

# Review on Effect of aluminum toxicity and genetic control mechanisms in crops

Tegegn Belete

E-mail address:tegegnbelete2011@gmail.com

Ethiopian Institute of Agricultural Research; Jimma Agricultural Research Center

P.O. Box: 192; Jimma, Ethiopia

## ABSTRACT

*Aluminum (Al) toxicity is one of the major factors constraining crop production on 67% of the total acid soil area in the world. Al toxicity restricts root growth and affects nutrient and water absorption with resultant stunted growth and reduced grain and biomass yield of crops. Cereals occupy about half of the world's cropland area and, therefore, take a lion share of the global Al-toxicity constraint. Al-toxicity is more serious in tropical environments, where the soil is highly resistant to improvement by lime application. In addition, in these environments, the use of adequate lime and organic fertilizer sources is constrained by various technological and socio-economic constraints. Studies on genetic control of Al toxicity are active areas of research for most of the globally important cereals. Development and use of Al-tolerant crop varieties is economically feasible and an environmentally friendly management option that can complement other non-genetic management options. This paper introduces the importance of soil acidity and effect of Al-toxicity on plant growth and development and yield.*

**Key word:** Aluminum toxicity, genetic control, crops, soil acidity

**DOI:** 10.7176/JBAH/14-2-01

**Publication date:** July 31<sup>st</sup> 2024

## 1. Introduction

Soil acidity is one of the most important factors that affect crop production worldwide. Acid soils (pH < 5.5 in surface layer) constitute 3,950 million ha or 30% of the world's total ice free land or about 40% of the arable land (Von Uexkull and Mutert, 1995). In Africa 22% or 659 million ha of the total 3.01 billion ha land area has soil acidity problem (Von Uexkull and Mutert, 1995). Al-toxicity is the single most important contributing factor constraining crop production on 67% of the total acid soil area in the world (Eswaran *et al.*, 1997). Aluminum is the most abundant metal and the third most common element in the earth's crust (Delhaize and Ryan, 1995; Vitorello *et al.*, 2005). In soils, it mostly exists as structural constituent of primary and secondary minerals especially of the aluminosilicates. Nonetheless, despite its abundance, Al is not known to be used in any living organisms (Vitorello *et al.*, 2005).

As the soil gets acidic, the silicon will be leached leaving aluminum in the solid forms as aluminum oxyhydroxides, such as boehmite and gibbsite. These forms release the phytotoxic aluminum species,  $Al^{+3}$  also known as  $Al(H_2O)^{+3}_6$  into the soil solution (Abebe, 2007). Even though, there are several forms of Aluminum species in the soil,  $Al^{+3}$  and monomeric Al-hydroxyl species ( $AlOH^{+2}$  and  $Al(OH)^{2+}$ ) are the most phytotoxic ones (Miyasaka *et al.*, 2007). The trivalent  $Al^{+3}$  is dominant in soil solutions when the soil pH is less than 5. The most common and immediate toxic effect of  $Al^{+3}$  in plants is inhibition of root growth which happens within few hours after exposure to micro molar concentrations of Al (Barcelo and Poschenrieder, 2002).

Root inhibition could be exhibited on primary and lateral root apices, and such roots become thick and develop brown colour (Vitorello *et al.*, 2005; Wang *et al.*, 2006). Suppression and abnormal root morphology directly hinders nutrient uptake as well as water absorption. Consequently, plants show stunted growth and become susceptible to drought (Vitorello *et al.*, 2005; Wang *et al.*, 2006; Miyasaka *et al.*, 2007). The yield loss associated with Al-toxicity varies depending on soil Al saturation, the crop species and the specific variety used. For instance, Al-tolerant maize variety gave 61% higher grain yield than Al-sensitive variety, and with lime treatment, yield increment of 208 and 82% was obtained for Al sensitive and Al-tolerant varieties of maize, respectively (The *et al.*, 2006).

