

Role of *RhIR* and *RhII* Genes in Biofilm and Multidrug Resistance of Burn Isolated *Pseudomonas aeruginosa*.

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Abstract

During the period of 3 months in Baghdad Teaching hospital from March to June 2014, (24) isolates of *Ps. aeruginosa* were assumed from burn infection. Further identification were done by using *rpsL* gene as house keeping gene by PCR. *Ps. aeruginosa* isolates from Baghdad hospital that identified by PCR method, antibiotic sensitivity had done by Kerbey Bauer Disk method for (10) antibiotics which showed that the highest resistance (100%) (n=24) to Tetracyclin, Chloramphenicol, Cefazidime, Cefoxitin and Ceftriaxone, while the resistance to the Cefepime (83.33%) (n=20), Pipracillin (62.5%) (n=15), the same resistance to Ciprofloxacin and Tobramycin (50%) (n=12), Azithromycin (41.66%) (n=10). Results of biofilm formation which detected by TCP Method showed that (66.66%) (n=16) of isolates had high biofilm production most of them are male isolates while the most of moderate production was in female isolates. The highest presence of quorum sensing genes was for *RhIR* gene (83.33%) (n=20) while *RhII* gene was (58.33%) (n=14). Through these results it was observed that there was not any effect of quorum sensing genes on biofilm formation or multidrug resistance while showed relationship between biofilm production and multidrug resistance of negative isolates for quorum sensing genes.

Introduction

Ps. aeruginosa is an opportunistic pathogen widely distributed in the environment and had a versatile metabolic activity. This non-fermenting Gram negative rod is responsible for about (10%) of all hospital acquired infections (Aloush, et al., 2006; Enoch, et al., 2007). Infections are opportunistic in nature and ranged from those associated with ventilator, catheter, burn and wound, to pulmonary infections in cystic fibrosis patients and keratitis in contact lens wearers (Lee, et al., 2003; Choyet, et al., 2008). In general, the bacteria control their environmental systems and cell populations through intracellular communications to have the best performance and response according to the demographic and the environmental conditions (Dechoet, et al., 2010; Horswill et al., 2007). Pathogenic bacteria take the best advantages of communication capability, as an example, they can overcome the host immune system barrier using the community, in this case, they estimate the cell density using these intercellular signals and given the concentration of signal transmitter factors, and when the density reaches its minimum extent in which the immune system cannot simply cope, bacteria will release the virulence factors, thus, the host immune system will be prevented to deliver rapid responses. The process is controlled by a system in bacteria called "Quorum Sensing" (Li, & Tian, 2012; Waters, & Bassler, 2005). Biofilm formation has tremendous detrimental effects and has brought huge amounts of problems to our everyday life. In addition, biofilms show resistance to a wide range of antibiotics such as ampicillin, streptomycin, tetracyclines, gentamicin etc. Also, the conventional methods such as antibiotics, and disinfectants to control the infections caused by biofilm formation are often ineffective due to their special physiology and physical matrix barrier (Stewart, 2002). The aim of this study was to show the relationship between biofilm formation and multidrug resistance and determine the role of quorum sensing.

Materials and Methods

Bacterial isolates: Sample were collected from burn infections and initially characterized as *Ps. aeruginosa* based on biochemical tests and gram staining, according to the criteria established by (Forbes et al., 2002), and by PCR method used *rpsL* house keeping gene primer (Alpha DNA/Canada) Table(1). The PCR amplification was performed to these primers in a total volume (25) μ L containing (5) μ l DNA template, (12.5) μ L Go Taq Green Master Mix 2X (Promega/USA), (1) μ L of each primer (10) pmol, then volume was completed to (25) μ L by nuclease free water.

