

Facultative Methylotrophic Bacterium *Brevibacterium Methylicum* in the Process of Preparation of Highly Deuterated Phenylalanine, Alanine, Valine, and Leucine/Isoleucine

Oleg Mosin¹

Ignat Ignatov^{2*}

1. PhD (Chemistry), Moscow State University of Applied Biotechnology,

Talalikhina St., 33, Moscow 109316, Russia

2. DSc, Professor, Scientific Research Center of Medical Biophysics (SRCMB),

Nik. Kopernik St., 32, Sofia, 1111 Bulgaria

* E-mail of the corresponding author: mbioph@dir.bg

Abstract

Microbiological preparation of [²H]amino acids was performed by conversion of low molecular weight substrates ([U-²H]MeOH and ²H₂O) by Gram-positive aerobic facultative methylotrophic bacterium, *Brevibacterium methylicum*, being L-phenylalanine producer having ribulose-5-monophosphate (RuMP) cycle for carbon assimilation. For this purpose, the cells of the methylotroph with improved growth characteristics were used on minimal salt media M9 supplemented with 2% (v/v) [U-²H]MeOH and increasing gradient of ²H₂O concentration from 0; 24.5; 49.0; 73.5 up to 98% (v/v) ²H₂O. L-phenylalanine was isolated from the growth medium after adding 5 M ²HCl (in ²H₂O), pH = 2.0 by extraction with isopropanol and subsequent crystallization in ethanol (output 0.65 g/l). Alanine, valine, and leucine/isoleucine were produced and accumulated exogenously in amounts of 5–6 μmol in addition to the main product of biosynthesis. The method allows to obtain [²H]amino acids with different levels of deuterium enrichment, depending on ²H₂O concentration in growth media, from 17% ²H (2 deuterium atoms) (on the growth medium with 24.5% (v/v) ²H₂O) up to 75% ²H (6 deuterium atoms) (on the growth medium with 98% (v/v) ²H₂O) with introduction of deuterium to benzyl C₆H₅CH₂-fragment of molecule that is confirmed with the data of electron impact (EI) mass spectrometry analysis of methyl ethers of N-5-dimethylamino(naphthalene)-1-sulfochloride [²H]amino acids after the separation by reverse-phase HPLC.

Keywords: *Brevibacterium methylicum*, [U-²H]MeOH, heavy water, biosynthesis, [²H]amino acids, EI mass spectrometry, HPLC.

1. Introduction

Labeling of amino acid molecules with deuterium is becoming an essential part for various biochemical and biomedical studies with ²H-labeled molecules and investigation of certain aspects of their metabolism (LeMaster, 1990; V értes, 2011). Tendencies to preferable using of stable isotopes in clinical diagnostics in comparison with their radioactive analogues are caused by absence of radiation danger and possibility of localization of a label in a molecule by highly resolution methods as NMR spectroscopy and mass spectrometry (Mosin et al., 1996). Development of these methods for detecting stable isotopes for last years has allowed to improve considerably carrying out numerous biological studies with amino acids de novo, and also to study the pathways of their metabolism. ²H-labeled amino acids are applied in medical

diagnostics and biochemical research (Kushner et al, 1999), as well as in chemical synthesis of various isotopically labeled compounds on their basis, for example, [^2H]phenylalanine – in synthesis of peptide hormones and neurotransmitters. Isotopically labeled analogues of L-phenylalanine find the increasing application in diagnostic purposes, for example, for detecting hereditary phenylketonuria (PKU) – a metabolic genetic disorder characterized by a mutation in the gene of the hepatic enzyme phenylalanine hydroxylase (PAH), making it non-functional and connected with disorders of phenylalanine metabolism in organism (DiLella et al., 1986; Surtees & Blau, 2000). For biomedical diagnostic of PKU is essential to study distribution of isotope label in end-products of metabolism of this amino acid by introducing isotopically labelled phenylalanin into organism (Dewick, 2009; Mosin, 1996). Therefore it is important to develop new biotechnological approaches for preparation of natural isotopically labelled analogues of phenylalanine, including its deuterated analogues. Advantages of biotechnological methods for synthesis of isotopically labelled amino acids in comparison with chemical synthesis consist in high yields of synthesised products and that they possess a natural L-configuration.

For introduction of deuterium into amino acid molecules either chemical or biosynthetic methods may be used. Chemical synthesis of ^2H -labeled compounds has one significant limitation; it is a very laborious and costly multistep process resulting in obtaining a racemic mixture of D,L- enantiomers (Blomquist et al., 1966). Although chemomicrobiological synthesis overcomes this problem (Faleev et al., 1989), the amounts of purified enzymes, required is prohibitive (Walker & Matheny, 1986). The biosynthesis of ^2H -labeled amino acids usually involves growth of an organism on selective growth media containing the labeled substrates: e.g., growth of algae autotrophically on media with 96% (v/v) $^2\text{H}_2\text{O}$, is a well established method for biosynthesis of highly deuterated biochemicals including amino acids (Cox & Kyli, 1988; Crespi, 1986). Though this method, being generally applicable, is still limited due to lower resistance of plant cells towards $^2\text{H}_2\text{O}$ along with the expense of the mixture of ^2H -labeled amino acids which is isolated from hydrolysates of biomass (Den'ko, 1970; Mosin & Ignatov, 2013).

Alternatively inexpensive sources for microbiological synthesis of [^2H]amino acids seem methylotrophic bacteria, capable to oxidize methanol (MeOH) and other one-carbon compounds containing methyl CH_3 -group to formaldehyde via ribulose-5-monophosphate and serine pathways of carbon assimilation (Antony, 1986). Interest to use methylotrophic bacteria in biotechnology has recently increased owing to development of new perspective technologies for chemical synthesis of methanol. Owing to 50% level of bioconversion of methanol (at efficiency of methanol conversion 15,5–17,3 gram of dry biomass for 1 gram of consumed substrate) methylotrophic bacteria are considered as cheap sources of deuterated protein and irreplaceable amino acids, whose technological expenses for their preparation are defined, basically by cost of $^2\text{H}_2\text{O}$ and [^2H]methanol. These bacteria have shown a great practical advantage in terms of its industrial usage because of their ability to produce and accumulate in culture liquid large quantities of ^2H -labeled amino acids during its growth on media supplemented with $^2\text{H}_2\text{O}$ and [^2H]MeOH and is a cost effective process due to lower price of [^2H]MeOH (Karnaukhova et al., 1993). The traditional approach for this aim is cultivation of strain-producers of amino acids on growth media containing [^2H]methanol and heavy water ($^2\text{H}_2\text{O}$) with subsequent fractionation of liquid cultural media for isolation of [^2H]amino acids (Mosin et al., 2012).

In the past few years, a certain progress was observed in terms of isolation of methylotrophs wherein RuMP cycle operates. However still for such studies, limited research is carried out with methylotrophs due to poor growth characteristics obtained on $^2\text{H}_2\text{O}$ containing media. Realizing this, the production of ^2H -labeled amino acids by obligate methylotrophs *Methylobacillus flagellatum* for carbon assimilation was conducted by 2-keto 3-deoxy 6-phospho-6-gluconate aldolase/transaldolase (KDPGA/TA) variant of RuMP cycle which involves the bacterial growth on salt medium supplemented with approx. 75% (v/v) $^2\text{H}_2\text{O}$ (Skladnev et al., 1996). In the present study, a mutant strain of Gram-positive aerobic facultative methylotroph, *Brevibacterium methylicum*, with NAD^+ dependent methanol dehydrogenase (EC 1.6.99.3) variant of RuMP cycle for carbon assimilation was selected, which seems more convenient for the preparation of ^2H -labeled amino acids because of its ability to grow on minimal salt media M9 with 98% (v/v) $^2\text{H}_2\text{O}$ as compared to the obligate methylotroph *M. flagellatum* (Mosin et al., 1998).

The main aim of the present study is to study the physiological adaptation through deuterated enrichment

technique for Gram-positive aerobic facultative methylotrophic bacterium *B. methylicum*, L-phenylalanine producer, to high level of $^2\text{H}_2\text{O}$ in order to increase level of [^2H]amino acids by this selected bacterium.

2. Material and Methods

2.1. Chemicals

$^2\text{H}_2\text{O}$ (99.9% (atom) ^2H) was purchased from JSC “Isotope” (Sankt Petersburg, Russia), [$\text{U-}^2\text{H}$]MeOH (97.5% (atom) ^2H) (Biophysics Center, Pushino, Russia). Dansylchloride (DNSCl) of sequential grade was from Sigma Chemicals Corp. (USA). Diazomethane (DZM) was prepared from N-nitroso-methylurea (N-NMU) (“Pierce Chemicals Corp.”, USA). Inorganic salts were crystallized in 99.9% (atom) $^2\text{H}_2\text{O}$; $^2\text{H}_2\text{O}$ was distilled over KMnO_4 with the subsequent control of isotopic enrichment by $^1\text{H-NMR}$ spectroscopy on Bruker WM-250 device (“Bruker Corp.”, USA) (working frequency – 70 MHz, internal standard – Me_4Si).

2.2. Bacterial Strain

A Gram-positive parental strain of L-leucine auxotroph of facultative methylotrophic bacterium *Brevibacterium methylicum* # 5662, producer of L-phenylalanine used in this research was obtained from Russian State Scientific Center for Genetics and Selection of Industrial Microorganisms GNIIGENETIKA. The parental strain was modified by adaptation to $^2\text{H}_2\text{O}$ *via* plating the cells onto 2% (w/v) agarose media with an increasing gradient of $^2\text{H}_2\text{O}$ concentration from 0; 24.5; 49.0; 73.5 up to 98% (v/v) $^2\text{H}_2\text{O}$ and the subsequent selection of separate colonies resistant to $^2\text{H}_2\text{O}$, capable to produce L-phenylalanine while grown on liquid growth media with the same isotopic content.