Applications of lime, manure compost, and use of tolerant crop species or varieties are the most common methods used to overcome the impact of Al-toxicity. Nevertheless, in the context of tropical Africa, utilization of lime, manure and other organic fertilizer sources have their own technical and or socio economic constraints. Lime has been extensively used to ameliorate acid soils of temperate areas. In these areas, soil acidity develops

mainly as a consequence of heavy use of chemical fertilizers and environmental pollution (Rao *et al.*,1993).In the tropics,several experimental reports also indicate significant yield increment with application of lime (The *et al.*,2006).However,the highly acidic soils of this region have strong buffering capacity against amendment by lime.Such soils demand heavy dose and need deeper incorporation to ameliorate the sub surface acidity.

Most of resource poor farmers in the tropics,however,are constrained by unavailability ,transport and high cost of this bulky dose (Rao *et al.*,1993).In addition,since lime incorporation to the subsoil is hardly possible,even when surface soil is neutralized, difficulty of ameliorating the subsoil restricts root growth of plants to surface soil and make them vulnerable to drought (Little,1989;Foy,1992).Runoff pollution and adverse effects of lime on rotation crops are also other side effects of lime application (Wang *et al.*,2006).Use of organic matter seems an applicable strategy to resource poor farmers of the tropics who cannot afford purchase of large volume of lime and fertilizers.However,regular and high volume application of manure and compost to the highly acidic soils is limited by competing uses of organic matter sources for fuel,animal feed and construction (Schlede,1989).On contrary,in the tropics,the use of acid forming fertilizers on cultivated land and expansion of crop production to forest inhabited areas accelerate development of the soil acidity and Al-toxicity (Giller *et al.*,1996).

Cereals,the predominant staple food crops of the world population,are cultivated roughly on the half of the world's crop land (Dyson,1999).By the year 2025,the world's farmers are expected to produce about 3 billion tons of cereals to feed the human population of the around 8 billion, and this requires an average world cereal yield of about 4 metric tons/ha (Dyson,1999).The current average cereal yields in Africa is below 1 ton per ha (Langyintuo,2011).The use of tolerant crop varieties is considered to be the best complement to non-genetic management option for combating Al-toxicity problem (Rao *et al.*,1993).This paper attempts to review the effect of Al-toxicity and genetic control mechanism in crops.The reviewer believe that the paper gives better insight to basic information on Al-toxicity and genetic control in crops.

## **2. Literature Review**

### **2.1 Effect of Al toxicity on plant growth and development**

#### **2.1.1 Aluminum Toxicity**

##### **2.1.1.1 Effects on leaves**

Aluminum toxicity is a potential growth-limiting factor for plants grown in acid soils in many parts of the world (Foy,1974).The symptoms of aluminum toxicity are not easily identifiable. In plants,the foliar symptoms resemble those of phosphorus (P) deficiency (overall stunting,small,dark green leaves and late maturity,purpling of stems,leaves,and leaf veins,yellowing and death of leaf tips).In some cases,Al toxicity appears as an induced Calcium (Ca) deficiency or reduced Ca transport problem (curling or rolling of young leaves and collapse of growing points or petioles).Excess Al even induces iron (Fe) deficiency symptoms in rice (*Oryza sativa* L.),sorghum and wheat (Clark *et al.*,1981).

##### **2.1.1.2 Effects on roots**

Aluminum does not affect the seed germination but helps in new root development and seedling establishment (Nosko *et al.*, 1988).Root growth inhibition was detected 2-4 days after the initiation of seed germination (Bennet *et al.*,1991).Vanpraag and Weissen *et al.*,(1985) reported that plant species and ecotypes growing on acid soils had become very resistant to the inhibitory effects of aluminum on root absorption and growth in course time and phenological evolution. The major Al toxicity symptom observed in plants is inhibition of root growth (Bennet *et al.*,1991).The roots exhibit greater signs of cellular damage than other parts of the plant (Rincon and Gonzales,1992).Al toxicity could be observed in the root system particularly in root-tips and in lateral roots; lateral roots become thickened and turn brown (Kinraide,1985).The root system as a whole is corraloid in appearance with many stubby lateral roots but lacks fine branching (Foy *et al.*,1978).The toxicity appears to be determined by the availability of certain monomeric species of Al to the plant roots (Bartlett and Reigo,1972).

