

A Study on the Effect of Ethidium Bromide on Virulence Factors (Protease and Biofilm Formation) by *Klebsiella Pneumoniae* Isolated from Different Clinical Sources

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Abstract

About 145 isolate of *Klebsiella* species were isolated from different clinical sources for 6 months ago from (1-9-2014) to (1-2-2015). These isolates were identified morphologically and biochemically and Api 20 kit, thus there were only 22 isolates identified as *Klebsiella pneumoniae* (15.1%). Protease activity was tested by measuring the diameter of cleared zones around wells containing bacterial cells harvested from 18 h of incubation at 37°C on skim milk agar medium or by (wells method). The results showed that k4 is the higher isolate of protease production (18 mm in diameter) while k2 showed a lowest protease activity of (12 mm in diameter). Biofilm formation for 22 *Klebsiella* isolates was tested on Congo-red agar medium and the results showed that all the isolates have (100%) activity by forming dark colonies on Congo-red medium. In our study, first attempt was made on the effect of ethidium bromide on the virulence factors of pathogenic bacteria (*Klebsiella pneumoniae*) at concentrations (10^{-1} to 10^{-6}) that have an effect at which the 22 isolates of (*K.p*) lose the protease activity and biofilm formation at these concentrations while little and normal activities were observed at concentrations (10^{-7} to 10^{-10}) of ethidium bromide on the 22 studied isolates of *K. pneumoniae*. Also, the results of Agarose-gel electrophoresis of both (normal case) *Klebsiella pneumoniae* k4 and cured isolates showed the presence of chromosomal and plasmid DNA bands in the normal case while only chromosomal DNA bands occur with the *Klebsiella* isolates treated with Ethidium-Bromide at concentrations of (10^{-2} to 10^{-4}).

Keywords: Ethidium-Bromide – *Klebsiella* – Protease – Biofilm .

Introduction

The genus *Klebsiella pneumoniae* is a gram negative, non fermentative bacillus; has emerged as a major pathogen among nosocomial infections which is related to possessing anti phagocytosis capsule (Levinon;2004;Umhe et al;2006). *Klebsiella* species are routinely found in the Human nose, mouth and gastrointestinal tract as normal flora; however they can also behave as opportunistic human pathogens (Einstein,2000;Abbot,2003;Vincent,2004). *Klebsiella* infections can involve pneumonia, urinary tract infection, septicemia, diarrhea and soft tissues infections (Kaye, et al ;2000). There are many factors contribute to pathogenicity of *Klebsiella pneumoniae* especially protease production and biofilm formation (in addition capsule antigen, ability of adhesion, siderophores and others). A protease (or proteinase) is any enzyme that performs proteolysis, that is, begins protein catabolism by hydrolysis of the peptide bonds that link amino acids together in a poly peptide chain. Proteases have evolved multiple times, and different classes of protease can perform the same reaction by completely different catalytic mechanisms. Proteases can be found in animals, plants, bacteria, archaea, and viruses. They can attack the immunoglobulins and immune cells (Nehad, T.A, 2003; Senior, 1999). Another important virulence factor contributing to the *K.pneumoniae* pathogenesis in clinical settings is the biofilm mode of growth involved in chronic as well as in acute infections (Schaber et al ;2007). Biofilm are formed from individual free-floating (planktonic) cells and are defined as an exopolysaccharide-surrounded bacterial complex on the biotic or a biotic surfaces (Hoiby et al 2001). Bacterial cells in the biofilm often display a variety of phenotypic differences from those in the planktonic culture. These include some phenotypic changes such as motility, production of extracellular polysaccharide and increased resistance to antibiotic and host defence system (Ceri et al, 1999; Tarkkanen et al , 1997; Rollin and Joseph, 2000). This study was viewed on the effect of Ethidium-Bromide on these two virulence factors (protease and biofilm formation) in which it is (Et-br) an intercalating agent commonly used as a fluorescent agent (nucleic acid stain), in molecular biology laboratories for techniques such as agarose gel electrophoresis, when exposed to ultraviolet, it will fluoresce with an orange color, intensifying almost 20 fold after binding to DNA under the name (ethidium). Ethidium-bromide may be a mutagen, a carcinogen, or a teratogen, although this depends on the organism exposed and the circumstances of exposure. (Sambrook et al 1989).

