

Investigation of Mutagenic Effects of Pea Green and Anti-carcinogenic Effects of Several Plants Extracts in *Drosophila Melanogaster* Using Wing Spot

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Abstract

In the present study, Pea green (E 102 - E 133) is coloring additive for its genotoxic effects and expected antigenotoxic effects of several plant extracts; *Hypericum perforatum* L., *Silybum marianum* (L.) Gaertn., *Lavandula stoechas* L. against the food coloring were examined using Wing spot test on *Drosophila melanogaster* in the laboratory experiments. The *Drosophila* lar-vae were chronically fed in food mediums where Pea green at 25 g/L, 50 g/L, and 75 g/L concentrations were experimentally applied. The results showed that Pea green has highly significant results at different exposure concentrations in trans-heterozygous flies (mwh/flr3) and balanc-er-heterozygous flies (mwh/TM3) for inducing the numbers of mutant wing spots compared to control groups (Chi-Square test; df=3, P<0.001). Thus, it seemed that the food coloring is a highly mutagenic agent on *Drosophila melanogaster* in the laboratory conditions. On the other hand, the concentrations of Pea green were used together different plant extracts at 100 g/L concentra-tion in the mediums. The numbers of mutant wing spots were compared to control groups showed that the three plant extracts have not effects to reduce the mutant spots in mwh/flr3 and mwh/TM3 flies and thus they have not antigenotoxic effects used experimental treatments (Kruskal-Wallis test; df=2, P>0.05)

Keywords: Genotoxicity, antigenotoxicity; Pea green; *Hypericum perforatum* L.; *Silybum marianum* (L.) Gaertn., *Lavandula stoechas* L.; *Drosophila melanogaster* Meigen; Wing spot test

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1. Introduction

With the new technologies used in the food industry and the increasing variety of products, the interest in ready-made foods is increasing day-by-day with consumer demand (AKBULUT 2011). Food additives are frequently found in ready-to-eat foods and are among the potentially important genotoxic agents (Vural 2005; Alkan and Anlas 2015). In addition to the nutritional values of consumed food, its safety and effects on human health are nowadays widely discussed (Alkan and Anlas 2015).

Food coloring agents can create allergic reactions, skin rashes, asthma, hyperactivity and concentration disorder, hypersensitivity, migraine, preterm delivery, salicylate sensitivity and cancer when not used within legal restrictions (Yentür 1996; Maier et al. 2010).

With genotoxicity tests and epidemiological studies on *Drosophila*, mouse, rat, and bacteria, when not used within the limits specified in regulations, the synthetic food colorings have been reported to be carcinogenic when used in order to imitate the high quality (Yentür 1996; Sarıkaya 2010). It has been determined that the level of food colorings used in studies on this subject in our country is well above the statute limits and may have harmful effects in terms of public health (Yentür 1996; Topsoy 1990; Yaman 1996).

Pea green used in the present study was preferred because, in the literature, there are very few scientific studies similar to the subject of the present research. There is no study carried out on Pea green in *Drosophila* by using Wing spot test. There are few studies carried out on *Drosophila* regarding the genotoxicity of the food coloring. In addition, the plant extracts, widely used among the public and thought to eliminate mutagenic effects, used in the present study was preferred because, in the literature, there is no study carried out on the plant extracts in

Drosophila by using Wing spot test. In *Drosophila*, while there are few scientific studies on the antigenotoxicity of *Hypericum perforatum* L., there is no study carried out on *Silybum marianum* (L.) Gaertn. and *Lavandula stoechas* L..

Using Wing spot test on *D. melanogaster* Meigen's *mwh* and *flr3* lines, the present research was conducted in order to determine whether Pea green at 25 g/L, 50 g/L, 75 g/L concentrations have a detrimental effect, *Hypericum perforatum* L., *Silybum marianum* (L.) Gaertn. and *Lavandula stoechas* L. at 100 g/L concentration be successful in reducing the genotoxic effect of the coloring or not.