Losses of phytoactive,monomeric Al can occur by polymerization of Al as the p<sup>H</sup> and the Al concentration rise (8) to make complex formation or chelation with phosphate and organic acids (Bartlett and Reigo,1972).Kinraide *et al.*,(1985) demonstrated rapid assay for aluminum phytotoxicity at sub micromolar concentration of Al to *Trifolium pretense*.Wagatsuma *et al.*,(1987) noted the role of aluminum on root cells of various crops.They reported that the cells of the epidermis and outer cortex of maize (Al-sensetive) in the portion approximately

1cm from the root–tip were damaged and the walls of these cells were abnormal and partially detached in barley (a plant highly sensitive to Al);more pronounced abnormality and detachment of the cell walls involved almost the whole cortex, and few cortex cells remained alive in oats (Al-tolerant) after 6 days' exposure to the Al treatment.

Aluminum was absorbed in large amounts in the tip portion of the root.In the tip portion, the K content decreased with the increase of the Al content,but the Ca content was almost constant.Bennet *et al.*,(1985) reported that an anisotropic growth response of cortical cells with 20-h root exposure to Al were associated with the collapse of the conducting tissue of the stele and disintegration of the outer cells of the root.

### 2.1.1.3 Effects on plant morphology and physiology

Aluminum is one of the most abundant elements in the earth's crust,and toxic for many plants when the concentration is greater than 2-3 ppm with a soil  $p^H < 5.5$  (Balsberg,1990).A significant correlation between low  $p^H$  and high Al concentration has also been shown in acidified freshwater, where this metal may reach levels of 0.3-1.6 mM (Dickson,1978) and cause serious metabolic derangement in some hydrophytes (150).In general,young seedlings are more susceptible to Al than older plants (Thawornwong and Diest,1974).So far as physiology is concerned.

Al has been shown to interfere with cell division in plant roots;fix phosphorus in less available forms in the soil and in or on plant roots; decrease root respiration; interfere with certain enzymes governing the deposition of polysaccharides in cell walls; increase cell wall rigidity (cross-linking pectins) and interfere with the uptake, transport and with some essential nutrients (Ca,Mg,K,P) and water supply to plants (Fleming *et al.*,1974); alters cell-wall Donnan free space (450,the plasma membrane transport proteins (Caldwell,1989) and regulates the activity of many enzymes (Copeland and DeLima,1992) and metabolic pathway for repair mechanism (Plucinska and Ziegler,1995).Trim (1985) reported that Al is known to form strong complexes to precipitate nucleic acids.Soileau and Engelstad (1969) and Soileau *et al.*,(1969) indicated that chemical factors were more important than physical factors in limiting cotton root growth in an acid ( $p^H 4.4$ ) fragipan soil.

Al becomes soluble or exchangeable and also toxic depending on the soil  $p^H$  and many other factors including the predominant clay minerals,organic matter levels,concentrations of other cations,anions and total salts,and the plant species(Foy,1984).Dickson (1978) reported that there was a significant correlation between low  $p^H$  and high aluminum concentration in fresh water, and metal may reach levels of 0.3 – 1.6 mM.It also causes serious metabolic derangement in some hydrophytes(Plieth *et al.*,1985).Berggren and Fiskessjo (1987) reported aluminum toxicity in *Allium cepa* with reference to root growth and morphology.Further,Severi (1991) analyzed the aluminum toxicity in *Lemna minor* with reference to citrate and cytokinin metabolism.Physiological mechanisms due to Al toxicity have been focused on field crops and other herbaceous plants (Foy *et al.*,1978).Plieth *et al.*,(1999) reported that low pH elevation in cytosolic calcium were inhibited by aluminum toxicity.They observed that plant roots responded to external low  $p^H$  by a sustained elevation in cytosolic free calcium concentration  $Ca^{+2}$  (C) in the presence of aluminum.They also suggested that a primary toxic effect of aluminum might impair calcium- mediated plant defence responses against low  $p^H$ .

