic bonds existing between sugars and phenolic compounds, causing a significant increase in the amount of different free phenolic compounds (Cho *et al.*, 2009; Lee *et al.*, 2013; Di Gioia *et al.*, 2014 ).

Table 1: Total phenolic content (mgGAE/g of dried extract) of methanol and water extracts of Kocho.

Fermentation period	Water extract	Methanol extract
Fresh	0.32 ± 0.08 <sup>a</sup>	1.32 ± 0.02 <sup>a</sup>
15 days	2.75 ± 0.05 <sup>b</sup>	5.57 ± 0.14 <sup>bc</sup>
30 days	4.86 ± 0.10 <sup>bc</sup>	5.88 ± 0.20 <sup>bc</sup>
45 days	4.98 ± 0.07 <sup>bc</sup>	9.27 ± 0.44 <sup>d</sup>
60 days	5.34 ± 0.88 <sup>c</sup>	8.00 ± 0.56 <sup>c</sup>
75 days	5.03 ± 0.11 <sup>c</sup>	5.93 ± 0.12 <sup>bc</sup>
90 days	4.98 ± 0.06 <sup>bc</sup>	5.82 ± 0.22 <sup>bc</sup>
105 days	4.78 ± 0.13 <sup>bc</sup>	5.60 ± 0.58 <sup>bc</sup>
120 days	4.41 ± 0.03 <sup>bc</sup>	5.53 ± 0.70 <sup>bc</sup>

Values are expressed as mean ± SD (n = 3) from triplicate experiments .Values within a column with different letters are significantly different at p < 0.05.

### 3.2. DPPH Free Radical Scavenging Activity

Scavenging the stable DPPH radical model is a widely used method to evaluate *in vitro* antioxidant activity. DPPH is a stable free radical with characteristic absorption at 520 nm and antioxidants react with DPPH radical and convert it to diamagnetic 2, 2-diphenyl-1-picrylhydrazine molecule. The degree of discoloration indicates the scavenging potential of the antioxidant extract, which is due to the hydrogen donating ability (Von Gadow *et al.*, 1997). Figures 2 and 3 show dose dependence curve for DPPH radical scavenging activity of methanol and water extracts of fermented Kocho which quantified by its absorbance reduction at a wavelength of 520 nm. As concentration of sample increased, the percent of DPPH scavenging increased. Generally, in response to fermentation treatment, there was an increase in relative percentage of radical scavenging activity in all fermentation periods, compared to the control. The result obtained was in accordance with previous research done on red cabbages by Hunaefi *et al.* (2013), who reported an increase in the percentage of radical scavenging activity in fermented cabbage compared with unfermented cabbage as a control. The increase may be due to the increase in acidic value during fermentation that is liberating bound flavonoid components and making it more bio available (Ashish *et al.*, 2014). Fermentation time did significantly increase the DPPH radical-scavenging activities for all extracts, as compared to unfermented pulverized corm. Contrary to the present result, fermented Kocho sample collected from South Western Ethiopia, showed weaker antioxidant activity than the unfermented pulverized corm of Enset (Forsido *et al.*, 2013).

Table 2 shows IC<sub>50</sub> value, which is the inhibition concentration of sample extract required to decrease initial DPPH activity by 50%. DPPH scavenging activities were also reflected in the IC<sub>50</sub> values where the value for methanol extract of Kocho fermented for 60 days (IC<sub>50</sub> = 0.15 ± 0.02 mg/mL) was significantly (p < 0.05) lower (stronger antioxidant activity) than that of all fermented Kocho samples. The unfermented fresh sample showed the weakest (IC<sub>50</sub> > 5mg/mL) DPPH scavenging activity. Similarly, the water extract of Kocho fermented for 60 days showed the strongest DPPH scavenging (IC<sub>50</sub> = 0.81 ± 0.47 mg/mL) activity. Further increasing the fermentation period (above 60 fermentation days) decreased the DPPH scavenging potential of Kocho extract. BHT as a reference showed stronger DPPH scavenging activity (p < 0.05) than that of all fermented Kocho extracts. Different studies reported (Moktan *et al.*, 2008; Ademiluyi and Obboh, 2011) that prolonged fermentation decreased the available phenolic compounds because of the microorganisms grow using these compounds as substrates (Ehsan *et al.*, 2010). Therefore the optimal fermentation period is important to get the maximum amount of phenolic compounds during fermentation of food products.

