

Control of Fungi Isolated from Date Palm Fruit in Yola, Adamawa State

Anjili S. M. Channya F. K. Chimbekujwo I. B.*

Department of Plant Science, Modibbo Adama University of Technology, Yola

Abstract

Samples of dates were purchased from four (4) major markets in Yola, and transferred to Modibbo Adama University of Technology. Isolation of moulds associated with the rot was carried out on solid Potato Dextrose Agar (PDA) on 9 cm diameter Petri-dish. Four fungal isolates were obtained and proved through pathogenicity test to be the responsible pathogens. These were *Aspergillus niger*, *Fusarium solani*, *Scopulariopsis brevicaulis*, and *Rhizopus stolonifer*. In-vitro and in-vivo control trials with ash and garlic oil showed effective mycelia growth inhibition at $p=0.001$. Inhibition improved with increase in concentration of the garlic oil and ash. Heat treatment equally proved worthwhile with significant inhibition compared to the control at $p=0.001$. Increase in temperature (0-550C) was rewarded with better control effect.

Keywords: Date palm, Fungi

1. Introduction

Date palm (*Phoenix dactylifera* L.) is a palm in the genus *Phoenix*, cultivated for its edible sweet fruit (Wikipedia, 2014). Dates are widely distributed facilitated by the fact that dates lend themselves perfectly to being carried along as a high calorie food, with a long-keeping quality (FAO, 2012). Date fruit also can be made into juice, vinegar, wine, beer, sugar, syrup, honey, chutney, pickle, paste, dip, and food flavoring (Barreveld, 1993 and Glasner *et al.*, 2002). It is an ideal food for all age phases, providing the most important essential nutrients like protein, fiber, carbohydrates, fat and minerals (Al-Farsi *et al.*, 2005 and Vyawahare *et al.*, 2009).

The plant is affected by various pests and diseases, but the nature of the problem varies with geographic location (Howard *et al.*, 2001). Atia (2011) observed that the date palm fruits are mostly loaded with mixture of microbes: bacteria, moulds and yeast but people go ahead eating after clearing the pericarp, while some eat it whole irrespective of the state of the pericarp. Species of *Alternaria*, *Aspergillus*, *Fusarium* and *Penicillium* cause fruit rots of date-palm (Bokhary, 2010).

A report stated that the major agents of date palm spoilage are moulds, followed by bacteria and yeast at all stages of ripening on trees, as well as during storage and processing (Amal *et al.*, 2014). Effects of rot causing organisms along with size, colour, texture, cleanliness, freedom from defects (sunburn, insect damage, sugar migration to surface, fermentation) are the important considerations affecting fruit quality (ChihCheng and Robert, 2007)

Some pathogens and potential mycotoxin producing microbes isolated from date fruits include species of *Aspergillus*, *Penicillium*, *Alternaria* and *Fusarium* (Kader, 2007; Hamad, 2008; Hayrettin *et al.*, 2012; Aido *et al.*, 1996; El-Sherbeeney *et al.*, 1985 and Abdulsalam *et al.*, 1991).

Hawking of date palm fruits is a common sight especially north of Nigeria where Yola is located. The high patronage this merchandise attracts (both fresh and dried fruits) is a cause for pathological concern in light of reports indicating that some potential mycotoxin producing moulds are implicated. Also, Christensen *et al.* (2007) reported that the agricultural industries sustained huge crop losses as a result of fungal diseases of fruits and plants. The Food and Agricultural Organization of the United Nations (FAO) has estimated that up to 25% of the world's food crops are significantly contaminated with mycotoxins (WHO, 1999). Mycotoxins are highly toxic and cause severe intoxications in human and animals, some of them are carcinogens (Khomutov *et al.*, 2011).

Amal *et al.* (2014) observed that methods that will curb the microbial infestation of fruits at post-harvest storage are required and should be made mandatory. The use of fungicides for the control of plant diseases is a common practice all over the world (Wazir, 2013). The use of chemical in control of plant disease posed a risk to the survival of human race (UNEP, 1995). Therefore there is need for other control means to this important menace.

Reports on the successful control of certain moulds associated with postharvest rot of crops are available (Agbeniyi, 1988; Channya and Chimbekujwo, 2002; Bristone *et al.*, 2011).

Many spices and herbs exert antifungal activity due to their essential oil fractions. The thyme oil had strong fungicidal effect on *Penicillium* and *Alternaria* (Mironescua and Georgescub, 2008), *Alternaria alternata* (Hadizadeh *et al.*, 2009). The essential oil of *Mentha spicata* showed cidal effect on mycelial growth of *Aspergillus ochraceus*, *Penicillium digitatum*, *Pyricularia oryzae* and *Alternaria alternata* (Singh *et al.*, 2006). Essential oil of *Ocimum gratissimum* inhibited the growth of *Botryosphaeria rhodina*, *Rhizoctonia* spp and two strains of *Alternaria* spp (Terezinha *et al.*, 2006). Post-harvest treatment with heat has been used to reduce decay

in several crop including pumpkin, strawberry and apple (Spotts and Chen, 1987).