### 2.1.3 Factors affecting Al- toxicity

Al toxicity is affected by many factors such as  $p^H$ ,concentration of Al,temperature, and concentration of cations and anions in culture solution.A  $p^H 5.0$  or above will reduce Al solubility (Reid *et al.*,1971) thus reducing Al toxicity.Root elongation depended critically on the concentration of  $Ca^{+2}$ ,whether in the presence or absence of Al, with at least 0.2mmol/L  $Ca^{+2}$  being essential for optimum growth (Kinraide *et al.*,1985).The concentration of  $Ca^{+2}$  greatly influences the Al toxicity at a given pH and Al concentration. As the  $Ca^{+2}$  concentration approached 1mmol/L the inhibition by 1micromol/L Al was nearly eliminated (Kinraide *et al.*,1985).Increased concentrations of basic cations in solution of the root rhizoshere,particularly calcium have been shown to ameliorate Al toxicity (Brady *et al.*,1993). $Mg^{+2}$  at concentration of 0.5mmol/L can also alleviate Al toxicity as did  $Ca^{+2}$  (Kinraide *et al.*,1985).Application of  $NH_4Cl$  to a soil with a high exchangeable Al significantly reduced barley seedling emergence,shoot and root weights,spike numbers/ $m^2$  and grain numbers/spike whereas  $NaNO_3$  significantly increased all these parameters. At harvesting, soil analysis showed that  $NH_4Cl$  significantly reduced soil pH and increased soil Al and Mn contents and this was confirmed by tissue analysis of shoots and roots (Stange *et al.*,1995). $NH_4^+-N$  induced release of  $H^+$  from the roots particularly whilst  $NO_3^- -N$  significantly increased pH by release of  $OH^-$  (Borie *et al.*,1994).Adding excess P in nutrient solution will precipitate and detoxify Al (Kinraide *et al.*,1985).

## 2.2 Genetic control of Al-tolerance in crop plants

Studies on genetic control of Al toxicity are active areas of research for most of the globally important cereals. In wheat, earlier reports presumed that Al toxicity in wheat is controlled at least by two major loci (Didier *et al.*, 1996). The two genes proposed were genes that encode for malate, and phosphate exudation to the rhizosphere (Didier *et al.*, 1996). A major aluminum tolerance gene in wheat, ALMT1 later renamed as TaTAALM1, is known to confer an Al-activated efflux of malate from root apices (Sasaki *et al.*, 2004). This gene is mapped to chromosome 4DL using 'Chinese spring' deletion lines. Absence or loss of this gene resulted in loss of Al-tolerance and malate exudation (Raman *et al.*, 2005a). Hence, it was suggested that Al-tolerance in diverse range of wheat genotypes to be primarily controlled by TaALMT1 located at Alt<sub>BH</sub> (Raman *et al.*, 2005a).

Very recently, with discovery of a new mechanism of Al-tolerance that involve efflux of citrate in root apices of Brazilian wheat cultivars, another gene that resides on chromosome 4BL has been identified (Ryan *et al.*, 2009). They also indicated that the citrate efflux is controlled by single gene which could explain 50% of the phenotypic variation in citrate efflux. In addition, Navacode *et al.*, (2009) located two major Al-tolerance QTL on chromosome arm 4DL and 3BL which could, respectively explain 49 and 31% of the phenotypic variance present in the population of 'Chinese Spring' wheat cultivar. These findings indicated that the trait is controlled by major and minor genes in wheat. In Barley, Lima Echart *et al.* (2002) indicated that the F<sub>2</sub> generation analysed with haematoxylin staining followed the Mendel's segregation ratio 3:1 for Al toxicity tolerant to susceptible plants; revealing the fact that the trait is controlled by single dominant gene. It is generally agreed that Al tolerance in barley is conditioned by the *Alp* locus which is located on the long arm of chromosome 4H. This locus is associated with Al-induced efflux of citrate from root apices of tolerant barley varieties (Wang *et al.*, 2006). A gene encoding a multidrug and toxic compound extrusion protein is proposed as a candidate gene for Al-tolerance in Barley (Wang *et al.*, 2007). In addition, quantitative trait loci that could explain 50% of the phenotypic variation are also associated with the same chromosomal location (Jian Feng *et al.*, 2004).