2. Materials and Methods

2.1. Culture of *Drosophila* Lines

D. melanogaster Meigen lines used in the study were obtained from Trakya University and Akdeniz University, Science Faculty, Biology Department. *Drosophila* flies were cultured in an incubator where optimum living conditions (25 ± 1 °C and 40-60 % relative humidity) were provided and adjusted to 12-hour light and 12-hour dark time period.

2.2. Creation of Experimental Groups

In determination the mutagenic and/or recombinogenic effects of Pea green and antigenotoxic effects of the plant extracts against the coloring in *D. melanogaster* Meigen lines SMART developed by (Graf et al. 1984) was used.

2.3. Genetic Structure of *Drosophila* Mutant Strains

In *Drosophila* Wing spot test, the *mwh*, *flr3* and *BdS* (beader-serrate) marker genes on the third largest chromosome of *Drosophila* are used. In *Drosophila* Wing spot test, *flr3/TM3*, *BdS* virgin females and *mwh/mwh* males are used for normal metabolic activity cross (Lindsley and Grell 1968; Lindsley and Zimm 2012; Garcia-Bellido and Dapena 1974).

Experiments of the application groups consisting of experimental, treatment and control groups in the study were conducted simultaneously. Distilled water was used in the negative control group and EMS (Kasimoğlu 2016) is stated in the literature that it has a mutagenic effect, was used at 1 mM concentration in the positive control group. The food coloring and the plant extracts used in the application groups were dissolved in distilled water. Pea green at 25 g/L, 50 g/L and 75 g/L concentrations in the experimental group and *Hypericum perforatum* L., *Silybum marianum* (L.) Gaertn. and *Lavandula stoechas* L. at 100 g/L concentration with the concentrations of the coloring were used for *Drosophila* larvae at 72 ± 4 hours.

D. melanogaster Meigen lines were fed with a standard *Drosophila* medium when kept in stock and in a cross bottle to perform fertilization and embryogenesis to obtain heterozygous larvae. *Drosophila* instant medium was used for *Drosophila* larvae at 72 ± 4 hours in the experimental, treatment and control groups. *Drosophila* instant medium and the food coloring and the plant extracts used in the study were purchased commercially. *Drosophila* instant medium was obtained from Carolina Biological Supply Company. Pea green was from Roha JJT Group Company. *Hypericum perforatum* L. (St. John's wort) and *Silybum marianum* (L.) Gaertn. (milk thistle) were from Solgar and *Lavandula stoechas* L. (topped lavender) was from Herbal Pharm and EMS was obtained from Sigma Aldrich GmbH.

2.4. Applications of Experimental Groups

Larvae in groups of 100 were left into falcon tubes where 1.5 g of *Drosophila* instant medium wetted with 5 ml of solutions containing at 25 g/L, 50 g/L and 75 g/L concentrations of Pea green and/or the plant extracts at 100 g/L concentration. While the wing edges of the adult flies developed from trans-heterozygous larvae (*mwh/flr3*) in each concentration of application groups have normal structure, the wing edges of the balancer-heterozygous flies (*mwh/TM3*) are in the form of serrate. In evaluation, 40 wings were used for each concentration in all application groups. Slides of normal and serrate wings were examined using a light microscope under 40×10 magnification to detect the presence of mutant clones (Kaya 2000). Classification of types were according to mutant clones: Small single spots containing of 1-2 *mwh*, large single spots containing ≥ 3 *mwh* or ≥ 4 *flr3* and twin spots containing adjacent *mwh* and *flr3* cells (Graf et al. 1984).

2.5. Statistical Analysis

Minitab package program was used in the statistical evaluation of the study results. In the analysis of the study data, it was investigated whether the total numbers of mutant spots in the wings of the flies in the positive and negative control groups was statistically different with the Chi-Square test ($df=1$). Chi-Square test ($df=3$) was

used in the analyzes where the total numbers of mutant spots in the wings of the flies in experimental and treatment groups were compared separately with those of the flies in the positive and negative control groups. The total numbers of mutant spots in the wings of the flies in experimental and treatment groups were compared with each other and whether there was a statistical difference was determined by the Chi-Square test (df=2).

Possible differences between the average of the total numbers of wing mutant spots of flies grown in experimental and treatment groups were compared with Non-Parametric Kruskal-Wallis test (df=2).