2.3. Adaptation and Growth Conditions

The parent strain was adapted to deuterium *via* plating the cells to individual colonies followed by subsequent selection of cell colonies which were resistant to deuterium on solid 2% (w/v) agarose media M9 with 2% (v/v) MeOH/[$\text{U-}^2\text{H}$]MeOH containing a step-wise increasing gradient of $^2\text{H}_2\text{O}$ concentration from 0; 24.5; 49.0; 73.5 up to 98% (v/v) $^2\text{H}_2\text{O}$. Minimal salt medium M9 (g/l): KH_2PO_4 – 3; Na_2HPO_4 – 6; NaCl – 0.5; NH_4Cl – 1, with MeOH (2%, v/v) as a carbon and energy source and L-Leu (100 mg/l) was used for bacterial growth. Selection of separate colonies was performed by their resistance to $^2\text{H}_2\text{O}$. Cell colonies grown on the growth media M9 with a low gradient of $^2\text{H}_2\text{O}$ concentration were transferred onto the growth media with higher gradient of $^2\text{H}_2\text{O}$ concentration; this was repeated up to 98% (v/v) $^2\text{H}_2\text{O}$. At a final stage of this technique on growth medium with 98% (v/v) $^2\text{H}_2\text{O}$ were isolated the separate cellular colonies representing posterity of one cell, resistant to the action of $^2\text{H}_2\text{O}$. Then cells were transferred to the liquid growth medium M9 with the same deuterium content, and grew up during 3–4 days at temperature 34 °C. The adaptation was monitored by formation of individual colonies onto the surface of solid agarose media with $^2\text{H}_2\text{O}$ and also by values of optical density (OD) of cell suspension in liquid media, determined in a quartz cuvette (light path 10 mm) at $\lambda = 620$ nm using a Beckman DU-6 spectrophotometer (“Beckman Coulter”, USA). The bacterium adapted to $^2\text{H}_2\text{O}$ was grown on the maximally deuterated M9 medium with 2% (v/v) [^2H]methanol and 98% (v/v) $^2\text{H}_2\text{O}$ in 250 ml Erlenmeyer flasks containing 50 ml of growth medium at 32–34 °C and vigorously aerated at 200 rpm on orbital shaker Biorad 380-SW (“Biorad Labs”, Poland). The exponentially growing cells (cell density 2.0 at absorbance 620 nm) were pelleted by centrifugation (1200 g for 15 min), the supernatant was lyophilized in vacuum and used for chemical derivatization.

2.4. Determination of Phenylalanine

The amount of phenylalanine was determined for 10 μl aliquots of liquid minimal salt medium M9 by TLC method with solvent mixture iso-PrOH–ammonia (7 : 3, v/v) using pure commercially available

phenylalanine as a standard. The spots were treated with 0.1% (w/v) ninhydrin solution (prepared in acetone), and was eluted with 0.5% (w/v) CdCl₂ solution prepared in 50% (v/v) EtOH (in H₂O) (2 ml). Absorbance of the eluates was measured on spectrophotometer Beckman DU-6 (“Beckman Coulter”, USA) at $\lambda = 540$ nm, the concentration of phenylalanine in samples was calculated using a standard curve.

2.5. Isolation of [²H]Phenylalanine from Growth Medium

Deutero-biomass of *B. methylicum* obtained after growth in maximally deuterated medium M9 with 2% (v/v) [U-²H]MeOH and 98% (v/v) ²H₂O, was separated using centrifugation (“Heraeus Sepatech”, Model T-24, Germany) at 1200 g for 15 min. Supernatant thus obtained was evaporated under vacuum. To 5 g of the lyophilized growth medium M9; 30 ml of iso-PrOH was added, and thus acidified to pH = 2.0 using 5 M ²HCl (in ²H₂O) and keep at room temperature for 4 h. The salts were removed through centrifugation and the supernatant was evaporated. Phenylalanine (0.65 g/l) was recrystallized from EtOH: $[\alpha]_D^{20} = 35^0$ (EtOH); UV-spectrum (0.1 M HCl): $\lambda_{\max} = 257.5$ nm, $\epsilon_{257.5} = 1.97 \cdot 10^2 \text{ M}^{-1} \cdot \text{cm}^{-1}$.

2.6. Synthesis of Methyl Esters of N-DNS-[²H]Amino Acids

To 200 mg of lyophilized growth media M9 in 5 ml 2 M NaHCO₃ (0.002 mol) solution, pH = 9–10, 320 mg (1.2 mol) DNSCl in 5 ml of acetone was added. A mixture was kept at 40 °C under vigorous stirring for 1 h. After the reaction was completed, the solution was acidified by 2 M HCl till pH = 3.0, and extracted with ethyl acetate (3 × 5 ml). The combined extracts were dried over sodium sulphate and evaporated under vacuum. The further derivatization to methyl esters of N-DNS-[²H]amino acids was performed using a standard chemical procedure with DZM. To, 20 ml of 40% (w/v) KOH in 40 ml of diethyl ether 3 g of N-NMU was added and mixed at 120 rpm on a water bath cum stirrer at 4 °C for 15–20 min. After intensive evolution of gaseous DZM ether layer was separated and washed out by cold water till pH = 7.0 was achieved, then dried over sodium sulphate for further processing and analysis.

2.7. High-Performance Liquid Chromatography

Analytical separation of methyl esters of N-DNS-[²H]amino acids was performed by a reversed-phase HPLC on liquid chromatograph Knauer (“Knauer GmbH”, Germany), supplied with UV-detector and integrator C-R 3A (“Shimadzu”, Japan). Methyl esters of N-DNS-[²H]amino acids were detected at $\lambda = 254$ nm by UV-absorbance. As a motionless phase was used Separon SGX C₁₈ (“Kova”, Czech Republic), 22 °C, 18 μm , 150 × 3.3 mm. The mobile phase composed of a mixture of solvents: (A) – acetonitrile–trifluoroacetic acid (20: 80%, v/v) and (B) – acetonitrile (100%, v/v). The gradient started from 20% A to 100% B for 30 min; at 100% B for 5 min; from 100% B to 20% A for 2 min; at 20% A for 10 min.

2.8. EI Mass-Spectrometry

EI MS was performed for the determination of deuterium enrichment levels using methyl esters of N-DNS-[²H] amino acids on Hitachi MB 80 mass spectrometer (“Hitachi”, Japan) at an ionizing energy of 70 eV, with an accelerating electrical voltage of 8 kV, and an ion source temperature of 180–200 °C. Each measurement was repeated at least three times. Calculation of deuterium enrichment of [²H]amino acids was performed on a parity of contributions of molecular ion (M⁺) peaks of N-DNS-[²H]amino acid methyl esters obtained in isotopic experiments relative to the control in protonated minimal medium M9. Statistical processing of experimental data was carried out with using the program of statistical package, STATISTICA 6.

3. Results and Discussion

3.1. Study of phenylalanine biosynthesis by the strain *B. methylicum* in $^2\text{H}_2\text{O}$ containing media

As is known, the majority of the microorganisms distributed in nature cannot serve as good producers of aromatic amino acids, owing to the presence in a cell effective mechanism of regulation of their biosynthesis, though this ability is known for a number of their mutant forms (Mosin et al., 1996). Effective microbial producers of *L*-phenylalanine are, as a rule, the mutants, which do not have negative control over such key enzymes of biosynthesis of this amino acid as prephenate dehydratase (EC 4.2.1.51), chorismate mutase (EC 5.4.99.5) and 3-deoxy-D-arabino-heptulosonate-7-phosphate synthase (EC 2.5.1.54). Phenylalanine is synthesized in most bacteria *via* shikimic acid pathway from prephenic acid, which through formation of phenylpyruvate that turns into phenylalanine under the influence of cellular transaminases (Conn, 1986; Herrmann & Weaver, 1999). The precursors for the biosynthesis of phenylalanine are phosphoenolpyruvate (PRP) and erythrose-4-phosphate (ERP). The latter compound is an intermediate in the pentose phosphate (PenP) pathway and, in some methylotrophs, during the RuMP cycle of carbon assimilation (Antony & Williams, 2003; Lindstrom & Stirling, 1990). The basic metabolic transformation of phenylalanine in animals is enzymatic hydroxylation of this amino acid with formation of tyrosine (Bohinski, 1983).

Phenylalanine is synthesized in most organisms *via* shikimic acid pathway from the common aromatic amino acid precursors as erythrose-4-phosphate (E4P) formed through formation stages of shikimic acid, chorismic acid and prephenic acid (Fig. 1). The basic intermediate products of phenylalanine catabolism and metabolically related tyrosine in various organisms are fumaric acid, pyruvate, succinic acid, acetaldehyde etc. At hereditary phenylketonuria disease the hepatic enzyme phenylalanine hydroxylase (PAH), which is necessary to metabolize phenylalanine to tyrosine is nonfunctional, that caused accumulation of phenylalanine and its metabolites (phenylpyruvate, phenyllactate, phenylacetate, *ortho*-hydroxyphenyl acetate), superfluous amounts of which negatively affect development of nervous system (Michals & Matalon, 1985; Filiano, 2006).