Thermal cycler was programmed as following conditions:

Initial denaturation 95 $^{\circ}$ C for 5min, followed by 30 cycles each cycle included 94 $^{\circ}$ C for 30 sec, 57 $^{\circ}$ C for 30 sec and 72 $^{\circ}$ C for 1min then final extension on 72 $^{\circ}$ C for 7 min. The PCR products were separated by 1% agarose gel electrophoresis and visualized by exposure to Ultra Violet Light.

1- Antimicrobial susceptibility test: disk diffusion test had done for (Tetracyclin(10)µg, Chloramphenicol(10)µg, Ceftazidime(10)µg, Cefoxitin(10)µg , Ceftriaxone(30)µg, Cefepime(30)µg, Piperacillin(30)µg, Ciprofloxacin(10)µg, Tobramycin(30)µg and Azithromycin(30)µg (Bioanalyse/Turkey). According to the (CLSI, 2013).

Biofilm Production: was detected by Tissue culture Plate Method (TCP) as described by (Sandoeet *al.*, 2003) this is a qualitative method for biofilm detection. A loopful of tested organism inoculated in (10) ml Brain Heart Infusion broth then tubes were incubated at 37c° for 24 h. After incubation tubes were centrifugated to remove the media and the bacterial suspension was washed with normal saline, then filled the flat bottom tissue culture plates(96)wells with (180)µL of brain heart infusion broth then (20) µL from bacterial suspension was added to the wells, the culture plate incubated at 37c° for 24h. After incubation the wells were washed with normal saline by ELISA washer to remove free-floating bacteria. Then (200)µL of formaldehyde was added to the wells for (10)min. Biofilm which remained to the walls and the bottoms of the wells stained with (200) µL of 0.1% Crystal violet for (10)min, excess stain was washed with normal saline and plates were dried then (200)µL of destaining solution was added (95% Etanol) for (10) min, finally (200)µL from each well was transferred to a new microtiter plates and measured at (630)nm by microplate reader.

Detection of *RhII* and *RhRI* genes by PCR: Genotype detection of these genes by specific primers Table(1). under following conditions: Initial denaturation 94 c° for 5 min, 35 cycles included: denaturation at 94c° for 30 sec, primer annealing with DNA template at 50 c° for 30 sec, extension at 72c° for 30 sec, and final extension at 72c° for 10 min, these condition for the two genes.

Table(1): Sequences and product size of specific primers

Gene		Sequence of primers (5'-3')	Product size bp	reference
<i>rpsL</i>	F	GCAAGCGCATGGTCGACAAGA	210	Xavier <i>et al.</i> , 2010
	R	CGCTGTGCTCTTGCAGGTTGTGA		
<i>RhIR</i>	F	CAATGAGGAATGACGGAGGC	370	Sandoeet <i>al.</i> , 2003
	R	GCTTCAGATGAGGCCAGC		
<i>RhII</i>	F	CTTGATCATGATCGAATTGCTC	625	Sandoeet <i>al.</i> , 2003
	R	ACGGCTGACGACCTCACAC		

Results and Discussion

A total of *Ps. aeruginosa* isolates were recovered from burn infections (n=24) which identified by PCR method according to house keeping gene *rpsL* Table 2. as described in (Xavier *et al.*, 2010).

The resistance patterns of *Ps. aeruginosa* isolates to (10) antibiotics were shown in Table 3. The highest resistance was observed to Tetracyclin, Chloramphenicol, Ceftazidime, Cefoxitin and Ceftriaxone which was (100%), followed by Cefepime (83.33%) and Piperacillin (62.5%), while the isolates show the same resistance to the Ciprofloxacin and Tobramycin (50%) and Azithromycin (41.66%). (Senturket *al.*, 2012) Observed that most of burn *Ps. aeruginosa* isolates showed high resistance rates to cefepime (98%) followed by piperacillin-ceftazidime (91%) and ceftriaxon (87%) All isolates were totally resistant to tetracycline. MDR *P. aeruginosa* develops resistance by various mechanisms like multidrug resistance efflux pumps, production of β-lactamases, aminoglycoside modifying enzymes, and decrease outer membrane permeability (Mahmoud *et al.*, 2013).