The average of the total numbers of mutant spots in the normal and serrate wings of the flies in all experimental groups were compared with the Mann-Whitney U test.

In all evaluation, the total numbers of wing mutant spots obtained from the study and presented as Fr (Frequency) values and the differences below $P < 0.05$ were considered to be statistically significant.

3. Results

Table 1. Mutant clones in *mwh/flr3* wings obtained from experimental group, treatment group, positive and negative control groups

Application groups / Concentration (g/L)	Number of wings	Small single spots (1-2 cell)		Large single spots (>2 cell)		Twin spots		Total <i>mwh</i> spots		Total spots X ²		
		No	Fr	No	Fr	No	Fr	No	Fr	No	Fr	
Distilled water	40	3	(0.08)	5	(0.13)	4	(0.13)	3	(0.08)	12	(0.30)	
1 mM EMS	40	49	(1.23)	130	(3.25)	68	(1.70)	99	(2.48)	247	(6.18)	a***
25 Pea green	40	1	(0.03)	40	(1.00)	0	(0.00)	1	(0.03)	41	(1.03)	a*** b***
50 Pea green	40	1	(0.03)	57	(1.43)	0	(0.00)	1	(0.03)	58	(1.45)	a*** b***
75 Pea green	40	3	(0.08)	59	(1.48)	0	(0.00)	4	(0.10)	62	(1.55)	a*** b***
25 Pea green- <i>H. perforatum</i>	40	2	(0.05)	51	(1.28)	0	(0.00)	2	(0.05)	53	(1.33)	a*** b***
50 Pea green- <i>H. perforatum</i>	40	3	(0.08)	46	(1.15)	0	(0.00)	3	(0.08)	49	(1.23)	a*** b***
75 Pea green- <i>H. perforatum</i>	40	3	(0.08)	59	(1.48)	0	(0.00)	5	(0.13)	62	(1.55)	a*** b***
25 Pea green- <i>S. marianum</i>	40	6	(0.15)	31	(0.78)	2	(0.05)	12	(0.30)	39	(0.98)	a*** b***
50 Pea green- <i>S. marianum</i>	40	6	(0.15)	31	(0.78)	0	(0.00)	7	(0.18)	37	(0.93)	a*** b***
75 Pea green- <i>S. marianum</i>	40	2	(0.05)	45	(1.13)	2	(0.05)	5	(0.13)	49	(1.23)	a*** b***
25 Pea green- <i>L. stoechas</i>	40	0	(0.00)	27	(0.68)	0	(0.00)	0	(0.00)	27	(0.68)	a*** b*** c**
50 Pea green- <i>L. stoechas</i>	40	4	(0.13)	47	(1.18)	1	(0.03)	6	(0.15)	52	(1.30)	a*** b*** c**
75 Pea green- <i>L. stoechas</i>	40	2	(0.05)	56	(1.40)	0	(0.00)	3	(0.08)	58	(1.45)	a*** b*** c**
H, P [#]											3.77, >0.05	
P [§]											<0.001	

No: Number of mutant clones, Fr: Frequency; #: It was evaluated by Kruskal-Wallis test;

§: It was evaluated by Mann-Whitney U test; X2: In evaluation with Chi-Square test; a: with Distilled water, b:

with EMS, c: with Pea green; *: P<0.05, **: P<0.01, ***: P<0.001

Table 2. Mutant clones in mwh/TM3 wings obtained from experimental group, treatment group, positive and negative control groups