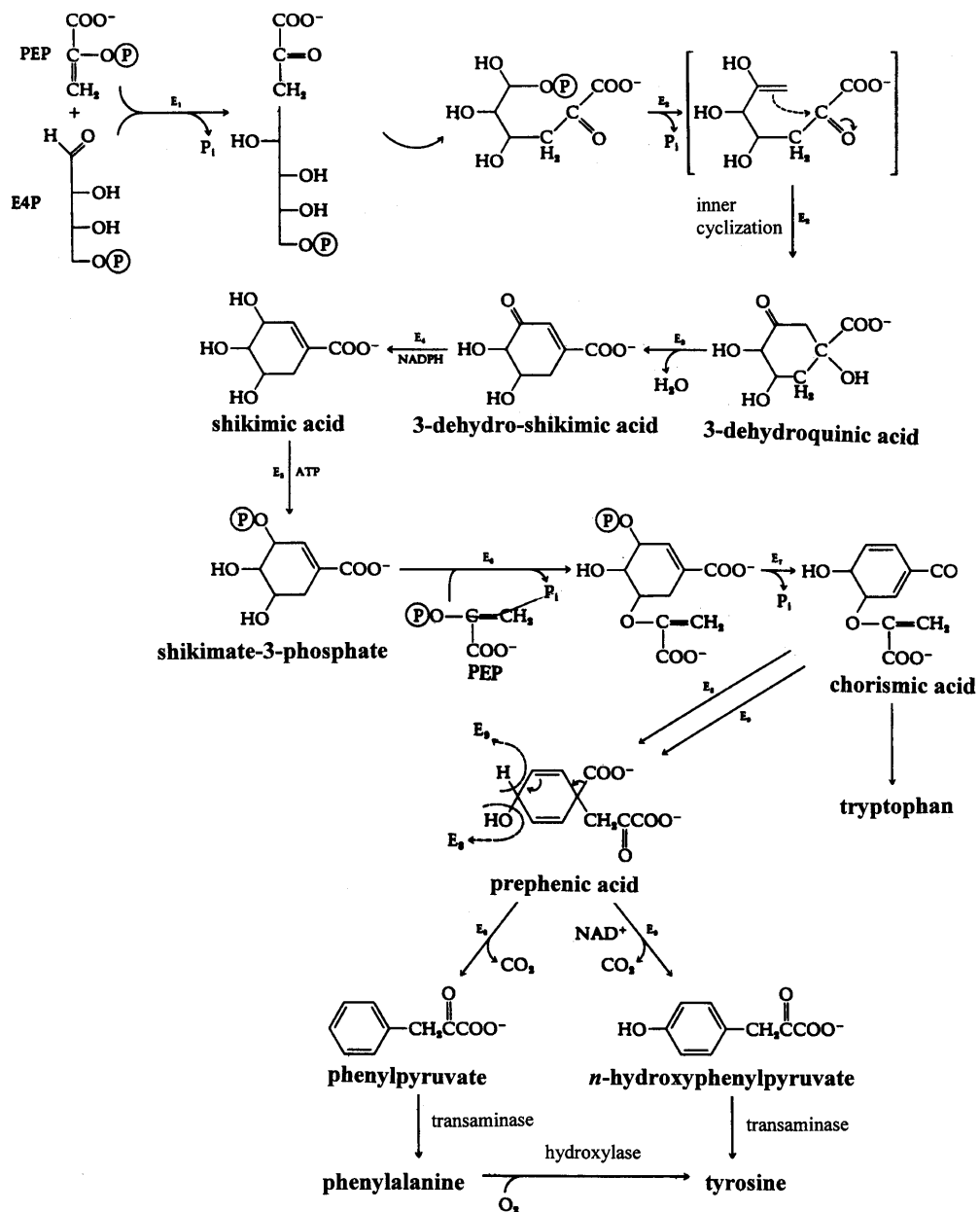


Figure 1. Scheme of biosynthesis of aromatic amino acids – phenylalanine, tyrosine and tryptophan from the general predecessors – phosphoenolpyruvate (PEP) and erythrose-4-phosphate (E4P) (Mosin & Ignatov, 2012)

The native bacterial strains can not be the potential producers of phenylalanine owing to their effective mechanisms in its metabolic regulation, although certain bacterial mutants with mutations in their metabolic machinery such as in their of prephenate dehydrogenase (EC 1.3.1.12), prephenate hydratase (EC 4.2.1.51), chorismate mutase (EC 5.4.99.5), 3-deoxy-D-arabino-heptulosonate-7-phosphate synthase (EC 2.5.1.54) and in other enzymes which thereby proves to be an active producers of this amino acid

(Wendisch, 2007). The best phenylalanine producing strains once selected were the mutants partially or completely dependent on tyrosine or tryptophan for growth. The reports about other regulative mechanisms for phenylalanine biosynthesis in bacterial cells are quite uncommon, though it is known that RuMP cycle operates in certain number of auxotroph mutants of methylotrophs, covers numerous steps in aromatic amino acid biosynthesis (de Boer et al., 2007; Abou-Zeid et al., 1995).

A certain practical interest represents the research of the ability to produce L-phenylalanine by a leucine auxotroph of Gram-negative aerobic facultative methylotrophic bacterium *B. methylicum* realizing the NAD^+ dependent methanol dehydrogenase (EC 1.6.99.3) variant of the RuMP cycle of carbon assimilation – convenient, but insufficiently explored object for biotechnological usage. Unlike other traditional producers of L-phenylalanine, which do not have negative control of prephenate dehydratase (EC 4.2.1.51) or 3-deoxy-D-arabino-heptulosonate-7-phosphate synthase (EC 2.5.1.54), the uniqueness of this strain consists in that it requires L-leucine for growth. The initial stage of biochemical research with this strain of methylotrophic bacteria was connected with obtaining auxotrophic mutants, which in majority cases are characterized by the limited spectrum of mutant phenotypes and, besides that the high level of reversions. The initial L-leucine dependent strain *B. methylicum*, producer of L-phenylalanine was obtained *via* selection at previous stage of research after processing of parental strain by nitrozoguanidin. Screening for resistant cell colonies was carried out by their stability to the analogue of phenylalanine – *meta*-fluoro-phenylalanine (50 $\mu\text{g}/\text{ml}$). The analogue resistant mutants allocated on selective media were able to convert methanol and accumulate up to 1 gram per 1 liter of L-phenylalanine into growth media. Comparative analyses (TLC, NMR) showed that L-phenylalanine, produced by this strain of methylotrophic bacteria is identical to the natural L-phenylalanine.

Further attempts were made to intensify the growth and biosynthetic parameters of this bacterium in order to grow on media M9 with higher concentration of deuterated substrates – $[\text{U}-^2\text{H}]\text{MeOH}$ and $^2\text{H}_2\text{O}$. For this, deuterium enrichment technique *via* plating cell colonies on 2% (w/v) agarose media M9 supplemented with 2% (v/v) $[\text{U}-^2\text{H}]\text{MeOH}$ with an increase in the $^2\text{H}_2\text{O}$ content from 0; 24.5; 49.0; 73.5 up to 98% (v/v) $^2\text{H}_2\text{O}$, combined with subsequent selection of cell colonies which were resistant to deuterium. The degree of cell survive on maximum deuterated medium was approx. 40%. The data on the yield of biomass of initial and adapted *B. methylicum*, magnitude of lag-period and generation time on protonated and maximum deuterated M9 medium are shown in Fig. 2. The yield of biomass for adapted methylotroph (c) was decreased approx. on 13% in comparison with control conditions (a) at an increase in the time of generation up to 2.8 h and the lag-period up to 40 h (Fig. 2). As is shown from these data, as compared with the adapted strain, the growth characteristics of initial strain on maximally deuterated medium were inhibited by deuterium.

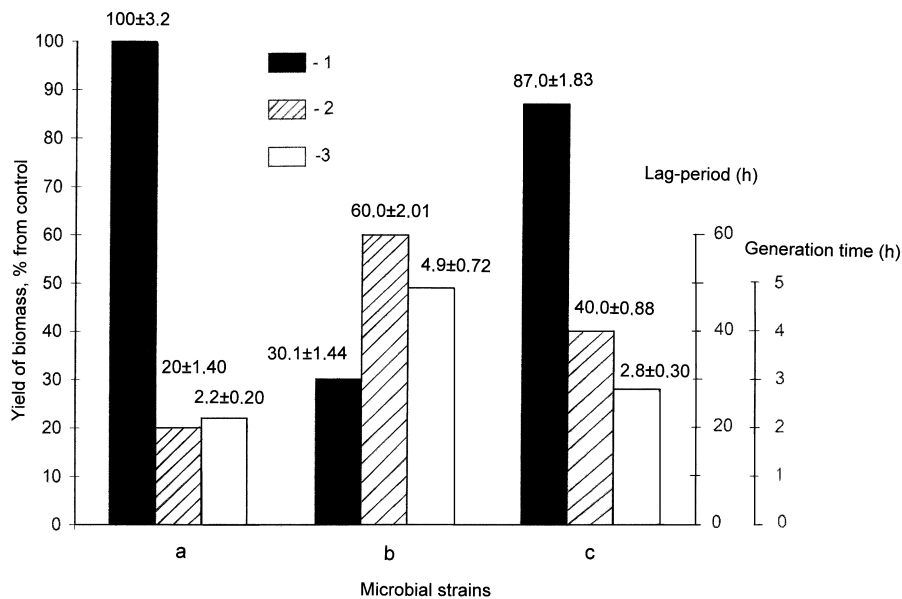


Figure 2. Yield of microbial biomass of *B. methylicum*, magnitude of lag-period and generation time in various experimental conditions: initial strain on protonated M9 medium (control) with water and methanol (a); initial strain on maximally deuterated M9 medium (b); adapted to deuterium strain on maximally deuterated M9 medium (c): 1 – yield of biomass, % from the control; 2 – duration of lag-period, h; 3 – generation time, h.