Materials and Method

Isolation and Identification of Bacterial Isolates

The present study included (145) samples from different clinical sites during 6 months included: 98 urine, 14 stool, 7 blood, 5 ear swap, 13 wound, 2 burn swap, 4 throat swap, 2 skin swap. These clinical samples were collected from the main three hospitals in Baghdad – Iraq (Al Kindy, Al Kadhamiyah, Al-Yarmoq) in addition

to general health Laboratories . Bacterial isolates were identified to the level of subspecies using the Traditional biochemical and morphological test described by (Baron et al ,1999) and then confirmed using rapid identification systems (Api 20 E) as recommended by the manufacture(Biomeriex _France)

Protease Activity

Protease activity were measured before and after ethidium bromide by measuring the diameter of lysis area after growing of (18-36 h) of incubation at 37^oc on skim milk agar media for all isolates,using (wells method) (Barron and Fine gold ,1994).

Biofilm Formation

The bacterial activity for biofilm formation were measured by culturing on congo –agar media,in which cells of 18 h to 36h of incubation at 37 c , dark ends of growing determined the biofilm production (Todar,2007).This activity tested before and after addition of ethidium bromide to nutrient agar medium at concentrations(10⁻¹ to 10⁻¹⁰) (Baron and Fingold1999)

DNA –Analysis

Chromosomal)and plasmid DNA was extracted from cultured cells using the alkaline –SDS method described by Kirby et al ,1995.

Results and Discussion

Isolation and Identification of Bacterial Isolates

Results of morphological and biochemical characterization tests revealed that a total of 22 isolates were belonged to *Klebsiella pneumoniae*

Protease production and Biofilm Formation by *Klebsiella* isolates

The production of extracellular protease from a number of pathogenic bacteria represent one of the most important virulence factors have a wide spread of interesting field for studying , the results of this study showed that *Klebsiella pneumoniae* 4was the highest activity (18 mm indiameter) by measuring the diameter of lysis area on skim milk agar media (or by wells method) while the k2 was the lowest protease activity (12mm in diameter) as shown in table (1) and figure (1).

These findings are in agreement with results obtained by Nehad (2003) who found similar and higher results of protease from *Proteus mirabilis* ,in some genetic studies of protease production from different bacteria found that it is chromosomally determined (Pons *et al* ,2004;Wassif *et al*1995).there are many factors affect protease production such as the time of incubation ,the presence of inhancers ,metals .in addition ,protease can cleavage the immunoglobulins and many immune cells .(Nehad,2003)

ACTIVITY ISOLATE NO.	PROTEASE ACTIVITY	BIOFILM
	DIAMETER OF LYSIS AREA IN (mm)	FORMATION BY(CRA)
K1	14	100
K2	12	100
K3	16	100
K4	18	100
K5	14	100
K6	14	100
K7	14	100
K8	14	100
K9	14	100
K10	16	100
K11	14	100
K12	14	100
K13	17	100
K14	14	100
K15	15	100
K16	14	100
K17	16	100
K18	12	100
K19	14	100
K20	13	100
K21	11	100
K22		
K(Control)	14	100

Table (1) showed both the protease activity of 6 *K.pneumoniae* isolates detected by measuring the

diameter of lysis area after 18h of incubation at 37°C on skim milk agar media and biofilm formation after incubation at 37°C on congo agar media.