Similarly, Raman *et al.*, (2005b) identified quantitative trait loci for root elongation under aluminum stress on 3H, 4H, 5H and 6H chromosomal locations. Alike other cereals, aluminum tolerance in rye is effected by efflux of organic acids. Segregation ratio of 3:1 (tolerant to sensitive) was found in three F<sub>2</sub> populations analysed indicating the fact that the trait is controlled by single dominant locus (Matos *et al.*, 2005). So far, four independent loci Alt<sub>1</sub>, Alt<sub>2</sub>, Alt<sub>3</sub> and Alt<sub>4</sub> located on chromosome arms 6RS, 3RS, 4RL and 7RS, are known to confer Aluminum toxicity tolerance in this crop (Matos *et al.*, 2007). Specifically, the Alt<sub>4</sub> locus contains cluster of genes homologues to the single copy Al-activated malate transporter (TaALMT1) (Collins *et al.*, 2008).

Tolerant and sensitive rye genotypes contain five and two genes of the clusters at the locus, respectively. Out of these, two ScALMT1-M39.2 and one ScALMT1-M77 genes are highly expressed in the root tip (Collins *et al.*, 2008). In rice, root growth under Al stressed condition is controlled by several quantitative trait loci (QTLs) genes. Two-three QTLs of largest effect, however, are identified to explain phenotypic variation for Al-tolerance (Ma *et al.*, 2002a). A recent study identified two genes STAR1 and STAR2 which function as bacterial-type ATP binding cassette (ABC) transporter to control Al-tolerance in rice (Hang *et al.*, 2009). The mechanism, however, is not yet clear enough.

## 3. CONCLUSION

Aluminum toxicity is an important growth-limiting factor for plants in many acid soils, particularly in PH of 5.0 or below. Aluminum toxicity in plants is often clearly identifiable through morphological and physiological symptoms. Differential tolerances to Al toxicity almost certainly involves differences in the structure and function of roots. Aluminum interferes with cell division in roots, decreases root respiration and uptake and use of water and nutrients, particularly calcium and phosphorus and metabolic pathway. Other promising approaches to studying metal toxicity in tolerant and sensitive plant genotypes are to determine the metal uptake and transportation in various plant parts, the mechanisms behind the interaction with mineral nutrients, specific genes responsible for tolerance, levels and kinds of organic and amino acids which act as metal chelators and detoxifiers, level and forms of enzymes and changes in root permeabilities to ions and molecules and its mechanisms.

## 4. REFERENCES

Abebe M (2007). Nature and Management of Acid Soils in Ethiopia. Ethiopian Institute of Agricultural Research, Addis Ababa, Ethiopia.