Experimental conditions are given in Table 1 (expts. 1–10) relative to the control (expt. 1) on protonated medium M9 and to the adapted bacterium (expt. 10'). Various compositions of [U-²H]MeOH and ²H₂O were added to growth media M9 as hydrogen/deuterium atoms could be assimilated both from MeOH and H₂O. The maximum deuterium content was under conditions (10) and (10') in which we used 98% (v/v) ²H₂O and 2% (v/v) [U-²H]MeOH. The even numbers of experiment (Table 1, expts. 2, 4, 6, 8, 10) were chosen to investigate whether the replacement of MeOH by its deuterated analogue affected growth characteristics in presence of ²H₂O. That caused small alterations in growth characteristics (Table 1, expts. 2, 4, 6, 8, 10) relative to experiments, where we used protonated methanol (Table 1, expts. 3, 5, 7, 9). The gradual increment in the concentration of ²H₂O into growth medium caused the proportional increase in lag-period and yields microbial biomass in all isotopic experiments. Thus, in the control (Table 1, expt. 1), the duration of lag-period did not exceed 20.2 h, the yield of microbial biomass (wet weight) and production of phenylalanine were 200.2 and 0.95 gram per 1 liter of growth medium. The results suggested, that below 49% (v/v) ²H₂O (Table 1, expts. 2–4) there was a small inhibition of bacterial growth compared with the control (Table 1, expt. 1). However, above 49% (v/v) ²H₂O (Table 1, expts. 5–8), growth was markedly reduced, while at the upper content of ²H₂O (Table 1, expts. 9–10) growth got 3.3-fold reduced. With increasing content of ²H₂O in growth media there was a simultaneous increase both of lag-period and generation time. Thus, on maximally deuterated growth medium (Table 1, expt. 10) with 98% (v/v) ²H₂O and 2% (v/v) [U-²H]MeOH, lag-period was 3 fold higher with an increased generation time to 2.2 fold as compared to protonated growth medium with protonated water and methanol which serve as control (Table 1, expt. 1). While on comparing adapted bacterium on maximally deuterated growth medium (Table 1, expt. 10') containing 98% (v/v) ²H₂O and 2% (v/v) [U-²H]MeOH with non adapted bacterium at similar concentration showed 2.10 and 2.89 fold increase in terms of phenylalanine production and biomass yield due to deuterium enrichment technique, while, the lag phase as well as generation time also got reduced to 1.5 fold and 1.75 fold in case of adapted bacterium.

Table 1. Effect of variation in isotopic content (0–98% $^2\text{H}_2\text{O}$, v/v) present in growth medium M9 on bacterial growth of *B. methylicum* and phenylalanine production

Bacterial strains	Exp. number	Media components, % (v/v)				Lag-period (h)	Yield in terms of wet biomass (g/l)	Generation time (h)	Phenylalanine production (g/l)
		H ₂ O	² H ₂ O	MeOH	[U- ² H]MeOH				
Non adapted	1 (control)	98.0	0	2	0	20.2±1.40	200.2±3.20	2.2±0.20	0.95±0.12
Non adapted	2	98.0	0	0	2	20.3±1.44	184.6±2.78	2.4±0.23	0.92±0.10
Non adapted	3	73.5	24.5	2	0	20.5±0.91	181.2±1.89	2.4±0.25	0.90±0.10
Non adapted	4	73.5	24.5	0	2	34.6±0.89	171.8±1.81	2.6±0.23	0.90±0.08
Non adapted	5	49.0	49.0	2	0	40.1±0.90	140.2±1.96	3.0±0.32	0.86±0.10
Non adapted	6	49.0	49.0	0	2	44.2±1.38	121.0±1.83	3.2±0.36	0.81±0.09
Non adapted	7	24.5	73.5	2	0	45.4±1.41	112.8±1.19	3.5±0.27	0.69±0.08
Non adapted	8	24.5	73.5	0	2	49.3±0.91	94.4±1.74	3.8±0.25	0.67±0.08
Non adapted	9	98.0	0	2	0	58.5±1.94	65.8±1.13	4.4±0.70	0.37±0.06
Non adapted	10	98.0	0	0	2	60.1±2.01	60.2±1.44	4.9±0.72	0.39±0.05
Adapted	10'	98.0	0	0	2	40.2±0.88	174.0±1.83	2.8±0.30	0.82±0.08

Notes:

* The data in expts. 1–10 described the growth characteristics for non-adapted bacteria in growth media, containing 2 % (v/v) MeOH/[U-²H]MeOH and specified amounts (% , v/v) of $^2\text{H}_2\text{O}$.

** The data in expt. 10' described the growth characteristics for bacteria adapted to maximum content of deuterium in growth medium.

***As the control used expt. 1 where used ordinary protonated water and methanol

The adapted strain of *B. methylicum* eventually came back to normal growth at placing over in protonated growth medium after some lag-period that proves phenotypical nature of a phenomenon of adaptation that

was observed for others adapted by us strains of methylotrophic bacteria and representatives of other taxonomic groups of microorganisms (Mosin & Ignatov, 2012; Mosin & Ignatov, 2013a, Ignatov & Mosin, 2013a; Ignatov & Mosin, 2013b) The effect of reversion of growth in protonated/deuterated growth media proves that adaptation to $^2\text{H}_2\text{O}$ is rather a phenotypical phenomenon, although it is not excluded that a certain genotype determines the manifestation of the same phenotypic trait in the growth media with different isotopic compositions. On the whole, improved growth characteristics of adapted methylotroph essentially simplify the scheme of obtaining the deuterio-biomass which optimum conditions are M9 growth medium with 98% $^2\text{H}_2\text{O}$ and 2% [^2H]methanol with incubation period 3–4 days at temperature 35 °C.

Literature reports clearly reveal that the transfer of deuterated cells to protonated medium M9 eventually after some lag period results in normal growth that could be due to the phenomenon of adaptation wherein phenotypic variation was observed by the strain of methylotrophic bacteria (Mosin & Ignatov, 2013b; Mosin et al., 2013). The effect of reversion of growth in protonated/deuterated growth media proves that adaptation to $^2\text{H}_2\text{O}$ is a phenotypical phenomenon, although it cannot be excluded that a certain genotype determined the manifestation of the same phenotypic attribute in media with high deuterium content. The improved growth characteristics of the adapted bacterium essentially simplify the obtaining of deuterio-biomass in growth medium M9 with 98% (v/v) $^2\text{H}_2\text{O}$ and 2% (v/v) [$\text{U-}^2\text{H}$]methanol.

Adaptation, which conditions are shown in experiment 10⁷ (Table 1) was observed by investigating of growth dynamics (expts. 1a, 1b, 1c) and accumulation of L-phenylalanine into growth media (expts. 2a, 2b, 2c) by initial (a) and adapted to deuterium (c) strain *B. methylicum* in maximum deuterated growth medium M9 (Fig. 3, the control (b) is obtained on protonated growth medium M9). In the present study, the production of phenylalanine (Fig. 3, expts. 1b, 2b, 3b) was studied and was found to show a close linear extrapolation with respect to the time up to exponential growth dynamics (Fig. 3, expts. 1a, 2a, 3a). The level of phenylalanine production for non-adapted bacterium on maximally deuterated medium M9 was 0.39 g/liter after 80 hours of growth (Fig. 3, expt. 2b). The level of phenylalanine production by adapted bacterium under those growth conditions was 0.82 g/liter (Fig. 3, expt. 3b). Unlike to the adapted strain the growth of initial strain and production of phenylalanine in maximum deuterated growth medium were inhibited. The important feature of adapted to $^2\text{H}_2\text{O}$ strain *B. methylicum* was that it has kept its ability to synthesize and exogenously produce L-phenylalanine into growth medium. Thus, the use of adapted bacterium enabled to improve the level of phenylalanine production on maximally deuterated medium by 2.1 times with the reduction in the lag phase up to 20 h. This is an essential achievement for this strain of methylotrophic bacteria, because up till today there have not been any reports about production of phenylalanine by leucine auxotrophic methylotrophs with the NAD^+ dependent methanol dehydrogenase (EC 1.6.99.3) variant of the RuMP cycle of carbon assimilation. This makes this isolated strain unique for production of phenylalanine.

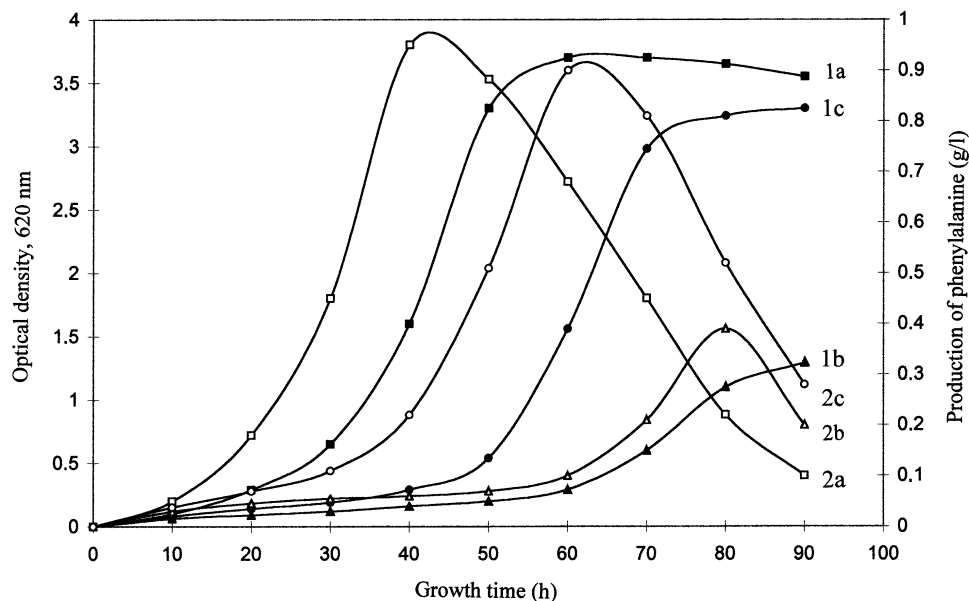


Figure 3. Growth dynamics of *B. methylicum* (1a, 2a, 3a) and production of phenylalanine (1b, 2b, 3b) on media M9 with various isotopic content: 1a, 1b – non-adapted bacterium on protonated medium (Table 1, expt. 1); 2a, 2b – non-adapted bacterium on maximally deuterated medium (Table 1, expt. 10); 3a, 3b – adapted bacterium on maximally deuterated medium (Table 1, expt. 10')

The general feature of phenylalanine biosynthesis in $H_2O/{}^2H_2O$ -media was increase of its production at early exponential phase of growth when outputs of a microbial biomass were insignificant (Fig. 3). In all the experiments it was observed that there was a decrease in phenylalanine accumulation in growth media at the late exponential phase of growth. Microscopic research of growing population of microorganisms showed that the character of phenylalanine accumulation in growth media did not correlate with morphological changes at various stages of the cellular growth. Most likely that phenylalanine, accumulated in growth media, inhibited enzymes of its biosynthetic pathways, or it later may be transformed into intermediate compounds of its biosynthesis, e.g. phenylpyruvate (Maksimova et al., 1990). It is necessary to notice, that phenylalanine is synthesised in cells of microorganisms from prephenic acid, which through a formation stage of phenylpyruvate turns into phenylalanine under the influence of cellular transaminases. However, phenylalanine was not the only product of biosynthesis; other metabolically related amino acids (alanine, valine, and leucine/isoleucine) were also produced and accumulated into growth media in amounts of 5–6 μmol in addition to phenylalanine. This fact required isolation of $[{}^2\text{H}]$ phenylalanine from growth medium, which was carried out by extraction of lyophilized LC with iso-PrOH and the subsequent crystallization of $[{}^2\text{H}]$ phenylalanine in EtOH. Analytical separation of $[{}^2\text{H}]$ phenylalanine and metabolically related $[{}^2\text{H}]$ amino acids was performed using a reversed-phase HPLC on Separon SGX C_{18} Column, developed for methyl esters of N-DNS- $[{}^2\text{H}]$ amino acids with chromatographic purity of 96–98% and yield of 67–89%.