This study seeks to try out garlic oil, ash and heat treatments as alternative control to this important menace that puts to waste this important crop with an antecedent health risk to consumers.

2. Material and Methods

The research was conducted in 2015. Date palm (*Phoenix dactylifera* L.) showing spoilage was obtained from four major markets in Yola which are Jimeta Old Market, Jimeta Ultra-Modern Market, Jimeta Shopping Complex and Yola Town Market. A sample of 250 fruits were randomly collected from sellers in each market. These were brought to the laboratory of Plant Science Department, Modibbo Adama University of Technology (MAUTECH) Yola, where isolation, pathogenicity test and control trials were carried out.

2.1. Isolation of Fungi from Date Palm Fruits

Under aseptic conditions the diseased sample from date palm fruit showing rot was sectioned into approximately 5 mm square with a heat-sterilized scalpel. The pieces were immersed into 1% sodium hypochlorite contained in a sterile 9 cm diameter Petri- dish for surface sterilization for 30 seconds using sterile forceps. The sterilized pieces were rinsed in three changes of sterile distilled water and then dried between sterile filter papers. With a flamed and cooled pair of forceps, a sterilized piece of date palm was blotted dry between sterile filter papers, then plated aseptically on 9 cm diameter Petri-dish containing sterile solidified Potato Dextrose Agar (PDA) and incubated at room temperature of $33\pm 2^{\circ}\text{C}$ for 7 days. Sub- culturing on fresh sterilize PDA using the method of Klick and Pitt (1988) and Robert *et al.* (1996) was carried immediately when new colonies began to grow through hyphal-tip transfer.

2.2 Identification of Fungi

Microscopic examination was carried out to observe the structure and characteristics of the fungal isolates in addition to macroscopic cultural observations. A sterile needle was used to pick a little portion of the hyphae containing spores and placed on a sterile glass slide stained with Lactophenol cotton blue and examined under the photographic microscope using the method of Fawole and Oso (1995). Micrograph of the isolates showing (conidia etc.) were taken. The morphological and cultural characteristics observed were compared with structures in the identification guides of the International Mycological Institute Kew and of Hunter and Barnett (1998).

2.3. Pathogenicity Test

Healthy date fruits (semi ripe) were obtained and surface-sterilized with 1% sodium hypochlorite for 30 seconds and rinsed in three (3) changes of sterile distilled water according to the method of Chukwuka *et al.* (2010). A sterile cork borer (2 mm in diameter) was used to puncture and inject healthy date fruits with spores' suspension of each isolated fungus in three replicates. Removed tissue was replaced and vaspar jelly was smeared to completely seal each hole to avoid contaminant. It was kept at room temperature of $33\pm 2^{\circ}\text{C}$. A similar set up was placed as control using distilled water to complement the fungal inocula. The set up was arranged in a completely randomized design. It was incubated for seven (7) days to allow for possible rot development and the isolates were re-isolated from the new host and shown to be the same as the originally inoculated pathogen.

2.4. Control Trials with Ash, Garlic oil

In-vitro control was carried out using food poisoning method of Raja *et al.* (2009). Different weights (grams) of heat-sterilized tomato leaf ash (at 160°C for 1 hour in an oven) of 1g, 3g, and 5g were added aseptically using a heat-sterilized spartula on cool sterilized PDA medium in conical flask swirled to mix before pouring into 9 cm Petri-dish, then allowed to solidify, followed by inoculation with 2 mm radius of 7-day old pure culture of each of the isolated fungi sectioned with a sterile cork-borer (*Aspergillus niger*, *Fusarium solani*, *Scopulariopsis brevicaulis* and *Rhizopus stolonifer*). Similar set up was made with ash free to serve as control. The fungi were inoculated at the centre of 9 cm diameter Petri-dish sealed with masking tape. The set up were inoculated for seven (7) days at room temperature of $33\pm 2^{\circ}\text{C}$ in the month of June and radial mycelial growth of each isolate was measured with caliper, rule, and with the aid of hand lens and recorded for analysis.

Different volumes of garlic oil were tested [0ml (control)], 2 ml, 3 ml and 5 ml]. These were added aseptically to cool sterile PDA medium in conical flasks and agitated before pouring into Petri-dishes. Similarly, 2 mm radius of 7-day old pure culture of each of the isolated fungi was inoculated on each of the media. Oils free medium served as control treatment. Petri-dishes were sealed with masking tape and similarly incubated and assessed. All *in-vitro* settings were of completely randomized design replicated thrice.