TCP Method showed that (66.66%) of isolates had high biofilm production most of them are male isolates while the most of moderate production was in female isolates Table 2. The association between the potential to form strong biofilms by *P. aeruginosa* and antibiotic resistance has also been shown which (Lambert, 2002).

The highest presence of quorum sensing genes was for *RhIR* gene (83.33%) while *RhII* gene was (58.33%). Bacteria are known as one of the simplest and the most primitive life forms, which have the single-cell life and their reproduction, feeding and communication mechanisms are identified as basic and simple patterns, however, the complex and interesting mechanisms are provided by studies in recent decades that control bacterial behaviors similar to what occurs in multicellular organisms. This mechanism is controlled by quorum sensing

system that enables intraspecies and interspecies communication (Drenkard, & Ausubel, 2002; Rutherford, & Bassler, 2012). In current study was observed that negative isolates (4) showed resistance for all antibiotics Table 2. So it due to think that was no role of Quorum sensing in antibiotic resistance while it was observed high biofilm producer isolates showed resistance for all antibiotics.

Table 2: TCP Method and PCR Results.

Male or female	TCP method	rpsL	RhIR	RhII
Female	moderate	+	+	-
Female	moderate	+	+	-
Female	moderate	+	+	+
Male	moderate	+	+	+
Male	high	+	+	+
Male	high	+	+	+
Male	high	+	+	+
Male	high	+	+	+
Female	moderate	+	+	+
Female	moderate	+	+	+
Female	high	+	+	+
Female	moderate	+	+	-
Male	high	+	+	+
Female	high	+	+	-
Female	high	+	+	-
Female	high	+	-	-
Female	moderate	+	+	+
Male	high	+	+	-
Male	high	+	-	-
Male	high	+	-	-
Female	high	+	-	-
Male	high	+	+	+
Female	high	+	+	+
Male	high	+	+	+
Male	high	+	-	-

Table3:Antibiotic sensitivity test

TE	CIP	C	CAZ	FOX	TOB	PRL	FEP	CRO	AZM
R	S	R	R	R	R	R	R	R	R
R	R	R	R	R	R	R	R	R	R
R	S	R	R	R	R	R	R	R	I
R	S	R	R	R	R	R	R	R	S
R	R	R	R	R	S	R	R	R	S
R	R	R	R	R	S	R	R	R	S
R	R	R	R	R	S	R	R	R	S
R	R	R	R	R	S	R	R	R	S
R	S	R	R	R	S	S	S	R	R
R	S	R	R	R	R	R	R	R	R
R	S	R	R	R	S	S	S	R	S
R	S	R	R	R	S	S	S	R	R
R	S	R	R	R	S	S	R	R	S
R	R	R	R	R	R	S	R	R	S
R	R	R	R	R	R	S	R	R	S
R	R	R	R	R	R	R	R	R	R
R	S	R	R	R	S	R	R	R	R
R	R	R	R	R	R	R	R	R	S
R	R	R	R	R	R	R	R	R	R
R	R	R	R	R	R	R	R	R	R
R	R	R	R	R	R	R	R	R	I
R	R	R	R	R	R	R	R	R	R
R	R	R	R	R	R	R	R	R	R
R	S	R	R	R	S	S	S	R	S
R	S	R	R	R	S	S	R	R	R
R	S	R	R	R	S	S	R	R	S

References

- 1- Aloush, V.; Navon-Venezia, S., Seigman-Igra Y., Cabili S. and Carmeli Y. (2006). Multidrug-resistant *Pseudomonas aeruginosa*: Risk factors and clinical impact. *Antimicrobial Agents and Chemotherapy* 50(1), 43-48.