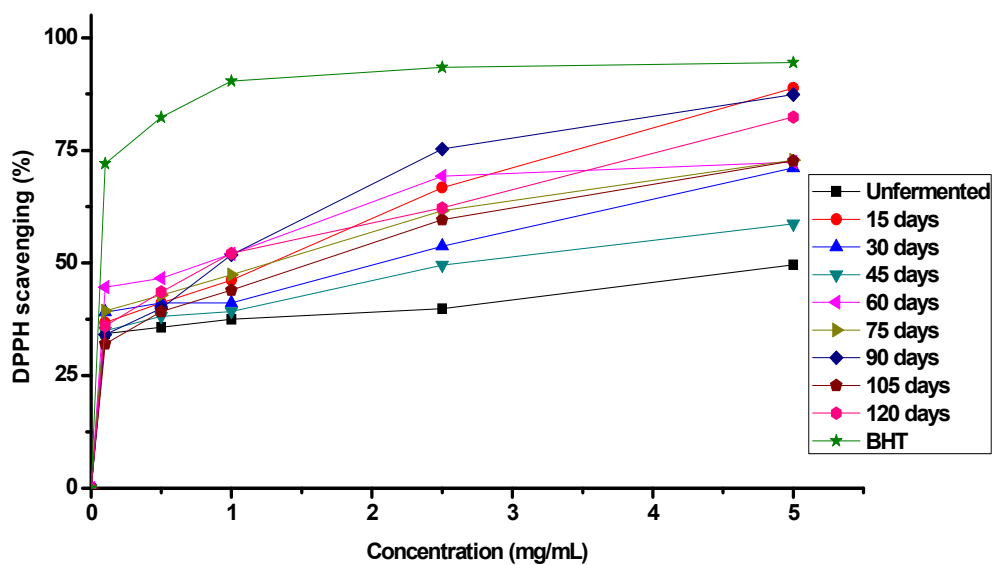


Figure 2: DPPH radical scavenging activity aqueous extracts of Kocho, fermented for different period of time and control (BHT). Values are average of triplicate measurements (mean  $\pm$  SD).

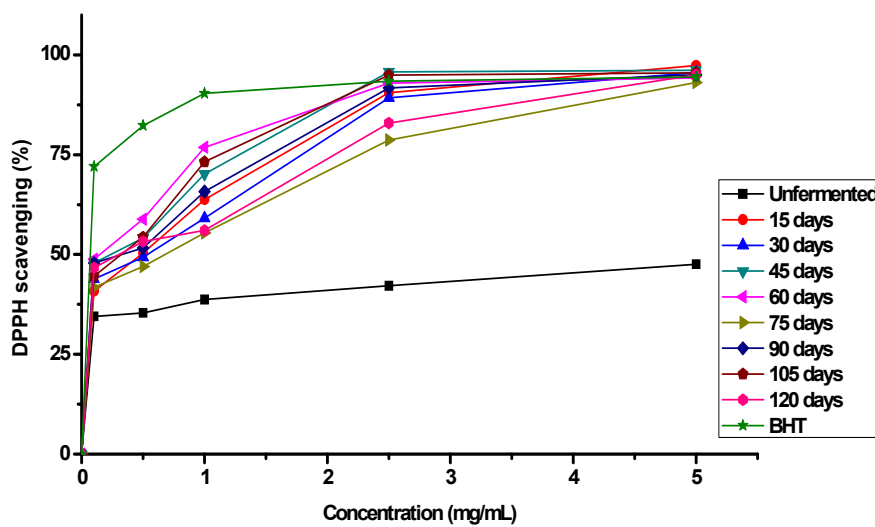


Figure 3: DPPH radical scavenging activity methanol extracts of Kocho, fermented for different period of time and control (BHT). Values are average of triplicate measurements (mean  $\pm$  SD).



Table 2: IC<sub>50</sub> (mg/mL) of DPPH scavenging activity of methanol and water extracts Kocho fermented for different period of time.

Fermentation period (days)	Aqueous extract	Methanol extract
Unfermented	> 5	> 5
15	1.29 ± 0.09 <sup>bc</sup>	0.47 ± 0.02 <sup>abc</sup>
30	2.10 ± 0.39 <sup>d</sup>	0.52 ± 0.04 <sup>bc</sup>
45	2.66 ± 0.42 <sup>e</sup>	0.25 ± 0.19 <sup>ab</sup>
60	0.81 ± 0.47 <sup>b</sup>	0.15 ± 0.02 <sup>a</sup>
75	1.16 ± 0.03 <sup>bc</sup>	0.67 ± 0.23 <sup>c</sup>
90	0.94 ± 0.10 <sup>bc</sup>	0.39 ± 0.23 <sup>abc</sup>
105	1.55 ± 0.18 <sup>c</sup>	0.33 ± 0.04 <sup>abc</sup>
120	1.03 ± 0.39 <sup>c</sup>	0.27 ± 0.08 <sup>ab</sup>
IC <sub>50</sub> (BHT) =	0.08 ± 0.02 mg/mL	

Values are expressed as mean ± SD (n = 3) from triplicate experiments.. Values within a column with different letters are significantly different at p < 0.05.