The method of Channya (1991) was used for the *in-vivo* test, in which a sterile 2 mm diameter cork borer was used to puncture and inject sterilized healthy semi-ripe fruits with spores' suspension of 7-day old pure culture of each of the isolated fungi using sterile needle. Weight of 0.5 g, 1.0 g and 1.5 g of ash was subjected to

the wounded date palm fruits. Inoculated fruits were put in sterile transparent polythene bags and incubated as for the *in-vitro* set up. Similar set up was made with ash free to serve as control. Three replicates for each treatment was used and plated at the centre of the 9 cm diameter Petri-dish sealed with masking tape. Radial rot of fruits was measured with caliper and rule and with the aid of hand lens.

A sterilized sample of date palm fruit was punctured with a sterile 2 mm diameter cork borer and spores' of the isolated fungi were inoculated to it as stated above. Similarly, 0.2, 0.3, and 0.4 ml of garlic oil were injected respectfully into the wounded date palm fruits. Oil free medium served as control. The set up was a completely randomized design. The set up were inoculated for seven (7) days at room temperature $33\pm 2^{\circ}\text{C}$ and radial rot of each fruit was measured with caliper and rule, and with the aid of hand lens.

2.5 Control using heat

Effect of heat on the fungal isolates was carried out using the method of Harrak and Chetto (2002) and Harrak (2003). An electric oven was set at four temperature regimes of room temperature of 33°C (control), 45°C , 50°C and 55°C for 3 hours. The isolates was inoculated into healthy semi-ripe date palm fruits by using a sterile 2 mm diameter cork borer to puncture and inject sterilized fruit with spores' suspension of the isolated organisms using 2.0 ml syringe. Samples were transferred into open glass Petri-dishes (9 cm in diameter) with a filter paper inserted to the lower lid, then incubated in an electric oven according to the above-stated temperatures. After treatment on hot air, samples were inoculated for seven (7) days and the filter paper was moistened with sterile distilled water once daily throughout the period. The set up was a completely randomized design in three replicates. The radial rot of each fruit was measured with caliper and rule, and with the aid of hand lens.

Results were analysed using Statistical Analysis System (SAS) version 7 and the means that were significant were separated using the Least Significant Difference (LSD).

3. Result and Discussion

Aspergillus niger, *Fusarium solani*, *Scopulariopsis brevicaulis* and *Rhizopus stolonifer* were the isolates that were associated with the rot of date palm fruits in Yola markets. The pathogenicity tests on these isolates proved they were the rot pathogens.

Analysis of variance for the efficacy of tomato ash on these moulds ($p < 0.0001$) was very significantly different from the control set up for both *in-vitro* and *in-vivo* tests. The performance of ash at different levels is shown in Tables 1 and 2 respectively. There was however a considerably lower effect of ash on *Scopulariopsis brevicaulis* than the other moulds. Higher quantity of ash produced a corresponding better control. This collaborates reports by Channya and Chimbekujwo (2002) that wood ash effectively controlled fungal rot of plantains (*Musa parasidiaca* L.). Agbeniyi (1988) also reported that when kola nut (*Cola nitida*) was treated with ash of *Vernonia* spp, *Tectonia grandis* and *Cassia siemia* before storage, it reduced the incidence of moulds. Bristone *et al.* (2011) reported that when tubers of sweet potatoes were treated with wood ash, rot caused by *Rhizopus stolonifer* and *Penicillium expansum* was reduced to minimal level. Agbeniyi (1988) and Channya (1991) adduced the performance of ash to pH of ash that is highly alkaline. The ash of tomato was found to have a pH of 9.

Control trials with garlic oil *in-vitro* and *in-vivo* produced even a better growth inhibition of all the moulds at $p < 0.0001$. Tables 3 and 4 show the effect of the different concentrations on the mycelial radial growth of the moulds with *Aspergillus niger* being the most inhibited compared to the control while *Scopulariopsis brevicaulis* been the least affected by garlic oil *in vitro*. *In-vivo* test result however shows that *Scopulariopsis brevicaulis* and *Rhizopus stolonifer* were the most susceptible to garlic oil control. Inhibition improved with increase in concentration of garlic oil for both *in-vitro* and *in-vivo* tests. The result is also similar to that of Atia (2011) who reported that increased in oil concentration resulted in reducing percentage of mycelia growth of the tested fungi. Garlic oil (*Allium sativum*) contain diallyl thiosulfinate as an active compound (Burt, 2004; Zaika, 1988; Ceylan, 2004). *Allium sativum* (garlic) juice extract belongs to such non-traditional treatments and among the natural fungicide substances, it has been found most active against many fungal species (Curtis *et al.*, 2004; Slusarenko *et al.*, 2008). Allicin is the most important biologically active substance of *A. sativum* crude extract; it is formed from its precursor, alliin, by the action of allinase enzyme (Josling, 2003). Pharmacologically, allicin is the most important and the most active substance and it is found in the fresh extract of *Allium* (Vasile *et al.*, 2012).s

Alice and Rao (1987) observed that *A. sativum* extracts significantly reduced seed infection by *Drechslera* on rice and treated seeds had significantly higher viability. On barley in green house and field experiments, allicin used as elicitor was as effective as fungicide against the leaf spot severity caused by *Bipolaris sorokiniana* (Silva *et al.*, 2001; Rodrigues *et al.*, 2002; Rodrigues and Bach, 2003; Antoniazzi *et al.*, 2008). Garlic extract treatment of wheat seeds significantly reduced the incidence of seed-borne fungi, increased seed germination, the number of healthy seedlings and vigour index (Grozav and Fource, 2005; Khalaf *et al.*, 2011).