- 2- Enoch, D. A., Birkett, C. I. and Ludlam, H. A. (2007). Non-fermentative Gram-negative bacteria. *International Journal Antimicrobial Agents* 29(3), S33-41.
- 3- Lee, E.J., Cowell, B.A., Evans, D.J. and Fleiszig, S. M. (2003). Contribution of ExsA-regulated factors to corneal infection by cytotoxic and invasive *Pseudomonas aeruginosa* in a murine scarification model. *Investigative Ophthalmology & Visual Science* 44(9), 3892-3898.
- 4- Choy, M. H., Stapleton, F., Willcox, M. D. P. and Zhu, H. (2008). Comparison of virulence factors in *Pseudomonas aeruginosa* strains isolated from contact lens- and non-contact lens-related keratitis. *Journal of Medical Microbiology* 57(12), 1539-1546.
- 5- Decho AW, Norman RS, Visscher PT (2010) Quorum sensing in natural environments: emerging views from microbial mats. *Trends Microbiol* 18: 73-80
- 6- Horswill AR, Stoodley P, Stewart PS, Parsek MR (2007) The effect of the chemical, biological, and physical environment on quorum sensing in structured microbial communities. *Anal Bioanal Chem* 387: 371-380
- 7- Li, YH1 and Tian, X. (2012) Quorum sensing and bacterial social interactions in biofilms. *Sensors (Basel)* 12: 2519-2538.
- 8- Waters, C.M. and Bassler, B.L. (2005). Quorum sensing: cell-to-cell communication in bacteria. *Annu Rev Cell Dev Biol* 21: 319-346.
- 9- Stewart, P.S. (2002). Mechanisms of antibiotic resistance in bacterial biofilms. *Int. J. Med. Microbiol*, 292, 107-113
- 10- Forbes, B.A.; Sahm, D.F.; and Weissfeld, A.S. (2002). *Diagnostic Microbiology* 11th edition. Mosby, Inc. Baltimore, USA. 302-309.
- 11- Xavier, D.E.; Renata, C.P.; Requel, G.; Lorena, C.C.F. and Ana, C.G. (2010). Efflux pump expression and its association with porin down-regulation and β -lactamase production among *Pseudomonas aeruginosa* causing bloodstream infection in Brazil. *BMC. microb.* 10:217.
- 12- CLSI. (2013). Performance Standards for Antimicrobial Susceptibility Testing ; Twenty-third informational supplement , M 100-S23 .
- 13- Sandoe, J.A.; Witherden, I.R.; Cove, J.H.; Heritage, I. and Wilco, M.H. (2003). Correlation between enterococcal biofilm formation in In Vitro and medical- device related infection. *In. J. Microbiol.* 2:545-500.
- 14- Senturk, S.S.U ; Gulgun, B. T.A. and Ulusoy, S. (2012). Quorum sensing and virulence of *Pseudomonas aeruginosa* during urinary tract infection. *J. Infect. Dev. Ctries.* 6(6):501-507.
- 15- Mahmoud, A, B.; Zahran, W.A.; Hindawi, G.R.; Labib, A.Z. and Galal, R. (2013) Prevalence of Multidrug-Resistant *Pseudomonas aeruginosa* in Patients with Nosocomial Infections at a University Hospital in Egypt, with Special Reference to Typing Methods. *Journal of Virology & Microbiology*
- 16- Lambert, P.A. (2002) Mechanisms of antibiotic resistance in *Pseudomonas aeruginosa*. *J. R Soc Med*; 95(Suppl. 41):22-26.
- 17- Drenkard, E. and Ausubel, F.M. (2002). *Pseudomonas* biofilm formation and antibiotic resistance are linked to phenotypic variation. *Nature*; 416(6882):740-3.
- 18- Rutherford, S.T and Bassler BL (2012) Bacterial quorum sensing: its role in virulence and possibilities for its control. *Cold Spring Harb Perspect Med* 2.
- 19- Miller, M.B.; Bassler, B.L. (2001) Quorum sensing in bacteria. *J. Annu Rev. Microbiol.* 55: 165-199

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