3.2. Study of deuterium inclusion into molecules of secreted amino acids

For evaluation of deuterium enrichment by EI MS method, methyl esters of N-DNS- $[{}^2\text{H}]$ amino acids were applied because the peaks of molecular ions (M^+) allow to monitor the enrichment of multicomponential mixtures of $[{}^2\text{H}]$ amino acids in composition with growth media metabolites. Furthermore, EI MS allows detect amino acid samples with concentrations 10^{-9} – 10^{-10} mol (Mosin, 1996). This method in combination with reversed-phase HPLC has proved to be better for research studies on deuterium enrichment levels of

[²H]amino acids in composition of their multicomponental mixtures of lyophilized growth media, and can be used for the analysis of amino acids of various natural sources. N-DNS-amino acids were obtained through the derivatization of lyophilized growth media with DNSCl. The reaction was carried out in the alkaline environment in the presence of 2 M NaHCO₃ (pH = 9–10) in water-organic solvent (acetone) in the mass ratio of DNSCl – the amino acid, (2: 1 w/w). The volatility of N-DNS-amino acids in the mass spectrometry analysis may be increased by additional derivatization on carboxyl group (esterification) by diazomethane. The choice of DZM as esterification reagent was caused by the necessity of carrying out the reaction of esterification in soft conditions excluding isotopic (¹H–²H) exchange in an aromatic fragment of the phenylalanine molecule. However, with DZM treatment derivatization occurs on αNH₂-group in molecules, so that their N-methylated derivatives could be formed in addition to the methyl ester of N-DNS-amino acids.

The control over deuterium inclusion into phenylalanine molecule due to the conversion of [U-²H]MeOH at growth of the bacterium on medium M9, containing H₂O and 2% (v/v) [U-²H]MeOH (Table 1, expt. 2) has shown insignificant amount of deuterium detected in molecule. The enrichment level was calculated on intensity of a peak (M⁺) at *m/z* 413 minus the contribution of peak of an impurity of a natural isotope (no more than 5%). This testified about delution of deuterium label *via* biochemical processes connected with disintegration of [U-²H]MeOH during its assimilation by a cell, as well as reactions of isotopic (¹H–²H) exchange and dissociation in ²H₂O. Thus, from 4 deuterium atoms in molecule C²H₃O²H, only deuterium at hydroxyl O²H-group is the most mobile and easily dissociates one deuterium atom in water with formation of C²H₃OH. Three remained deuterium atoms in C²H₃OH are included into a cycle of biochemical oxidation of methanol leading to formation of substances more oxidized than methanol, *e.g.* formaldehyde. In particular, this confirms the classical scheme of biochemical oxidations of methanol to formaldehyde by methylotrophic bacteria, after that formaldehyde assimilates by this strain of methylotrophic bacteria *via* RuMP cycle of carbon assimilation.

EI mass spectra of methyl esters of N-DNS-amino acids obtained from growth media M9 where used 0; 49.0; 73.5 and 98% (v/v) ²H₂O with 2% (v/v) [U-²H]MeOH (Table 1, expts. 1, 6, 8, 10) are shown in consecutive order in Fig. 4–7. The fragmentation pathways of methyl esters of N-DNS-amino acids by EI MS method lead to the formation of distinguished peaks of molecular ions (M⁺) from which the fragments with smaller *m/z* ratio further are formed. The peak of amino fragment A at *m/z* 353 generally has a low intensity, while and peak of aminoacyl fragment B at *m/z* 381 has the lowest intensity in EI mass spectra, or ever absent (see as example Fig. 4 corresponds to expt. 1, Table 1). A high continuous left background region in EI mass spectrum at *m/z* 100–200 is associated with peaks of contaminant metabolites and products of derivatization of metabolites of growth medium with DNSCl and DZM, and peaks at *m/z* 250, 234, 170 are fragments of further decay of dansyl fragment to N-(dimethylamino)naphthalene. A right region in EI mass spectra contains four peaks of molecular ions (M⁺) of N-DNS-amino acid methyl esters: Phe at *m/z* 412; Leu/Ile at *m/z* 378; Val at *m/z* 364; Ala at *m/z* 336 (Fig. 4), which fragmentation by the method of EI mass spectrometry will allow to carry out mass spectrometry monitoring of [²H]amino acids in composition of intact LC of the strain-producer, containing mixes of amino acids and other metabolites of growth media before their chromatographic separation. However, since the value of (M⁺) for Leu is the same as for Ile, these two amino acids could not be clearly estimated by EI MS method.

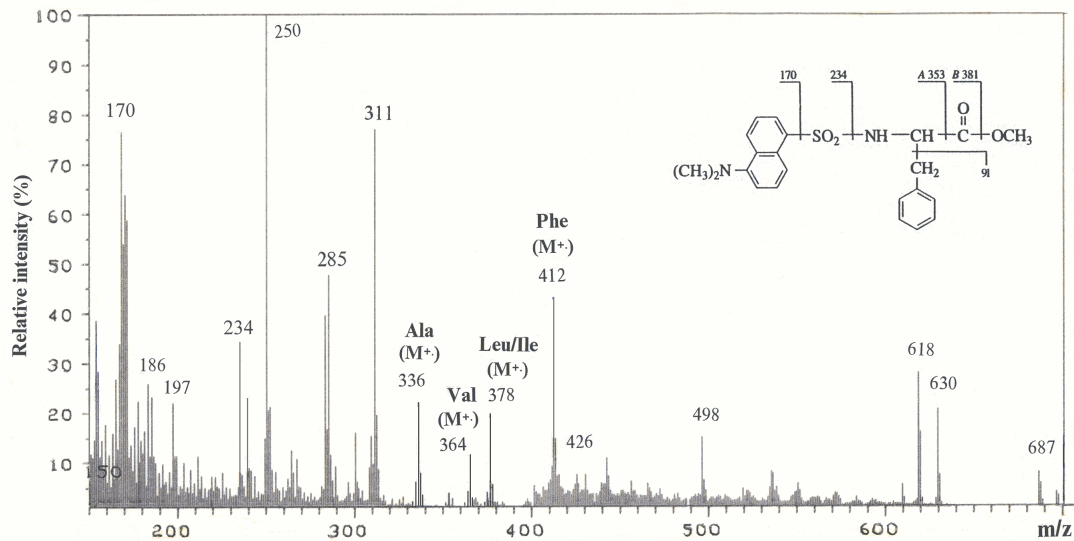


Figure 4. EI Mass spectrum of methyl esters of N-DNS-amino acids from protonated growth medium M9 (expt. 1, Table 1) after processing by DNSCI and DZM, and fragmentation of methyl ester of N-DNS-phenylalanine by EI MS method. Symbols of amino acids refer to the peaks of molecular ions (M^+) of methyl esters of N-DNS-amino acids.

The results confirmed the character of labeling of [^2H]amino acids as heterogeneous, judging by the presence of clusters of adduct peaks in molecular ions (M^+) in EI mass spectra; the species of molecules with different numbers of deuterium atoms were detected. Therefore, the peaks of a molecular ion of amino acid derivatives (M^+) were split polymorphously on separate clusters with an impurity of molecules having a statistical set of mass numbers m/z with the various contribution to a total level of deuterium enrichment of the molecule. The most abundant peak (M^+) in each cluster (the peak with the greatest contribution to level of deuterium enrichment), was a peak with an average m/z ratio registered by mass spectrometer in each experimental condition, relative to which the deuterium enrichment of each individual [^2H]amino acid was calculated. Thus, in experiment (expt. 6, Table 1) shown in Fig. 5 where was used 49% (v/v) $^2\text{H}_2\text{O}$ deuterium enrichment of Phe molecule was 2 (27.5% ^2H), calculated by (M^+) at m/z 414 (instead of (M^+) at m/z 412 for non-labeled compound); Leu/Ile – 5 (50% ^2H) ((M^+) at m/z 383 instead of (M^+) at m/z 378)); Val – 4 (50% ^2H) ((M^+) at m/z 368 instead of m/z (M^+) at 364)); Ala – 3 (50% ^2H) deuterium atoms ((M^+) at m/z 339.2 instead of (M^+) at m/z 336)). The area of EI mass spectrum with values m/z 90–300 corresponds to products of derivatization of metabolites of growth media by dansylchloride and diazomethane. The low intensity peak at m/z 431 detected in EI mass spectra in all isotopic experiments, corresponds to a product of additional methylation of [^2H]phenylalanine on α -NH-(Dns)-group. The peak at m/z 400 (Fig. 5) corresponds to a product of chip off methyl CH_3 -group from deuterated methyl ester of N-Dns- [^2H]phenylalanine.