Table 1: Effect of Ash on Fungal Pathogens of Date Fruit *In-vitro*

| Concentration (g) | Means for mycelial radial growth (mm) | | | |
|-------------------|---------------------------------------|------------------|-----------------------|----------------------|
| | <i>A. niger</i> | <i>F. solani</i> | <i>S. brevicaulis</i> | <i>R. stolonifer</i> |
| 1 | 8.26 | 8.23 | 12.53 | 30.21 |
| 3 | 7.75 | 6.65 | 12.01 | 28.61 |
| 5 | 6.62 | 5.51 | 8.90 | 16.80 |
| 0 | 17.28 | 17.18 | 13.98 | 40.23 |
| LSD (0.0001) | 1.21 | 1.26 | 1.53 | 4.47 |

Table 2: Effect of Ash on Fungal Pathogens of Date Fruit *In- vivo*

| Concentration (g) | Mean of mycelial radial growth (mm) | | | |
|-------------------|-------------------------------------|------------------|-----------------------|----------------------|
| | <i>A. niger</i> | <i>F. solani</i> | <i>S. brevicaulis</i> | <i>R. stolonifer</i> |
| 0.5 | 8.89 | 6.10 | 6.52 | 6.52 |
| 1.0 | 7.09 | 5.63 | 3.93 | 5.78 |
| 1.5 | 3.59 | 4.94 | 1.49 | 4.72 |
| 0 | 10.43 | 6.12 | 8.26 | 7.94 |
| LSD (0.0001) | 1.19 | 0.56 | 1.31 | 0.70 |

A report by Hassan *et al.* (2005) showed that garlic completely controlled the intensity of *B. sorokiniana* and *Fusarium* spp after the treatment of wheat seeds. It is generally accepted that the essential oil components act on the functionality and the structure of the cell membrane (Viuda-Martos *et al.*, 2008). Low concentrations result in changes of the cell structure, inhibiting respiration and changing the permeability of the cell membrane, whereas high concentrations lead to severe membrane damage, loss of homeostasis and cell death (Carson *et al.*, 2002).

Conner and Beuchat (1984) suggest that the antifungal activity is the product of essential oil components' interaction with enzymes responsible for energy production and the synthesis of structural compounds of the cell. On the other hand, Omidbeygi *et al.* (2007) suggested that the essential oil components pass through the cell membrane, integrating with enzymes and proteins of membranes, causing loss of macromolecules from the interior of the cell, leading to changes in the cell and ultimately to its death. Cristani *et al.* (2007) indicate that the antifungal activity of terpene relates to their ability to act not only on the permeability, but also on other functions of cell membranes. These components can pass through the cell membrane and interact with intracellular structures. Daferera *et al.* (2000) state that fungitoxic effect of essential oil is a consequence of hydrogen bonds formation between hydroxyl groups of phenolic compounds and active sites of cellular enzymes.

According to Sharma and Tripathi (2006), the active components cause loss of integrity of the cell wall, and thus the loss of cytoplasmic constituents from the cells of hyphae. Lucini *et al.* (2006) indicated that the inhibition of mycelial growth of fungi is caused by monoterpenes. These components can increase the concentration of lipid peroxides, such as hydroxyl, alkoxy and alkoperoxy radicals, leading to the destruction of cells.

It has been observed that application of essential oils for postharvest disease control of fresh product, as a novel emerging alternative to hazardous anti-fungal treatments will allow a safer and environmentally more acceptable management of post-harvest diseases (Hadizadeh *et al.*, 2009). The antifungal activity of the essential oils is different, depending on the mould type (Mironescua and Georgescub, 2008). Suwitchayanon and Kunasakdaku (2009) reported that the inhibitory effects of plant oils might be regarded as cidal agent against fungal growth and showed abnormal conidia and malformations as swollen, often septated and pale colour of hypha.