Incontrast,our results showed that allthe 22 *Klebsiella* isolates showed complete biofilm formation(100%)as in table (1) by forming deep darkng colonies on congo-red agar medium figure(3).These results are agreement with that of Podschan *et al* (2000)which they can isolate ahigh frequency of Germany clinical isolates that have similar virulence factors activity like capsule production ,siderophores ,resistance to serum and biofilm formation(Podschan et al,2000).Also a high Beta-TEM-59-lactamaseresistance and other s including biofilm formation by *K. oxytoca* were isolated at (26%)from different European clinical sources.(Bermuds *et al*,1999)

The Effect of Ethidium –Bromide on both Protease and Biofilm Formation by *Klebsiella pneumoniae* isolates

The results were shown in table (2) represent that the mutagenic agent (ethidium-bromide) have an effect at concentrations (stock solution,to 10⁻⁶)at which all 22 isolates loss the protease activity and biofilm formation as in figure (3),(4) while little and normal activities were observed at concentrations(10⁻⁷ to 10⁻¹⁰) as in table (2).These findings are in agreement with the results obtained by many researchers(;Kafaf,2000) who found many of antibiotic resistance were plasmid determined and affected after curing experiments .Incontrast ,many results revealed the chromosomally determined abilities such as Actinorhodin-like substance production by *Streptomyces* IQ45(Nehad,1998) Jones *et al* (1990) isolate many *Proteus mirabilis* mutants affect their virulence factors .

BACTERIAL ACTIVITY ETHIDIUM BROMIDE SOLUTION	PROTEASE ACTIVITY	BIOFILM FORMATION
Stock solution	No-activity	No-activity
10 ⁻¹	No-activity	No-activity
10 ⁻²	No-activity	No-activity
10 ⁻³	No-activity	No-activity
10 ⁻⁴	No-activity	No-activity
10 ⁻⁵	No-activity	No-activity
10 ⁻⁶	No-activity	No-activity
10 ⁻⁷	Little (10mm)	Normal darking (100%)
10 ⁻⁸	Normal (12mm)	Normal darking (100%)
10 ⁻⁹	Normal(14mm)	Normal darking (100%)
10 ⁻¹⁰	Normal(14mm)	Normal darking (100%)

Table (2) show the effect of Ethedium bromide concentration on both protease and biofilm formation by *K.pneumoniae* isolates .

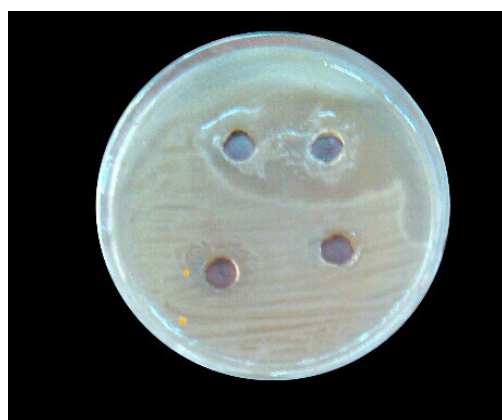


Figure (1) showed the protease activity (12 mm) in diameter of *K. pneumonia* (K4) measured by wells method after 18 h of incubation at 37°C

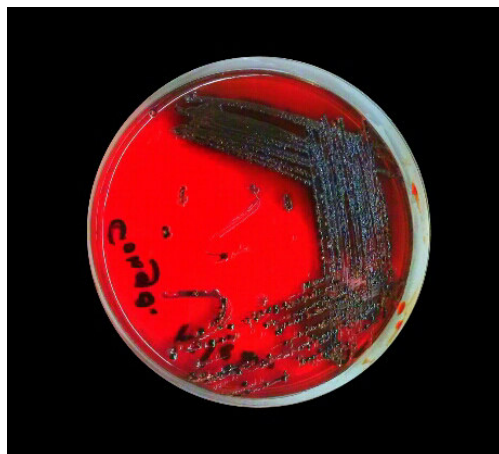


Figure (2) showed the biofilm activity of *K.pneumoniae* (K6) measured by congo-red agar method (CRA)



Figure (3) showed the absence of protease activity after growing on medium containing Et-Br concentrations (0 to 10^{-7})