- Balsberg Pahlsson A.M., Influence of aluminum on biomass, nutrients, soluble carbohydrate and phenols in beech (*Fagus sylvatica*), *Physiol. Plant.* 78 (1990) 79–84.
- Barcelo J, Poschenrieder C (2002). Fast root growth responses, root exudates, and internal detoxification as clues to the mechanisms of Al toxicity resistance : a review. *J. Environ. Exp. Bot.*, 48:75-92.
- Bartlett R.J., Riego D.C., Effect of chelation on the toxicity of aluminium, *Plant and Soil* 37 (1972) 419–423.
- Bennet R.J., Breen C.M., Fey M.V., The aluminium signal: new dimensions of aluminum tolerance, *Plant and Soil* 134 (1991) 153–166.
- Bennet R.J., Breen C.M., Fey M.V., Aluminium-induced changes in the morphology of the quiescent centre, proximal meristem and growth region of the root of *Zea mays*, *S. Afr. Tydskr. Planik.* 51 (1985) 355–362
- Berggren D., Fiskesjö G., Aluminium toxicity and speciation in soil liquids-experiments with *Allium cepa* L., *Environ. Toxicol. Chem.* 6 (1987) 771–779.
- Brady, D.J., Edwards, D.G., Asher, C.J., Blamey, F.C.P., 1993. Calcium amelioration of aluminium toxicity effects on root hair development in soybean (*Glycine max* L.) *Merr. New Phytol.*, 123(3):531-538. [doi:10.1111/j.1469-8137.1993.tb03765.x]
- Borie, B.F., Stange, J.B., Morales, L.A., Pino, B.M., 1994. Effect of aluminium and acidity on root growth in barley (*Hordeum vulgare* L.) and oats (*Avena sativa* L.). *Agricultura Tecnica (Santiago)*, 54:224-230.
- Caldwell C.R., Analysis of aluminum and divalent cation binding to wheat root plasma membrane proteins using terbicum phosphorescence, *Plant Physiol.* 91 (1989) 233–241.
- Collins NC, Shirley NJ, Saeed M, Pallotta M, Gustafson JP (2008). An *LMT1* Gene Cluster Controlling Aluminum Tolerance at the *Alt4 A* Locus of Rye (*Secale cereale* L.). *Genetics.* 179: 669-682.
- Clark R.B., Pier H.A., Knudsen D., Maranville J.W., Effect of trace element deficiencies and excesses on mineral nutrients in sorghum, *J. Plant Nutr.* 3 (1981) 357–374.
- Copeland L., DeLima M.L., The effect of aluminium on enzyme activities in wheat roots, *J. Plant Physiol.* 140 (1992) 641–645.
- Delhaize E, Ryan PR (1995). Aluminum toxicity and tolerance in plants. *J Plant Physiol.*, 107: 31 35-321
- Dickson W., Some effects of the acidification of Swedish lakes, *Verh. Int. Verein. Limnol.* 20 (1978) 851–856.
- Didier MP, Lisa AP, V. KL (1996). Multiple aluminium-resistance mechanism in wheat: roles of root apical phosphate and malate exudation. *Plant Physiol.*, 112: 591-597.
- Dyson T (1999). World food trends and prospects to 2025. The National Academy of Sciences colloquium „Plants and Population: Is There Time?“, Arnold and Mabel Beckman Center in Irvine, CA., pp. 5929-5936
- Eswaran H, Reich P, Beinroth F (1997). Global distribution of soils with acidity. In: A.Z. Moniz AMCF, R.E. Schaffert, N.K. Fageria, C.A. Rosolem and H. Cantarella (eds) *Plant-Soil Interactions at Low pH*. Brazilian Soil Science Society, pp.159-164.
- Fleming A.L., Schwartz J.W., Foy C.D., Soil: Aluminium toxicity in plants, *Agron. J.* 66 (1974) 715–719.
- Foy C.D., Effect of aluminium on plant growth, in: Carson E.W. (Ed.), *The Plant Root and its Environment*, Charlottesville, Univ. Press, Virginia, 1974, pp. 601–642.
- Foy CD (1992) Soil chemical factors limiting plant root growth. *Adv. Soil Sci.* 19: 97-148.
- Foy C.D., Chaney R.L., White M.C., The physiology of metal toxicity in plants, *Annu. Rev. Plant Physiol.* 29 (1978) 511–566.