Table 3: Effect of Garlic Oil on Fungal Pathogens of Date Palm Fruit Rot (*In- vitro*)

| Concentration (ml) | Mean of radial mycelia growth (mm) | | | |
|--------------------|------------------------------------|------------------|-----------------------|----------------------|
| | <i>A. niger</i> | <i>F. solani</i> | <i>S. brevicaulis</i> | <i>R. stolonifer</i> |
| 2 | 10.35 | 12.65 | 13.05 | 35.03 |
| 3 | 9.55 | 10.24 | 9.93 | 33.61 |
| 5 | 7.23 | 4.71 | 6.10 | 27.51 |
| 0 | 17.28 | 17.18 | 13.98 | 40.23 |
| LSD (0.0001) | 1.21 | 1.43 | 4.24 | 1.24 |

Table 4: Effect of Garlic Oil on Fungal Pathogens of Date Palm Fruit Rot (*In-vivo*)

| Concentration (ml) | Mean of radial mycelia growth (mm) | | | |
|--------------------|------------------------------------|------------------|-----------------------|----------------------|
| | <i>A. niger</i> | <i>F. solani</i> | <i>S. brevicaulis</i> | <i>R. stolonifer</i> |
| 0.2 | 7.16 | 4.62 | 3.90 | 3.23 |
| 0.3 | 4.33 | 3.39 | 3.39 | 1.19 |
| 0.4 | 3.40 | 2.04 | 1.29 | 0.91 |
| 0 | 10.43 | 6.12 | 8.26 | 7.94 |
| LSD (0.0001) | 1.38 | 0.74 | 1.20 | 1.16 |

Treated date fruit with heat showed effective growth inhibition as compared with control at $p=0.0001$ for all the pathogens. Temperature increase within the range of 45-55 °C had a corresponding improvement in mycelial growth inhibition. *Scopulariopsis brevicaulis* and *Fusarium solani* were the most susceptible to temperature control (Table 5). These temperature regimes did not change the physiology of the fruit. This is also similar with the result of Atia (2011) whose treatment on Deglate and Elak varieties of date reduced the number of date fruits infected by fungi compared with the untreated controls. Similarly, Harrak and Chetto, (2002) and Harrak, (2003) reported that heat treatment at 60 for 1-1.5 h lead to complete control other organisms on stored date and did not change its physical or chemical characters. Kader and Hussein (2009) also reported that, drying the dates to 20% moisture or lower greatly reduced incidence of moulds and yeasts.

Table 5: Effect of Heat on Fungal Agents of Date palm Fruit Rot

| Temperature (°C) | Mean of radial mycelia growth (mm) | | | |
|------------------|------------------------------------|------------------|-----------------------|----------------------|
| | <i>A. niger</i> | <i>F. solani</i> | <i>S. brevicaulis</i> | <i>R. stolonifer</i> |
| 45 | 6.64 | 3.51 | 2.63 | 3.27 |
| 50 | 6.01 | 1.86 | 0.76 | 2.97 |
| 55 | 5.98 | 1.12 | 0.47 | 0.58 |
| 0 | 10.43 | 6.12 | 8.26 | 7.94 |
| LSD (0.0001) | 1.01 | 0.86 | 1.41 | 1.11 |

4. Conclusion

Fungal contamination is a direct relationship with both the physical initial dates and environmental conditions of the premises including the storage temperature and humidity with consequent decrease on the market value. The sensitivity of dates to fungal spoilage is related to its poor conservation of places of production and storage which is a big challenge to operators. This alteration is particularly important as the storage conditions and storage are inadequate

Therefore ash, garlic oil and heat have the potential to reduce the date loss and consumer exposure to mycotoxin poisoning and these are more environmentally friendly than fungicides.

References

- Abdulsalam K. S., Ahmed A. M. and Ahmed A. M. (1991). The Effect of Three Fungi and Their Combinations on the Chemical Constituents of Two Cultivars of Date Palm Fruits. *Emirate Journal of Agricultural Science*, 3:81-95.
- Agbeniyi, S. O. (1988). Efficacy of Milton Solution and Wood Ash in the Control of Storage Moulds of Kola nut (*Cola nitida*). A paper presented at *Nigeria society for plants protection* (NSPP). Bida Nigeria, 21-24.
- Aido K. E., Tester R. F., Morrison J. E. and Mac-Farlane, D. (1996). The Composition and Microbial Quality of Prepacked Dates Purchased in Greater Glasgow. *International Journal of Food Science and Technology*, 31:433-438.
- Al-Farsi, M., Alasalvar, C., Morris, A., Baron, M. and Shahidi, F. (2005) "Comparison of Antioxidant Activity, Anthocyanins, Carotenoids, and Phenolics of three Native Fresh and Sun-dried Date (*Phoenix dactylifera* L.) Varieties Grown in Oman," *Journal of Agricultural and Food Chemistry*, 53(19): 7592–7599.
- Alice D. and Rao A. V. (1987). Antifungal Effects of Plant Extracts on *Drechslera oryzae* in Rice. *International Rice Reserve, Newsletter*. 12(2):28.
- Amal, A. A., Afaf, I. S., Humaira, R., Nadine, M. S. M., Ali, A. A., Anjana, M. and Gehan, E. (2014). Postharvest Fruit Spoilage Bacteria and Fungi Associated with Date palm (*Phoenix dactylifera* L.) from Saudi Arabia. *African Journal of Microbiology Research*, 8(11): 1228-1236.
- Antoniazzi N., Deschamps C. and Bach E. E. (2008). Effect of Xanthan and Allicin as Elicitors Against *Bipolaris sorokiniana* on Barley Infield Experiments. *Journal of Plant Disease and Protection*, 115:104-107.
- Atia, M. M. M. (2011). Efficiency of Physical Treatment and Essential Oil in Controlling Fungi Associated with Some Stored Date Palm Fruits. *Australian Journal Basic Applied Science*, 5(6):1572.
- Barreveld, W. H. (1993). Date Palm Products (Food and Agriculture Organization of the United Nations, Agricultural Services Bulletin no. 101, *Food and Agriculture Organization of the United*