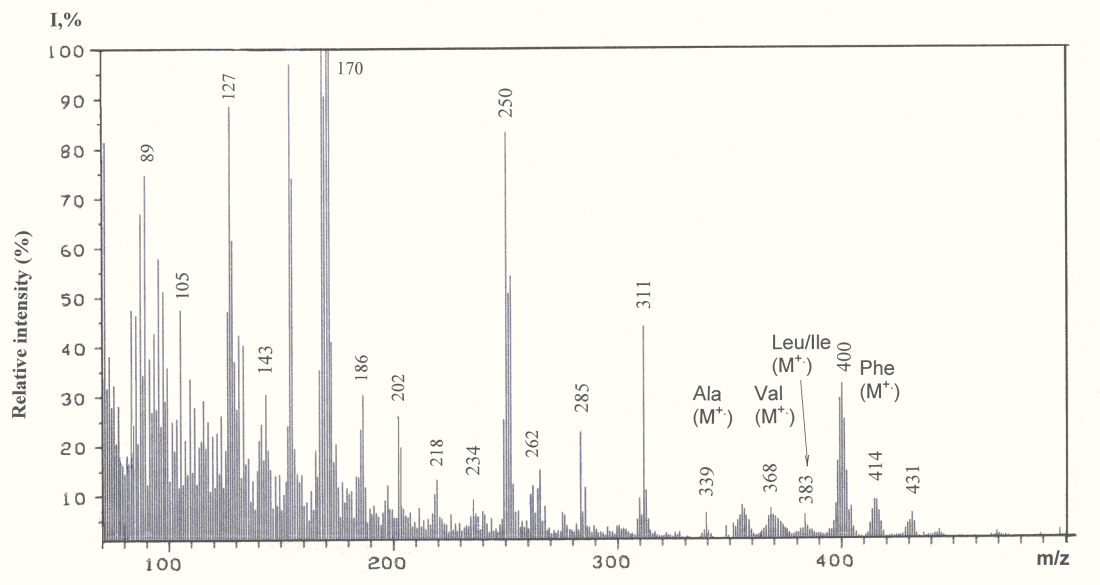


Figure 5. EI mass spectrum of methyl esters of N-DNS- ^{2}H amino acids from growth medium M9 containing 2% (v/v) $[\text{U-}^{2}\text{H}]\text{MeOH}$ and 49.0% (v/v) $^{2}\text{H}_2\text{O}$ (expt. 6, Table 1).

The similar result on proportional specific increase of levels of deuterium enrichment in ^{2}H phenylalanine and other metabolically related ^{2}H amino acids was observed in all isotopic experiments wherein used increasing concentration $^{2}\text{H}_2\text{O}$ in growth media. With increasing $^{2}\text{H}_2\text{O}$ content in growth media M9, the levels of deuterium enrichment in ^{2}H amino acids varied proportionally. As shown in Fig. 6 in experiment (expt. 8, Table 1) where in 73.5% (v/v) of $^{2}\text{H}_2\text{O}$, the deuterium enrichment of Phe was 4 (50% ^{2}H) (M^{+}) at m/z 416 instead of m/z 412 (M^{+}); Leu/Ile – 5 (50% ^{2}H) (M^{+}) at m/z 383 instead of m/z 378 (M^{+}); Val – 4 (50% ^{2}H) (M^{+}) at m/z 368 instead of m/z 364 (M^{+}); Ala – 3 (50% ^{2}H) deuterium atoms (M^{+}) at m/z 339 instead of m/z 336 (M^{+}). Evidently, the deuterium atoms at carbon backbone of ^{2}H amino acid molecules were synthesized *de novo* from deuterated substrates. Easily exchanged protons (deuterons), *e.g.* protons at heteroatoms in NH_2 - and COOH -groups of amino acids could be replaced by deuterium due to dissociation in $\text{H}_2\text{O}/^{2}\text{H}_2\text{O}$.

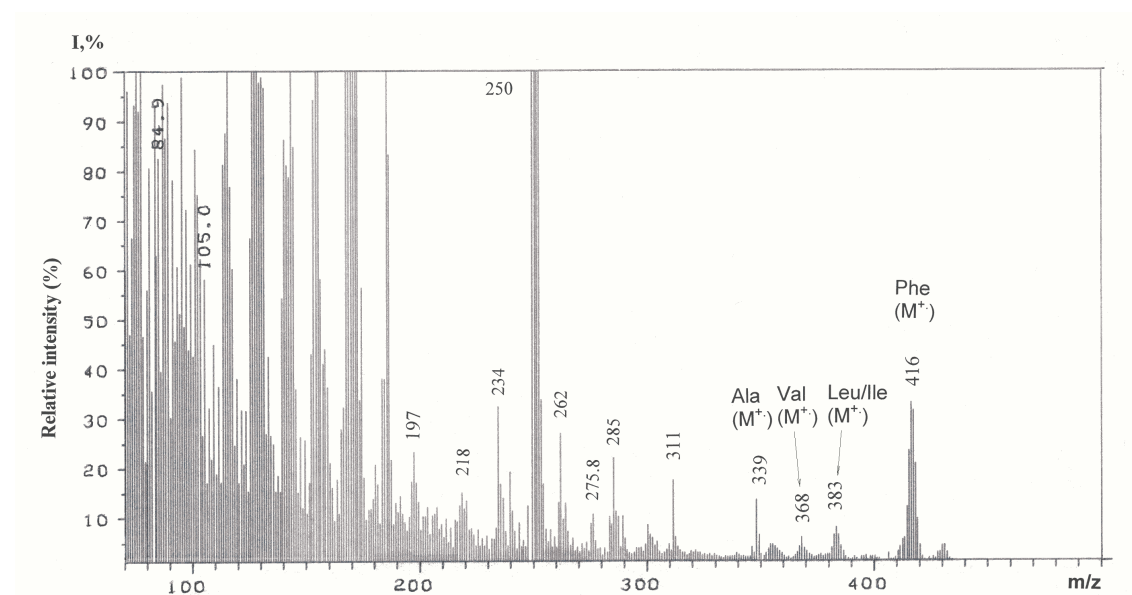


Figure 6. EI mass spectrum of methyl esters of N-DNS- ^{2}H amino acids from growth medium M9 containing 2% (v/v) $[\text{U-}^{2}\text{H}]\text{MeOH}$ and 73.5% (v/v) $^{2}\text{H}_2\text{O}$ (expt. 8, Table 1).

The used methylotrophic bacterium was leucine auxotroph with leucine being added to growth media in protonated form; therefore the levels of deuterium enrichment in ^{2}H amino acids of pyruvic acid family as alanine, valine and leucine were less than for phenylalanine (phenylalanine related to the family of the aromatic amino acids synthesized from shikimic acid). This is distinctly visible on the maximally deuterated M9 medium. As shown in Fig. 7 in experiment (expt. 10, Table 1) wherein 98% (v/v) $^{2}\text{H}_2\text{O}$ was used the deuterium enrichment of Phe molecule was 6 (75 at.% ^{2}H) (M^+) at m/z 418 instead of m/z 412 (M^+); Leu/Ile – 5 (50% ^{2}H) (M^+) at m/z 383 instead of m/z 378 (M^+); Val – 5 (62.5% ^{2}H) (M^+) at m/z 369 instead of m/z 364 (M^+); Ala – 3 (50% ^{2}H) deuterium atoms (M^+) at m/z 339 instead of m/z 336 (M^+). The peak at m/z 432, detected in EI mass spectrum of methyl esters of N-DNS-amino acids in Fig. 7 corresponds to a product of additional methylation of ^{2}H phenylalanine on $\alpha\text{-NH}_2$ -group. Additionally, in EI mass spectrum is detected a peak of deuterium enriched benzyl $\text{C}_6\text{H}_5\text{CH}_2$ -fragment of phenylalanine molecule at m/z 97 (instead of m/z 91 in the control), that specifies sites of localization of 6 deuterium atoms in ^{2}H phenylalanine at positions C1–C6 of aromatic protons in benzyl $\text{C}_6\text{H}_5\text{CH}_2$ -fragment. EI MS data suggest that at other concentration of $^{2}\text{H}_2\text{O}$ deuterium atoms also incorporated into the aromatic ring of the ^{2}H phenylalanine molecule as metabolism of adapted to $^{2}\text{H}_2\text{O}$ bacterium does not undergo essential changes in $^{2}\text{H}_2\text{O}$. The obtained result on distribution of atoms of deuterium into ^{2}H phenylalanine molecule is important for its further use in medical diagnostics where it is necessary to use ^{2}H amino acids with high levels of isotope enrichment.

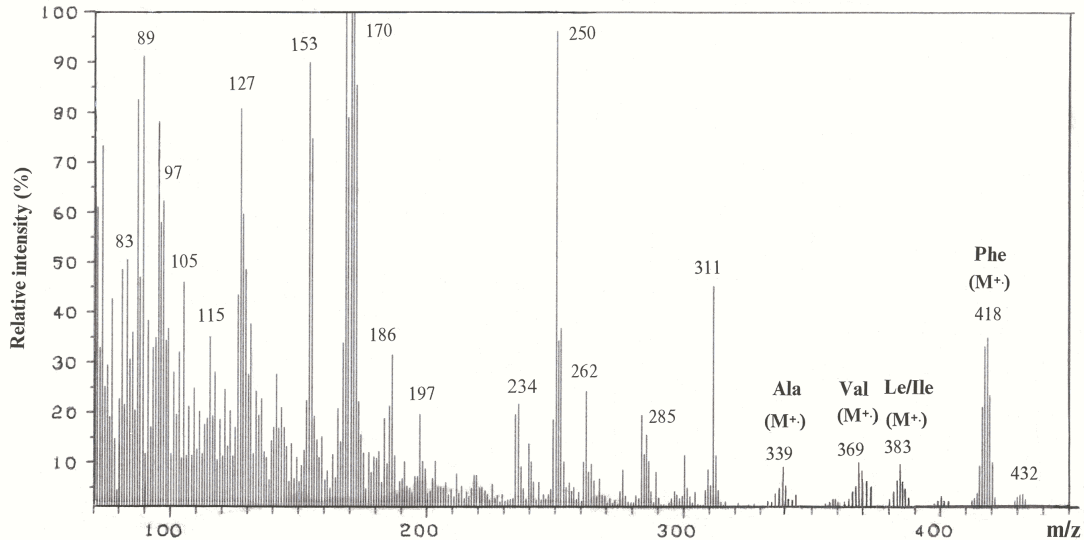


Figure 7. EI mass spectrum of methyl esters of N-DNS- ^2H amino acids from growth medium M9 containing 2% (v/v) $[\text{U-}^2\text{H}]\text{MeOH}$ and 98% (v/v) $^2\text{H}_2\text{O}$ (expt. 10, Table 1).