Application groups / Concentration (g/L)	Number of wings	Small single spots (1-2 cell)		Large single spots (>2 cell)		Twin spots		Total mwh spots		Total spots X ²		
		No	Fr	No	Fr	No	Fr	No	Fr	No	Fr	
Distilled water	40	1	(0.03)	6	(0.15)	0	(0.00)	1	(0.03)	7	(0.18)	
1 mM EMS	40	22	(0.55)	66	(1.65)	0	(0.00)	71	(1.78)	88	(2.20)	a***
25 Pea green	40	4	(0.03)	32	(0.80)	0	(0.00)	4	(0.10)	36	(0.90)	a*** b***
50 Pea green	40	1	(0.00)	31	(0.78)	0	(0.00)	1	(0.03)	32	(0.80)	a*** b***
75 Pea green	40	5	(0.08)	38	(0.95)	0	(0.00)	5	(0.13)	43	(1.08)	a*** b***
25 Pea green- <i>H. perforatum</i>	40	1	(0.05)	23	(0.58)	0	(0.00)	1	(0.03)	24	(0.60)	a*** b***
50 Pea green- <i>H. perforatum</i>	40	0	(0.13)	28	(0.70)	0	(0.00)	0	(0.00)	28	(0.70)	a*** b***
75 Pea green- <i>H. perforatum</i>	40	4	(0.03)	31	(0.78)	0	(0.00)	5	(0.13)	35	(0.88)	a*** b***
25 Pea green- <i>S. marianum</i>	40	1	(0.00)	30	(0.75)	0	(0.00)	1	(0.03)	31	(0.78)	a*** b***
50 Pea green- <i>S. marianum</i>	40	3	(0.03)	30	(0.75)	0	(0.00)	3	(0.08)	33	(0.83)	a*** b***
75 Pea green- <i>S. marianum</i>	40	0	(0.03)	26	(0.65)	0	(0.00)	0	(0.00)	26	(0.65)	a*** b***
25 Pea green- <i>L. stoechas</i>	40	1	(0.00)	23	(0.58)	0	(0.00)	3	(0.08)	24	(0.60)	a*** b***
50 Pea green- <i>L. stoechas</i>	40	2	(0.03)	24	(0.60)	0	(0.00)	3	(0.08)	27	(0.68)	a*** b***
75 Pea green- <i>L. stoechas</i>	40	2	(0.00)	33	(0.83)	0	(0.00)	2	(0.05)	35	(0.88)	a*** b***
H, P [#]											3.85, >0.05	
P [§]											<0.001	

No: Number of clones, Fr: Frequency, #: It was evaluated by Kruskal-Wallis test;

§: It was evaluated by Mann-Whitney U test; X²: In evaluation with Chi-Square test; a: with Distilled water, b: with EMS, c: with Pea green, *: P<0.05, **: P<0.01, ***: P<0.001

When the results of the study were evaluated, the total numbers of mutant spots in normal (X²=213.22, df=1, P<0.001) wings (in Table 1) and serrate (X²=69.06, df=1, P<0.001) wings (in Table 2) of *Drosophila* flies in the positive control group was found to be much more than those of negative control group.

Wing spot test data obtained from experimental group studies of Pea green were compared with results of positive and negative control groups. The total numbers of mutant spots in the normal wings (in Table 1) and serrate wings (in Table 2) of the *Drosophila* flies grown in the containing with 25 g/L, 50 g/L and 75 g/L concentrations of the Pea green is higher than those grown in the medium prepared with distilled water (normal wings: X²=35.85, df=3, P<0.001; serrate wings: X²=24.98, df=3, P<0.001) but it was found to be less than those grown in medium with EMS addition (normal wings: X²=277.27, df=3, P<0.001; serrate wings: X²=40.45, df=3, P<0.001).

The total numbers of mutant spots in the normal and serrate wings of the *Drosophila* flies grown in the mediums having three different concentrations of Pea green with *Hy-pericum perforatum* L. at 100 g/L concentration is

higher than grown that of the flies in the medium prepared with distilled water (normal wings in Table 1: $X^2=33.04$, $df=3$, $P<0.001$; serrate wings in Table 2: $X^2=18.08$, $df=3$, $P<0.001$) but it was found to be less than those grown in medium added to EMS (normal wings in Table 1: $X^2=270.87$, $df=3$, $P<0.001$; serrate wings in Table 2: $X^2=61.09$, $df=3$, $P<0.001$).