- Nations, Rome, Italy).*
- Bokhary, H. A. (2010). Seed-borne Fungi of Date Palm, *Phoenix dactylifera* L. from Saudi Arabia. *Saudi Journal of Biological Sciences*, 17: 327–329.
- Bristone, B., Chimbekujwo, I. B. and Pukuma M. S. (2011). Control of Post-harvest Fungal Rot of Sweet Potatoes (*Ipomea batatas*) in Yola. *Nigerian Journal of Botany*, 24(1): 43-51.
- Burt S. (2004). Essential Oils: their antibacterial properties and potential applications in foods - a review. *International Journal of Food Microbiology*, 94(3): 223-253.
- Carson C. F., Mee B. J. and Riley T. V. (2002). Mechanism of Action of *Melaleuca alternifolia* (tea tree) Oil on *Staphylococcus aureus* Determined by Time-kill, Lysis, Leakage and Salt Tolerance Assays and Electron Microscopy. *Antimicrobial Agents and Chemotherapy*, 46:1914-1920.
- Ceylan E., and Fung Y. C. D. (2004). Antimicrobial Activity of Spices. *Journal of Rapid Methods and Automation in Microbiology*, 12: 1- 55.
- Channya F. K. and Chimbekujwo I. B. (2002). Pathogens of Post-harvest Fruits Rot of Plantain (*Musa Parasidiaca* L.) in South- Western Nigeria. *Journal of Tropical Bioscience*, 21-24.
- Channya, F. K. (1991). Fungi Associated with Post-harvest Rot of Plantains (*Musa parasidiaca* linn) in South-western Nigeria. M.Sc. Thesis, Department of Botany/ Microbiology, University of Ibadan, Ibadan.
- ChihCheng, T. C. and Robert R. K. (2007). The Date Palm (*Phoenix dactylifera* L.): Overview of Biology, Uses, and Cultivation.
- Christensen, M. J., Folloan, R. E. and Skip, R. A. (2007). Plant Pathology. *Australian Plant Pathology*, 17(2): 45-47.
- Chukwuka, K. S., Okonko, I. O. and Adekunle, A. A. (2010) Microbial Ecology of Organisms Causing Pawpaw (*Carica papaya* L.) Fruit Decay in Oyo State, Nigeria. *American-Eurasian Journal of Toxicological Sciences*, 2 (1): 43-50.
- Conner D. E. and Beuchat L. R. (1984). Effect of Essential Oils from Plants on Growth of Food Spoilage Yeasts. *Journal of Food Science*, 49:429-434.
- Cristani M., d'Arrigo M., Mandalari G., Castelli F., Sarpietro M. G., Micieli D., Venuti V., Bisignano G., Saija A. and Trombetta D. (2007). Interaction of Four Monoterpenes Contained in Essential Oils with Model Membranes: Implications for their antibacterial activity. *Journal of Agricultural and Food Chemistry*, 55:6300-6308.
- Curtis H., Noll U., Störmann J. and Slusarenko A. J. (2004). Broad-spectrum Activity of the Volatile Phytoanticipin Allicin in Extracts of Garlic (*Allium sativum* L) Against Plant Pathogenic Bacteria, Fungi and Oomycetes. *Physiology and Molecular Plant Pathology*, 65:79-89.
- Daferera D. J., Ziogas B. N. and Polissiou M. G. (2000). GC–MS Analysis of Essential Oils from Some Greek Aromatic Plants and their Fungitoxicity on *Penicillium digitatum*. *Journal of Agricultural and Food Chemistry*, 48:2576–2581.
- El-Sherbeeny M. R., Saddik M. F. and Bryan F. L. (1985). Microbial Profiles of Foods Served by Street Vendors in Egypt. *International Journal of Food Microbiology*, 2: 355-364.
- FAO, (2012). Plant Production and Protection Paper No. 35. Date Production and Protection. FAO. Rome, 198-294.
- Fawole, M. O. and Oso, B .A. (1995). Laboratory Manual of Microbiology. 1st Edition. *Spectrum Books Ltd*, Ibadan, Nigeria, 34 - 35.
- Glasner, B., Botes, A., Zaid, A. and Emmens, J. (2002). In *Date Palm Cultivation*, Date Harvesting, Packinghouse Management and Marketing Aspects, ed Zaid A. (Food and Agriculture Organization Plant Production and Protection paper no. 156. *Food and Agriculture Organization of the United Nations, Rome, Italy*), 177–208.
- Grozav M. and Foarce A. (2005). Preliminary Study on the Biological Activity of *Allium sativum* Essential Oil as Potential Plant Growth Regulators. 3rd International Conference of Seed Pathology. Bydgoszcz, Poland, 6th- 8th September 2006. Abstracts, 87p. Electron. *Journal of Environment and Agricultural Food Chemistry*, 4(6):1138-1142.
- Hadizadeh, I., Peivastegan, B. and Hamzehzarghani, H. (2009). Antifungal Activity of Essential Oils from Some Medicinal Plants of Iran against *Alternaria alternata*. *American Journal Applied Science*, 6(5): 857-861.
- Hamad, S. H. (2008). Microbial Spoilage of Date Rutab Collected from the Markets of Al-Hofuf City in the Kingdom of Saudi Arabia. *Journal of Food Protection*, 71: 1406-1411.
- Harrak, H. (2003). Activités de Recherche Matière de la Valorisation Technologique des Dattes, in Rapport d'activités Compagne 2002-2003, INRA, Centre Régional de la Recherche Agronomique de Marrakech, Morocco.
- Harrak, H. and Chetto, A. (2002). Valorisation et Commercialisation des dattes au Maroc. *Edition INRA, Maroc*, 222.