- Foy C.D., Physiological effects of hydrogen, aluminium and manganese toxicities in acid soils, in: Adams F. (Ed.), *Soil Acidity and Limiting*, Second Edition, Amer. Soc. Agron., Madison, Wisconsin, 1984, pp. 57–97
- Giller KE, . Cadisch G, Ehaliotis C, Adams E, Sakala WD, Mafongoya PL (1996) Building soil nitrogen capital in Africa. *In: Buresh RJ, P. A. Sanchez, Calhoun F (eds) Replenishing Soil Fertility in Africa: Proceedings of an international symposium, Indianapolis, Indiana. 6 November 1996. Soil Science Society of America (SSSA), Madison, Wisconsin, USA, pp. 151-192.*
- Huang CF, Yamaji N, Mitani, N, Yano M, Nagamura Y, Maa JF (2009). A Bacterial-type ABC transporter is involved in aluminum tolerance in rice. *Plant Cell* 21:655-667.
- Jian Feng M, Sakiko N, Kazuhiro S, Hiroyuki I, Jun F, Kazuyoshi T (2004) Molecular mapping of a gene responsible for Al-activated secretion of citrate in barley. *J. Exp. Bot.*, 55:1335-1341.
- Kinraide T.B., Proton extrusion by wheat roots exhibiting severe aluminium toxicity symptoms, *Plant Physiol.* 88 (1988) 418–423.
- Kinraide T.B., Arnold R.C., Baligar V.C., A rapid assay for aluminium phytotoxicity at submicromolar concentrations, *Physiol. Plant.* 65 (1985) 245–250.
- Kinraide, T.B., Arnold, R.C., Baligar, V.C., 1985. A rapid assay for aluminium phytotoxicity at submicromolar concentrations. *Physiol. Plant.*, 65(3):245-250. [doi:10. 1111/j.1399-3054.1985.tb02390.x].
- Langyintuo A (2011). African agriculture and productivity. Sharing knowledge across the Mediterranean (6) Conference, Villa Bighi, Malta.
- Little R (1989) A review of breeding wheat for tolerance to Aluminium toxicity In: Van Ginkel M, Tanner DG (eds) Fifth Regional Wheat Workshop for Eastern, Central, and Southern Africa and the Indian Ocean. CIMMYT, Antsirabe, Madagascar, October 5-10, 1987, pp.83-97.
- Lima Echart C, Fernandes Barbosa-Neto J, Garvin D, Cavalli-Molina S (2002). Aluminum tolerance in barley: Methods for screening and genetic analysis. *Euphytica.* 126: 309-313.
- Matos M, Pérez-Flores V, Camacho M, Pernaute B, Pinto-Carnide O, Benito C (2007). Detection and mapping of SSRs in rye ESTs from aluminium-stressed roots. *Molecular Breed.* 20:103-115
- Matos M, Camacho MV, Pérez-Flores V, Pernaute B, Pinto-Carnide O, Benito C (2005). A new aluminum tolerance gene located on rye chromosome arm 7RS. *Theor. Appl. Genet.*, 111:360-369
- Ma JF, Shen R, Zhao Z, Wissuwa M, Takeuchi Y, Ebitani T, Yano M (2002a). Response of rice to Al stress and identification of quantitative trait loci for Al tolerance. *Plant Cell Physiol.*, 43:652-659.
- Miyasaka SC, Hue NV, Dunn MA (2007) Aluminum. In: Barker AV, Pilbeam DJ (eds) *Handbook of Plant Nutrition*. Taylor and Francis Group, Boca Raton, pp. 439-497.
- Navakode S, Weidner A, Lohwasser U, Röder M, Börner A (2009). Molecular mapping of quantitative trait loci (QTLs) controlling aluminium tolerance in bread wheat. *Euphytica* 166:283-290.
- Nosko P., Brassard P., Kramer J.R., Kershaw K.A., The effect of aluminium on seed germination and early seedling establishment growth and respiration of white spruce (*Picea glauca*), *Can. J. Bot.* 66 (1988) 2305–2310.
- Plucinska G.L., Ziegler H., The effect of aluminium on adenylate levels in Scots pine roots, *Acta Physiol. Plant.* 17 (1995) 225–232.
- Pettersson A., Hallbom L., Bergman B., Physiological and structural responses of the cyanobacterium *Anabaena cylindrica* to aluminium, *Physiol. Plant.* 63 (1985)153–158.
- Raman H, Wang JP, Read B, Zhou MX, Venkataganappa S, Moroni JS, O'Bree B, Mendham N (2005b) Molecular mapping of resistance to aluminium toxicity in barley. *Proceedings of Plant and Animal Genome XIII Conference, San Diego, USA, p. 154*

Raman H, Kerong Z, Mehmet C, Rudi A, David FG, Lyza GM, Kochian LV, J. Sergio M, Rosy R, Muhammad I, Fiona Drake-Brockman, Irene Waters, Peter M, Takayuki S, Yoko Y, Hideaki M, Diane MH, Emmanuel D, Peter RR (2005a). Molecular characterization and mapping of ALMT1, the aluminium-tolerance gene of bread wheat (*Triticum aestivum* L.). *Genome*, 48:781–791

Rao IM, Zeigler RS, Vera R, Sarkarung S (1993) Selection and breeding for acid-soil tolerance in crops. *BioSci.* 43:454-465

Reid, D.A., Fleming, A.I., Foy, C.D., 1971. A method for determining aluminium response of barley in nutrient solution in comparison to response in Al-toxic soil. *Agron. J.*, 63:600-603.