The similar result on proportional specific increase of levels of deuterium enrichment into ^2H phenylalanine and other metabolically related ^2H amino acids (alanine, valine and leucine/isoleucine) was observed in all isotopic experiments where used increasing concentration $^2\text{H}_2\text{O}$ in growth media (Table 2). Predictably, enrichment levels of ^2H phenylalanine related to the family of aromatic amino acids synthesised from shikimic acid and metabolically related ^2H amino acids of pyruvic acid family – alanine, valine and leucine at identical $^2\text{H}_2\text{O}$ concentration in growth media are correlated among themselves. Such result is fixed in all isotope experiments with $^2\text{H}_2\text{O}$ (Table 2). Unlike ^2H phenylalanine, deuterium enrichment levels in accompanying ^2H amino acids – Ala, Val and Leu/Ile keep a stable constancy within a wide interval of $^2\text{H}_2\text{O}$ concentration: from 49% (v/v) to 98% (v/v) $^2\text{H}_2\text{O}$ (Table 2). Summarizing these data, it is possible to draw a conclusion on preservation of minor pathways of the metabolism connected with biosynthesis of leucine and metabolic related amino acids of pyruvic acid family – alanine and valine, which enrichment levels were in correlation within identical concentration of H_2O in growth media (phenylalanine is related to the family of aromatic amino acids synthesized from shikimic acid). Since leucine was added into growth media in protonated form, another explanation of this effect, taking into consideration the various biosynthetic pathways of Leu and Ile (Ile belongs to the family of aspartic acid, while Leu belongs to the pyruvic acid family), could be cell assimilation of protonated leucine from growth media. Since Leu and Ile could not be clearly estimated by EI MS method, nothing could be said about possible biosynthesis of ^2H isoleucine. Evidently, higher levels of deuterium enrichment can be achieved by replacement of protonated leucine on its deuterated analogue, which may be isolated from hydrolysates of deuterated biomass of this methylotrophic bacterium.

Table 2. Effect of deuterium enrichment levels (atom%) in the molecules of ^2H amino acids excreted by *B*.

*methylicum**

[² H]amino acid	Concentration of ² H ₂ O in growth media, % (v/v)**			
	24.5	49.0	73.5	98.0
Alanine	24.0±0.70	50.0±0.89	50.0±0.83	50.0±1.13
Valine	20.0±0.72	50.0±0.88	50.0±0.72	62.5±1.40
Leucine/isoleucine	20.0±0.90	50.0±1.38	50.0±1.37	50.0±1.25
Phenylalanine	17.0±1.13	27.5±0.88	50.0±1.12	75.0±1.40

Notes:

* At calculation of enrichment levels protons (deuterons) at COOH- and NH₂-groups of amino acids were not considered because of dissociation in H₂O (²H₂O).

** The data on enrichment levels described bacteria grown on minimal growth media M9 containing 2% (v/v) [U-²H]MeOH and specified amounts (% , v/v) of ²H₂O.

4. Conclusions

As a result of the selection approach used in this research it was possible to adapt L-phenylalanine producer strain of aerobic Gram-positive facultative methylotrophic bacterium *B. methylicum* to maximal concentration of deuterated substrates for microbiological preparation of [²H]phenylalanine and other metabolically related [²H]amino acids (alanine, valine and leucine/isoleucine) with various levels of deuterium enrichment. Advantages of this methylotroph for synthesis of [²H]amino acids are improved growth and biosynthetic characteristics on maximally deuterated growth medium, which was achieved by adaptation to ²H₂O. By using the adapted methylotroph it was possible to obtain 0.82 g/liter of [²H]phenylalanine (75% ²H), which was isolated from growth medium by extraction with iso-PrOH and the subsequent crystallization in EtOH. [²H]phenylalanine was also isolated from growth medium by reversed-phase HPLC as methyl ester of N-DNS-[²H]phenylalanine with a yield of 85% and a purity of 97%. The method is suitable for preparation of other [²H]amino acids produced by methylotrophic bacteria. It should be noted, however, that higher levels of deuterium enrichment into [²H]amino acid molecule can be achieved via replacement of protonated leucine on its deuterated analogue.

Acknowledgements

The authors wish to thank V.I. Shvets and D.A. Skladnev for helpful discussions and valuable remarks.

References

- Abou-Zeid, A., Euverink, G., Hessels, G.I., Jensen, R.A. & Dijkhuizen, L. (1995) Biosynthesis of L-phenylalanine and L-tyrosine in the actinomycete *Amycolatopsis methanolica*, *Applied and Environmental Microbiology*, **61**(4): 1298–302.
- Antony, C. (1986) Bacterial oxidation of methane and methanol. *Advances in Microbial Physiology*, **27**: 113–210.
- Antony, C. & Williams, P.W. (2003) The structure and function of methanol dehydrogenase. *Biochimica et Biophysica Acta*, **1467**: 18–23.
- Blomquist, A.T., Cedergren, R.J., Hiscock, B.F., Tripp, S.L. & Harp, D.N. (1966) Synthesis of highly deuterated amino acids, *PNAS*, **55**(3): 453–456.
- Bohinski, R.C. (1983) *Modern Concepts in Biochemistry*. Boston, London, Sydney, Toronto, Allyn &

- Bacon Inc., Newton, Massachusetts, pp. 1-378.
- Crespi, H.L. (1986) Biosynthesis and uses of per-deuterated proteins, in: R.R. Muccino, (Ed.) *Synthesis and Applications of Isotopically labeled Compounds*, Proceedings of the 2nd Intern Symp, Kansas City, Missouri, USA, 3–6 September 1985, Amsterdam-Oxford-New York, Elsevier, pp. 111–112.
- Conn, E.E. (1986) Recent Advances in Phytochemistry, in: E.E. Conn, (Ed.) *The Shikimic Acid Pathway*, 2nd ed., Plenum Press, New York, pp. 20–22.
- Cox, J. & Kyli, D. (1988) Stable isotope labeled biochemicals from microalgae. *Trends in Biotechnology*, **6**: 279–282.
- de Boer, L., Harder, W. & Dijkhuizen, L. (1988) Phenylalanine and tyrosine metabolism in the facultative methylotroph *Nocardia sp.* 239. *Archives of Microbiology*, **149**: 459–465.
- Den'ko, E.I. (1970) Influence of heavy water (D₂O) on animal, plant and microorganism's cells. *Usp. Sovrem. Biol.*, **70**(4): 41–49 [in Russian].
- Dewick, P.M. (2009) in: *Medicinal Natural Products: A Biosynthetic Approach*. – NY: John Wiley & Sons Ltd, pp. 137–186.
- DiLella, A.G., Kwok, S.C.M., Ledley, F.D., Marvit, J., & Woo, S.L. C. (1986). Molecular structure and polymorphic map of the human phenylalanine hydroxylase gene. *Biochemistry*, **25**(4): 743–749. doi:10.1021/bi00352a001.
- Faleev, N.G., Ruvinov, S.B., Saporovskaya, M.B., Belikov, V.M. & Zakomyrdina, L.N. (1989) Preparation of α -deuterated amino acids by *E. coli* cells containing tryptophanase. *Izv. Akad. Nauk. USSR Ser. Khim.*, **10**: 2341–2343 [in Russian].
- Filiano, J.J. (2006). Neurometabolic Diseases in the Newborn. *Clinics in Perinatology*, **33**(2): 411–479. doi:10.1016/j.clp.2006.03.013.
- Herrmann, K.M & Weaver, L.M. (1999) The shikimate pathway. *Annual Review of Plant Physiology and Plant Molecular Biology*, **50**: 473–503.
- Ignatov, I. (2005) *Energy Biomedicine*, Gea-Libris, Sofia, 1–88.
- Ignatov, I. (2010) Which water is optimal for the origin (generation) of life? *Euromedica*, Hanover: 34-35.
- Ignatov, I. (2011) Entropy and time in living matter, *Euromedica*: 74.
- Ignatov, I. (2012) Origin of Life and Living Matter in Hot Mineral Water, Conference on the Physics, Chemistry and Biology of Water, Vermont Photonics, USA.
- Ignatov, I., & Mosin, O.V. (2012) Isotopic Composition of Water and its Temperature in Modeling of Primordial Hydrosphere Experiments, VIII Intern. Conference Perspectives of the Development of Science and Technique, *Biochemistry and Biophysics*, **15**: 41–49.
- Ignatov, I., Mosin, O. V. & Naneva, K. (2012) Water in the Human Body is Information Bearer about Longevity, *Euromedica*, Hanover: 110-111.
- Ignatov I., Mosin O.V. (2013) Possible Processes for Origin of Life and Living Matter with Modeling of Physiological Processes of Bacterium *Bacillus Subtilis* in Heavy Water as Model System, *Journal of Natural Sciences Research*, **3** (9): 65-76.
- Ignatov, I., Mosin, O. V. (2013) Modeling of Possible Processes for Origin of Life and Living Matter in Hot Mineral and Seawater with Deuterium, *Journal of Environment and Earth Science*, **3**(14): 103-118.
- Ignatov, I., Mosin, O. V. (2013) Structural Mathematical Models Describing Water Clusters, *Journal of Mathematical Theory and Modeling*, **3** (11): 72-87.
- Ignatov, I., Mosin, O. V. (2014) The Structure and Composition of Carbonaceous Fullerene Containing Mineral Shungite and Microporous Crystalline Aluminosilicate Mineral Zeolite. Mathematical Model of Interaction of Shungite and Zeolite with Water Molecules *Advances in Physics Theories and Applications*, **28**: 10-21.
- Ignatov, I., Mosin, O.V., Velikov, B., Bauer, E. & Tyminski, G. (2014) Longevity Factors and Mountain

Water as a Factor. Research in Mountain and Field Areas in Bulgaria, *Civil and Environmental Research*, **6** (4): 51-60.