- Hassan M. M., Chowdhury S. P., Alam S., Hossain B. and Alam M. S. (2005). Antifungal Effects of Plant Extracts on Seed-borne Fungi of Wheat Seed Regarding Seed Germination, Seedling Health and Vigour Index. *Pakistan Journal of Biological Science*, 8(9): 1284-1289.
- Hayrettin, O., Hatice I. O. B. G. and Guner O. (2012). Mycotoxin Risks and Toxigenic Fungi in Date, Prune and Dried Apricot among Mediterranean Crops; *Phytopathologia Mediterranea*, 51: 148–157.
- Howard, F. W., Moore, D., Giblin-Davis, R. M., and Abad, R. G. (2001). Insects on Palms (*CABI International*, Wallingford, Oxon, UK).
- Hunter B. B. and Barnett, H. I. (1998). *Illustrated genera of imperfect fungi*, (4th ed.). Minnesota. *American Phytopathological Society*.
- Josling P. (2003). Allicin. The earth of garlic. NWI Publishing Callahan. Florida pp.141-149.
- Kader, A. A. (2007). Recommendations for Maintaining Postharvest Quality. Department of Plant Science, University of California, Davis. Available at: <http://postharvest.ucdavis.edu/ProduceFacts/Fruit/Dates.shtml>.
- Kader, A. A. and Hussein, A. (2009). Harvesting and Postharvest of Dates. *ICARDA, Aleppo, Syria. iv +, 15*.
- Khalaf A., Emad I. H., Khalid M. A., Mahmoud A., Wesam A. K., Jacob H. J., Mohamad A. S., Ashraf K. and Mohamed I. H. (2011). Identification and Controlling *Pythium* sp. Infecting Tomato Seedlings Cultivated in Jordan Valley using Garlic Extract. *Asian Journal of Plant Pathology*, 5:84-92.
- Khomutov, R., Dzhevakhya, V., Khurs, E., Ospova, T., Shcherbakova, L., Zhemchuzhina, N., Mikityuk, O. and Nazarova, T. (2011). Chemical Regulation of Mycotoxin Biosynthesis. *Doklady Biochemistry and Biophysics*, 1:25-28.
- Klick, M. A. and Pitt. J. I. (1988). A Laboratory Guide to Common *Aspergillus* species and their Teleomorphs. Published by *Commonwealth Scientific and Industrial Research Organisation*, Division of Food Processing. North Ryde, NSW Australia, 116.
- Lucini E. I., Zunino M. P., Lopez M. L. and Zygodlo J. A. (2006). Effect of Monoterpenes on Lipid Composition and Sclerotial Development of *Sclerotium cepivorum* Berk. *Journal of Phytopathology*, 154:441–446.
- Mironescua, M. and Georgescu, C. (2008). Preliminary Researches on the Effect of Essential Oils on Moulds Isolated from Surfaces. *Journal of Agroalimentary Processes and Technologies*, 14: 30-33.
- Omidbeygi M., Barzegar M., Hamidi Z. and Naghdibadi H. (2007). Antifungal Activity of Thyme, Summer Savory and Clove Essential Oils Against *Aspergillus flavus* in Liquid Medium and Tomato Paste. *Food Control*, 18:1518–1523.
- Raja, G. R., Nirmala, C. R. and Ramanamma, C. H. (2009). Efficacy of Phytoextracts and Oils of Certain Medicinal Plants against *Cercospora moricola* Cooke, Incitant of Mulberry (*Morus alba* L.) leaf spot. *Journal of Biopesticides*, 2(1): 77-83.
- Robert, A. S., Hoekstra, Frisvad, J. C. and Filtenborg, O. (1996). Introduction to Food-borne Fungi. Printed by *Ponsen and Looyen, Wageningen, The Netherlands*.
- Rodrigues E. L. and Bach E. E. (2003). Alicina Como Elicitor de Resistência na Cultivar de Cevada AF 94135. In: XXIII Reuniao Anual de Pesquisa de cevada, 2003, Passo Fundo: EMBRAPA pp. 557-570.
- Rodrigues E. L., Milanes, and Bach E. E. (2002). Utilização da Alicina Como Elicitor de Resistência em Plantas de Cevada (variedade EMBRAPA 128) Contra *Bipolaris sorokiniana*. In: XXII Reuniao Anual de Pesquisa de cevada, 2002, Passo Fundo, EMBRAPA pp.519-530.
- Sharma N. and Tripathi A. (2006). Effects of *Citrus sinensis* (L.) Osbeck epecarp Essential Oil on Growth and Morphogenesis of *Aspergillus niger* (L.) Van Tieghem. *Microbiological Research*, 163:337-344.
- Silva A. A. O., Rodrigues E., Antomiazzi N., Milanez A. and Bach E. E. (2001). Allicin Effect for Control *Bipolaris sorokiniana* in Barley. *Summa Phytopathology*, 27: 95.
- Singh, R. Y., Kumar, S. and Dikshit, A. (2006). Antifungal Properties of Essential Oil of *Menthaspicata* L. var. MSS-5 Ramesh Singh Yadav. *Indian Journal Crop Science*, 1(1-2): 197-200.
- Slusarenko A., Patel A. and Portz D. (2008). Control to Plant Diseases by Natural products: Allicin from garlic as a case study. *Eur. Plant Pathology*, 121:313-322.
- Spotts, R. A. and Chen, P. M. (1987). Post-harvest Teat Treatment for Control Decay of Pear Fruit. *Phytopathology*, 67: 1578-1582.
- Suwitchayanon, P. and Kunasakdakul, K. (2009). *In vitro* Effects of Clove and Turmeric Extracts Controlling Crucifer Pathogens. *Journal Agricultural Technology*, 5(1): 193-199.
- Terezinha, J. F., Ferreira, R. S., Yassumoto, L. and Roberto, J. (2006). Antifungal Activity of Essential Oil Isolated from *Ocimum gratissimum* L. (eugenol chemotype) against Phytopathogenic Fungi. *Brazilian Archives of Biology and Technology*, 49: 867-871.
- UNEP (1995). Montreal Protocol on Substance that Depletes the Ozone Layer. *Methyl Bromide Technical Option Committee Kenya*, 304.
- Vasile B. R., Vlaicu B. and Butnariu M. (2012). Chemical Composition and *in Vitro* Antifungal Activity Screening of the *Allium ursinum* L. (Liliaceae). *International Journal of Molecular Science*, 13: 1426-