Rincon M., Gonzales R.A., Aluminium partitioning in intact roots of aluminium-tolerant and aluminium-sensitive wheat (*Triticum aestivum*) cultivars, *Plant Physiol.* 99 (1992) 1021–1028.

Ryan PR, Harsh R, Sanjay G, Walter JH, Emmanuel D (2009). A second mechanism for aluminum resistance in wheat relies on the constitutive efflux of citrate from roots. *Plant Physiol.*, 149:340-351

Sasaki T, Yamamoto Y, Ezaki E, Katsuhara M, Ju A, Ryan P, Delhaize E, Matsumoto H (2004) A wheat gene encoding an aluminium-activated malate transporter. *Plant J.*, 37:645-653

Severi A., Effects of aluminium on some morpho-physiological aspects of *Lemna minor* L., *Atti. Soc. Nat. e Mat. di Modena* 122 (1991) 95–108

Schlede H (1989) Distribution of Acid Soils and Liming Materials in Ethiopia. Ethiopian Institute of Geological Survey, Ministry of Mines and Energy, Addis Ababa, Ethiopia.

Soileau J.M., Engelstad O.P., Cotton growth in an acid fragipan subsoil. I. effects of physical soil properties, limiting and fertilization on root penetration, *Soil Sci. Soc. Am. Proc.* 33 (1969) 915–919.

Soileau J.M., Engelstad O.P., Martin J.B., Cotton growth in an acid fragipan subsoil. II. effects of soluble calcium, magnesium and aluminium on roots and tops, *Soil Sci. Soc. Am. Proc.* 33 (1969) 915–919

Stange, M.B., Beratto, M.E., Montenegro, B.A., Peyrelongue, C.A., Borie, B.F., 1995. Effect of nitrogen source on growth of barley on a soil with a high aluminium content. *Agricultura Tecnica (Santiago)*, 55:118-126.

Thawornwong N., van Diest A., Influences of high acidity and aluminium on the growth of lowland rice, *Plant and Soil* 41 (1974) 141–159.

Trim A.R., Metal ions as precipitants for nucleic acid and their use in the isolation of polynucleotides from leaves, *Biochem. J.* 73 (1959) 298–304

The C, Calba H, Zonkeng C, Ngonkeu E, Adetimirin V, Mafouasson H, Meka S, Horst W (2006) Responses of maize grain yield to changes in acid soil characteristics after soil amendments. *Plant Soil* 284:45- 57.

Vanpraag H.J., Weissen F., Aluminium effects on spruce and beech seedlings, *Plant and Soil* 83 (1985) 331–338.

Vitorello VA, Capaldi FR, Stefanuto VA (2005) Recent advances in aluminum toxicity and resistance in higher plants. *Brazilian J. Plant Physiol.*, 17:129-143.

Von Uexküll HR, Mutert E (1995) Global extent, development and economic impact of acid soils. *Plant Soil.* 171:1–15.

Wang J-p, Raman H, Zhang G-p, Mendham N, Zhou MX (2006). Aluminium tolerance in barley (*Hordeum vulgare* L.): Physiological mechanisms, genetics and screening methods. *J. Zhejiang Univ– Sci. B.*, 7:769-787.

Wang J, Raman H, Zhou M, Ryan P, Delhaize E, Hebb D, Coombes N, Mendham N (2007). High-resolution mapping of the *Alp* locus and identification of a candidate gene *HvMATE* controlling aluminium tolerance in barley (*Hordeum vulgare* L.). *Theor. Appl. Genet.*, 115:265-276.

Wagatsuma T., Kaneko M., Hayasaka Y., Destruction process of plant root cells by aluminium, *Soil. Sci. Plant Nutr.* 33 (1987) 161–175.