Ignatov, I., Mosin, O. V., Niggli, H.&Drossinakis, Ch. (2014) Evaluating Possible Methods and Approaches for Registering of Electromagnetic Waves Emitted from the Human Body, *Advances in Physics Theories and Applications*, **30**: 15-33.

Ignatov, I., Mosin, O.V.&Drossinakis, Ch. (2014) Infrared Thermal Field Emitted from Human Body. Thermovision, *Journal of Medicine, Physiology, Biophysics*, **1**:1-12.

Ignatov, I., Mosin, O.V.&Velikov, B. (2014) Longevity Factors and Mountain Water of Bulgaria in Factorial Research of Longevity, *Journal of Medicine, Physiology, Biophysics*,**1**:13-33.

Ignatov, I.&Mosin,O.V. (2014) Visual Perception. Electromagnetic Conception for the Eyesight. Rhodopsin and Bacteriodopsin, *Journal of Medicine, Physiology and Biophysics*, **2**:1-19.

Ignatov, I.&Mosin,O.V. (2014) The Structure and Composition of Shungite and Zeolite. Mathematical Model of Distribution of Hydrogen Bonds of Water Molecules in Solution of Shungite and Zeolite, *Journal of Medicine, Physiology and Biophysics*, **2**: 20-36.

Ignatov, I., Mosin,O.V., Velikov, B., Bauer, E.&Tyminski, G. (2014) Research of Longevity Factors and Mountain Water as a Factor in Teteven Municipality, Bulgaria, *Journal of Medicine, Physiology and Biophysics*, **2**: 37-52.

Ignatov, I.&Mosin,O.V. (2014) Modeling of Possible Processes for Origin of Life and Living Matter in Hot Mineral Water. Research of Physiological Processes of Bacterium *Bacillus Subtilis* in Hot Heavy Water, *Journal of Medicine, Physiology and Biophysics*, **2**: 53-70.

Ignatov, I.&Mosin,O.V. (2014) Mathematical Models of Distribution of Water Molecules Regarding Energies of Hydrogen Bonds, *Medicine, Physiology and Biophysics*, **2**: 71-94.

Karnaukhova, E.N., Mosin, O.V. & Reshetova. O.S. (1993) Biosynthetic production of stable isotope labeled amino acids using methylotroph *Methylobacillus flagellatum*. *Amino Acids*, **5**(1): 125–126.

Kushner, D.I., Baker, A. & Dunstall, T.G. (1999) Pharmacological uses and perspectives of heavy water and deuterated compounds. *Canadian Journal of Physiology and Pharmacology*, **77**(2): 79–88.

LeMaster, D.M. (1990) Deuterium labeling in NMR structural analysis of larger proteins. *Quarterly Reviews of Biophysics*, **23**: 133–174.

Lindstrom, M.E. & Stirling, D.I. (1990) Methylotrophs: genetics and commercial applications. *Annual Review of Microbiology*, **4**: 27–58.

Maksimova, N.P., Dobrozhinetskaia, E.V. & Fomichev, Iu.K. (1990) Regulation of phenylalanine biosynthesis in the obligate methylotroph *Methylobacillus* M75. *Mol. Gen. Microbiol. Virusol.*, **10**: 28–30.

Michals, K., Matalon, R. (1985). Phenylalanine metabolites, attention span and hyperactivity. *American Journal of Clinical Nutrition*, **42**(2): 361–365.

Mosin, O.V.(1996) Studying of methods for biotechnological preparation of amino acids, protein and nucleosides labeled with stable isotopes ^2H and ^{13}C with high levels of isotopic enrichment, Cand. Diss. (PhD) Thesis, M.V. Lomonosov State Academy of Fine Chemical Technology, Moscow, May 1996.

Mosin, O.V., Skladnev, D.A., Egorova, T.A. & Shvets, V.I. (1996a) Mass-spectrometric determination of levels of enrichment of ^2H and ^{13}C in molecules of amino acids of various bacterial objects. *Bioorganic Chemistry*, **22**(10–11) : 56–869.

Mosin, O.V., Skladnev, D.A., Egorova, T.A., Yurkevitch, A.M. & Shvets, V.I. (1996b) Studying of biosynthesis of amino acids by a strain of *Brevibacterium methylicum* at growth on media, containing heavy water and deuterium-methanol. *Biotechnologija*, **3**: 3–12 [in Russian].

Mosin, O.V., Skladnev, D.A. & Shvets, V.I. (1998) Biosynthesis of ^2H -labeled phenylalanine by a new methylotrophic mutant *Brevibacterium methylicum*. *Bioscience, biotechnology and biochemistry*, **62**(2):

225–229.

Mosin, O.V. & Ignatov, I. (2012) Isotope effects of deuterium in bacterial and microalgae cells at growth on heavy water (D₂O). *Water: Chemistry and Ecology*, **3**: 83–94 [in Russian].

Mosin, O.V., Shvets, V.I., Skladnev, D.A. & Ignatov, I. (2012) Studying of microbic synthesis of deuterium labeled L-phenylalanine by methylotrophic bacterium *Brevibacterium methylicum* on media with different content of heavy water. *Russian Journal of Biopharmaceuticals*, **4**: 11–22 [in Russian].

Ignatov, I., Mosin, O. V. (2012) Isotopic Composition of Water and its Temperature in Modeling of Primordial Hydrosphere Experiments, *Euro-Eco*, Hanover: 62.

Mosin, O.V., Shvets, V.I., Skladnev, D.A., & Ignatov, I. (2013) Microbiological synthesis of [²H]inosine with high degree of isotopic enrichment by Gram-positive chemoheterotrophic bacterium *Bacillus subtilis*. *Applied Biochem. Microbiol.*, **49**(3): 233–243.

Mosin, O.V., Shvets, V.I., Skladnev, D.I. & Ignatov, I. (2013) Microbial synthesis of ²H-labelled L-phenylalanine with different levels of isotopic enrichment by a facultative methylotrophic bacterium *Brevibacterium methylicum* with RuMP assimilation of carbon. *Biochemistry (Moscow) Supplement Series B: Biomedical Chemistry*, **7**(3): 249-260.

Mosin, O.V. & Ignatov, I. (2013a) Studying of the biosynthesis of ²H-labeled inosine by a Gram-positive chemoheterotrophic bacterium *Bacillus subtilis* B-3157 on heavy water (²H₂O) medium. *Chemical and Process Engineering Research*, **15**: 32–45.

Mosin, O.V. & Ignatov, I. (2013b) Microbiological synthesis of ²H-labeled phenylalanine, alanine, valine, and leucine/isoleucine with different degrees of deuterium enrichment by the Gram-positive facultative methylotrophic bacterium *Brevibacterium methylicum*. *International Journal of Biomedicine*, **3**(2): 132–138.

Mosin, O. V., Ignatov, I., Skladnev, D. A & Shvets, V. I. (2013) Use of Gram-positive Chemoheterotrophic Bacterium *Bacillus Subtilis* B-3157 with HNP-cycle of Carbon Assimilation for Microbiological Synthesis of [2H] riboxine with High Level of Deuterium Enrichment, *European Journal of Molecular Biotechnology*, **2**: 63-78.

Mosin, O.V. & Ignatov, I. (2014) Biosynthesis of photochrome transmembrane protein bacteriorhodopsin of *Halobacterium halobium* labeled with deuterium at aromatic amino acids residues of 2,3,4,5,6-²H₅]Phe, [3,5-²H₂]Tyr and [2,4,5,6,7 -²H₅]Trp. *Chemistry and Materials Research*, **6**(3): 38-48.

Skladnev, D.A., Mosin, O.V., Egorova, T.A., Eremin, S.V. & Shvets, V.I. (1996) Methylotrophic bacteria are sources of isotopically labelled ²H- and ¹³C-amino acids. *Biotechnologija*, **5**: 25–34 [in Russian].

Surtees, R., Blau, N. (2000). The neurochemistry of phenylketonuria. *European Journal of Pediatrics*, **169**: 109–113. [doi:10.1007/PL00014370](https://doi.org/10.1007/PL00014370).

V értes, A., Nagy, S., Klencs ár, Z., Lovas, R.G. & Rosh, F. (2011) *Handbook of Nuclear Chemistry, Elements and Isotopes: Formation, Transformation, Distribution*, 2nd ed., in: A. V értes, S. Nagy and Z. Klencs ár, (Eds.) Springer, Dordrecht, Helderberg, London, New York, Berlin, Vol. 2, pp. 699-727.

Wendisch, V.F. (2007) *Amino Acid Biosynthesis – Pathways, Regulation and Metabolic Engineering*, in: F. Volker, (Ed.) Springer Verlag, Helderberg, pp. 237–240.

Walker, T.E. & Matheny, C. (1986) An efficient chemomicrobiological synthesis of stable isotope-labeled L-tyrosine and L-phenylalanine. *Journal of Organic Chemistry*, **51**: 1175–1179.

The IISTE is a pioneer in the Open-Access hosting service and academic event management. The aim of the firm is Accelerating Global Knowledge Sharing.

More information about the firm can be found on the homepage:
<http://www.iiste.org>

CALL FOR JOURNAL PAPERS

There are more than 30 peer-reviewed academic journals hosted under the hosting platform.

Prospective authors of journals can find the submission instruction on the following page: <http://www.iiste.org/journals/> All the journals articles are available online to the readers all over the world without financial, legal, or technical barriers other than those inseparable from gaining access to the internet itself. Paper version of the journals is also available upon request of readers and authors.

MORE RESOURCES

Book publication information: <http://www.iiste.org/book/>

IISTE Knowledge Sharing Partners

EBSCO, Index Copernicus, Ulrich's Periodicals Directory, JournalTOCS, PKP Open Archives Harvester, Bielefeld Academic Search Engine, Elektronische Zeitschriftenbibliothek EZB, Open J-Gate, OCLC WorldCat, Universe Digital Library, NewJour, Google Scholar