- 1436.
- Viuda-Martos M., Ruiz-Navajas Y., Fernández-López J. and Pérez-Álvarez J. (2008). Antifungal Activity of Lemon (*Citrus lemon* L.), Mandarin (*Citrus reticulata* L.), Grapefruit (*Citrus paradisi* L.) and Orange (*Citrus sinensis* L.) Essential Oils. *Food Control*, 19:1130-1138.
- Vyawahare, N., Pujari, R., Khsirsagar, A., Ingawale, D., Patil, M. and Kagathara, V. (2009). *Phoenix dactylifera*: An Update of its Indegenous Uses Photochemistry and Pharmacology. *International Journal of Pharmacology*, 7(1): 1531-2976.
- Wazir, A. M., Ghulman, S. M., Abul-soad, A. A., Abdulmubeen, L. and Mushtaque A. J. (2013). Chemical Control of Sudden Decline Disease of Date Palm (*Phoenix dactylifera* L.) in Sindh, Pakistan. *Pakistan Journal Botany*, 45 (s1): 7-11.
- WHO (1999). World Health Organization. Basic Food Safety for Health Workers.
- Wikipedia, (2014). Date Palm. http://en.Wikipedia.Org/wiki/date_palm.
- Zaika L. L. (1988). Spices and Herbs: Their antimicrobial activity and its determination. *Journal of Food Safety*, 9(2): 97–118.