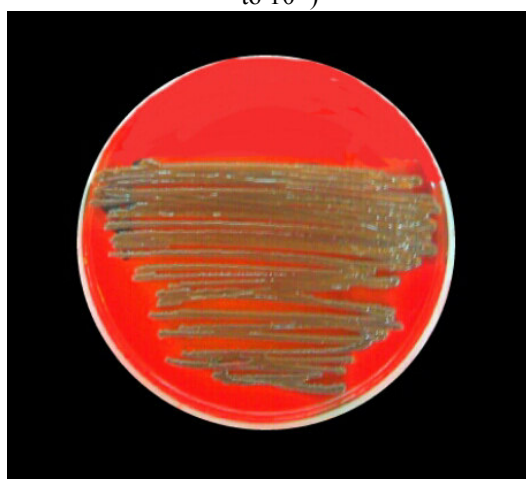
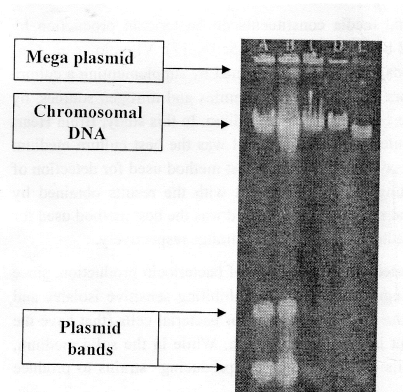


Figure (4) showed the absence of biofilm activity (0%) after growing on medium containing Et-Br concentration (10^{-4}).

DNA –Analysis

The results of DNA –analysis of both normal and cured isolates showed the presence of chromosomal and plasmids bands in (normal case) while only chromosomal bands observed in *Klebsiella* isolates treated with Ethidium –Bromide at concentrations (10 and 10) as in figure (5), the absence of plasmids was correlated with the absence of protease production and biofilm formation by *Klebsiella* isolates as mentioned below which explain the fact of their genetics ,they may be ,in most probable plasmids determined neither than

chromosome. Finally, our results showed that Ethidium-Bromide have an effect on the virulence factors especially protease and biofilm formation due to the mutagenic effect on the specific genes of their production by *Klebsiella* isolates under this study.



Figure(5) : Agarose gel electrophoresis of chromosomal and plasmid DNA isolated from *K. pneumoniae* K for (line A,B) normal and curing isolates treated with Eth. Bromide at concentrations (10^{-2} and 10^{-4}) panel (C,D,E)