### 3.3. Total antioxidant activity using phosphomolybdenum assay

Similar to DPPH scavenging, the methanol extract showed stronger total antioxidant activity than that of aqueous extracts (Table 3). Among the methanol extracts, Kocho fermented for 60 days showed the strongest total antioxidant activity with the value of 0.43 ± 0.04 mgAAE/g. whereas unfermented sample showed the significantly the weakest (p < 0.05) total antioxidant. No significant difference (p > 0.05) was found between the total antioxidant activities of Kocho fermented 15 up to 30 days and also these values have no significant difference with Kocho fermented 75 up to 90 days. But these values showed significantly (p < 0.05) stronger total antioxidant activity than Kocho fermented between 105 and 120 days. The total antioxidant activities of all fermented Kocho samples were significantly stronger (p < 0.05) than that of unfermented sample. Similarly, the water extract of Kocho fermented for 60 days showed the strongest total antioxidant activity (p < 0.05).

Table 3: Total antioxidant activity (mgAAE/g of dried extract) of methanol and water extracts of Kocho fermented for different period of time.

Fermentation period	Aqueous extract	Methanol extract
Unfermented	0.03 ± 0.01 <sup>a</sup>	0.17 ± 0.01 <sup>a</sup>
15 days	0.13 ± 0.01 <sup>b</sup>	0.28 ± 0.09 <sup>bc</sup>
30 days	0.18 ± 0.02 <sup>bc</sup>	0.33 ± 0.04 <sup>cd</sup>
45 days	0.18 ± 0.07 <sup>bc</sup>	0.38 ± 0.04 <sup>de</sup>
60 days	0.28 ± 0.06 <sup>d</sup>	0.43 ± 0.04 <sup>e</sup>
75 days	0.23 ± 0.08 <sup>cd</sup>	0.38 ± 0.02 <sup>de</sup>
90 days	0.20 ± 0.04 <sup>bcd</sup>	0.37 ± 0.03 <sup>de</sup>
105 days	0.17 ± 0.03 <sup>bc</sup>	0.23 ± 0.07 <sup>ab</sup>
120 days	0.17 ± 0.04 <sup>bc</sup>	0.23 ± 0.08 <sup>ab</sup>

Values are expressed as mean ± SD (n = 3) from triplicate experiments. Values within a column with different letters are significantly different at p < 0.05.

### 3.4. Correlation analysis

It is well known that the antioxidant activity of a plant extracts are largely depending on both the concentration and the nature of phenolic compounds present in the extract (Catherine *et al.*, 1996; Weng and Huang. 2014). In this study, the relationship between total phenolic content and antioxidant activity Kocho fermented for different period of time extracted by methanol and water solvents was evaluated (Table 4). The results showed good linear correlation in the cases of DPPH scavenging activity (R<sup>2</sup> = 0.53, p > 0.05) and total antioxidant activity (R<sup>2</sup> = 0.52, p > 0.05) with total phenolic content of methanol extracts of Kocho fermented up to 120 days. In case of aqueous extracts the total phenolic was weakly correlated ((R<sup>2</sup> = 0.48, p > 0.05) with total antioxidant activity but moderately correlated (R<sup>2</sup> = 0.63, p > 0.05) with DPPH scavenging activity of all Kocho samples.

Table 4: Correlation of antioxidant activities with TPC of Kocho samples during fermentation processes.

Extract	Correlation factor	Correlation coefficient (R <sup>2</sup> )
Water	TPC	+0.53
	Vs. DPPH Scavenging (%)	
	TPC	+0.52
Methanol	Vs. mg BHTE/100g	
	TPC	+0.63
	Vs. DPPH Scavenging (%)	
	TPC	+0.48
	Vs mg BHTE/100g	

#### 4. Conclusions

This study demonstrated that fermentation caused a marked increase in total phenolic content which then enhanced DPPH radical-scavenging ability and total antioxidant activity of the Kocho extracts. Fermentation time affected the antioxidant activities of Kocho and fermentation for 60 days seemed to be applicable as exemplified by the least IC<sub>50</sub> value of DPPH radical-scavenging ability and highest value of total antioxidant power. The methanol extract showed the strongest DPPH scavenging and total antioxidant activity. Whereas, in all extraction periods aqueous extracts exhibited weaker DPPH scavenging and total antioxidant activities.

In conclusion, extending of fermentation time had resulted differences in total phenolic contents and antioxidant activity of Kocho. Therefore, fermentation played a significant role in Kocho preparation by enrichment of the product through development and release of phenolic compounds due to the action of cell wall-degrading enzymes produced through fermentation of bioactive components and enhancing antioxidant capacity. Generally, the selection of appropriate Enset varieties, improved processing methods, and recommended fermentation time are good practice for the processing of tradition food products, such as Kocho an Ethiopian traditional fermented food. This work was carried out with the analysis of crude Kocho extract. However, further studies are needed to evaluate the *in vivo* experiments of various solvent extracts and individual bioactive compounds in various animal models.

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