References

- Levinson, W. (2004). *Klebsiella*: in medical microbiology and immunology examination and broad review 8th ed the Mc C ROW-Hill companies Appleton –USA. Invasive *Klebsiella pneumoniae* in North America, Clin. Infect. Dis. 45:25-28.
- Umhe, O., Berkowitz, L. and Case, C. (2006). Infectious disease, *Klebsiella* infection. J. e. Medicine: 27(1).
- Enstein, B. (2000). Enterobacteriaceae in Mandel, Douglas and Bennetts. principles and practice of infectious disease 5th ed. New York, NY. Churchill Livingstone, 2:294-310.
- Abbot, S. (2003). *Klebsiella enterobacter Citrobacter, Serratia Pleisiomonas* and other Enterobacteriaceae. In clinical Microbiology. edited by Mummy 8th ed. ASM press.
- Vincent, W. (2004). Infectious caused by member of the genus *Klebsiella*. Infectious Disease. 11(5):28-33.
- Kay, K.; Fraimow, H. and Arbutyn, E. (2000). Pathogens resistance to antimicrobial agents. epidemiology, molecular mechanisms and clinical management. Infect. Dis. Clin. North. Am. 14(2):319-393.
- Nehad, A. T. (2003). A study on two enzymes (IgA Protease and Urease) isolated from *Proteus mirabilis* caused urinary tract infections PhD thesis, college of science, Al-Nahrain university. Iraq.
- Senior, B. W. (1999). A survey of IgA protease production among clinical isolates of the proteases. J. Med. Microbiol. 25:27-35.
- Schaber, A.; Jeffrey, T. S.; Oliver, J.; Hastert, C.; Griswold, A.; Manfred, A.; Abdul, Hamood and Kendra, R. (2007). *Pseudomonas aeruginosa* forms biofilms in acute infections independent of cell-to cell signaling. Infection and Immunity. pp3715-3721.
- Hoiby, N.; Krough, J. H.; Moser, N.; Song, Z.; Ciofu, O. and Khaarazmi, A. (2001). *Pseudomonas aeruginosa* and the in vitro and in vivo biofilm mode of growth. Microbes. Infect. 3:23-25.
- Ceri, H. M.; Olsmo, C.; Stremic, R. R.; Read, D.; Morck and Buret, A. (1999). The Calgary Biofilm Device: new technology for rapid determination of antibiotic susceptibilities of bacterial biofilms. J. Clin. Microbiol. 37:1771-1776.
- Tarrkannan, A. M.; Virkola, R. and Clegg, S. (1997). Binding of the type 3 fimbria of *Klebsiella pneumoniae* to human epithelial and urinary bladder. Infect. Immun. 65:1546-1549.
- Rollin, D. and Joseph, S. (2000). Pathogenic microbiology. Maryland university Press USA.
- Sambrook, T.; Fritsch, E. F. and Maniatis, T. (1989). Molecular Cloning, Laboratory manual, Cold Spring Harbour, NY.
- Baron, E. J.; and Finegold, S. M. (1994). Baily and Scotts Diagnostic Microbiology. 8th ed. The C. V. Mosby company St. Louis, Missouri.
- Todar, K. (2007). The Mechanism of Bacterial Pathogenicity (*Klebsiella*). Todars Textbook of bacteriology. Wisconsin. Madison Inc. USA.
- Baron, E. J.; and Finegold, S. M. (1999). Diagnostic Microbiology. 9th ed. Baily and Scotts. The C. V. Mosby company.
- Kirby, A. S.; Posplech, A. K. and Neuman, E. D. (1995). Laboratory Manual of Experimental Microbiology. (2nd ed.). Preparation and Analysis of genomic and plasmid DNA. P16. John Innes Center, Norwich, NR4, 7UH, UK.
- Taheer, N. A. (1998). A study on Actinorhodine-like substance production by *Streptomyces* IQ45. Msc Thesis, college of science, Al-Nahrain University.

- Pons, A.M.; Delaland, F.; Duart, M.; Benoit, S.; Lannelu, I.; Sable, Van Dorsselear, A and Cotte nuae, G (2004).
Wassief, C.D; Cheek, D; Belas, R. (1995). Molecular Analysis of metalloprotease from *Proteus mirabilis*. *J. Bacteriol* 177:5790-5798.
- Podschun, R.; Fischer, A; and Alluman, U. (2000). Expression of putative virulence factors by clinical isolates of *Klebsiella planticola*. *J. Med. Microbiol.* 49(2):115-119.
- Bermud, H.; Jude, F.; Chaibi, E.; Arpin, C.; Labia, R. and Quentin, C. (1999). Molecular characterization of TEM-derived B-Lactamase in clinical isolates of *Klebsiella oxytoca*. *Antimicrob. Agent. Chemother.* 43(7):1667-1681.
- Dionsio, F.; Matic, L.; Radman, M.; Rodringuse O, R and Toddei F. (2002). Plasmid spread very fast in heterogenous bacterial communities. *Genetics*, 162:1525-1532.
- Loclerq, R.; Dorlot, E.; Dural, J, and Courvalin, P. (1988). Plasmid mediated resistance to vancomycin and teicoplanin in *Enterococcus faecium*. *North. Engl. J. Med.* 319:157-161.
- Kafaf, P.A. (2000). Genetic study on antibiotic resistance of some gram negative bacteria isolated from U.T.I. Msc Thesis, college of science, Al-Mustansiryha university.
- Jones, B.D.; Lockotel, D.E.; Johnson, J; and Warren, W. (1990). Construction of urease-negative mutant of *Proteus mirabilis* analysis of virulence in a mouse model in ascending urinary tract infection. *Infect. Immun.* 58:1120-1123.