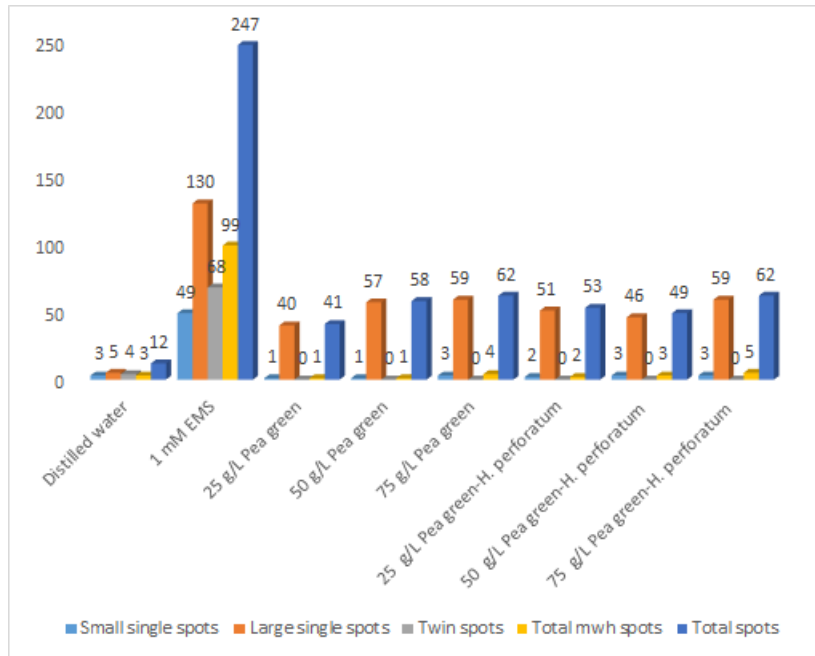


Figure 1. shows classification of mutant clones with the normal wings of the *Drosophila* flies in the experimental group, treatment group with *Hypericum perforatum* L. and the positive with negative control groups.

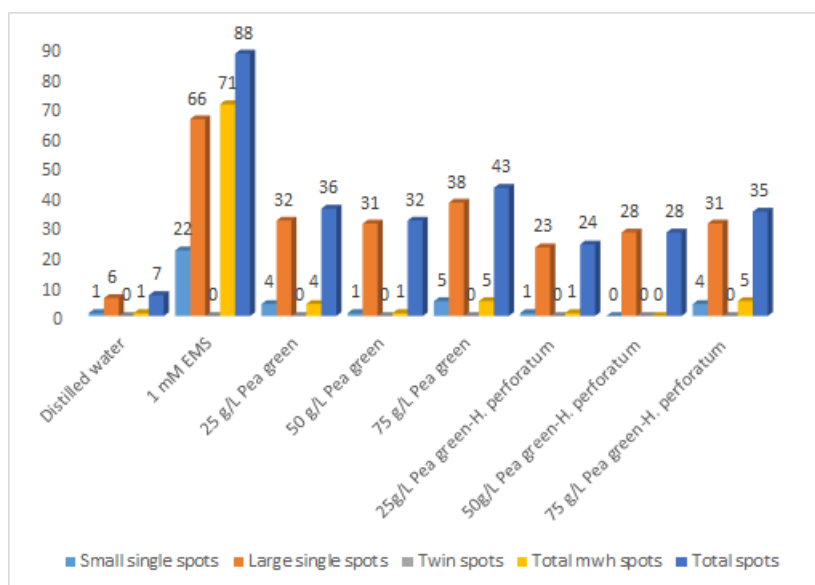


Figure 2. shows classification of mutant clones with the serrate wings of the *Drosophila* flies in the experimental group, treatment group with *Hypericum perforatum* L. and the positive with negative control groups.

The total numbers of mutant spots in the normal and serrate wings of the *Drosophila* flies grown in the mediums with three different concentrations of Pea green and 100 g/L concentration of *Silybum marianum* (L.) Gaertn are more than that of the flies grown in medium containing distilled water (normal wings in Table 1: $X^2=21.68$, $df=3$, $P<0.001$; serrate wings in Table 2: $X^2=17.43$, $df=3$, $P<0.001$) but it was found to be lower than those grown in medium with EMS (normal wings in Table 1: $X^2=340.90$, $df=3$, $P<0.001$; serrate wings in Table 2: $X^2=57.28$,

df=3, P<0.001).

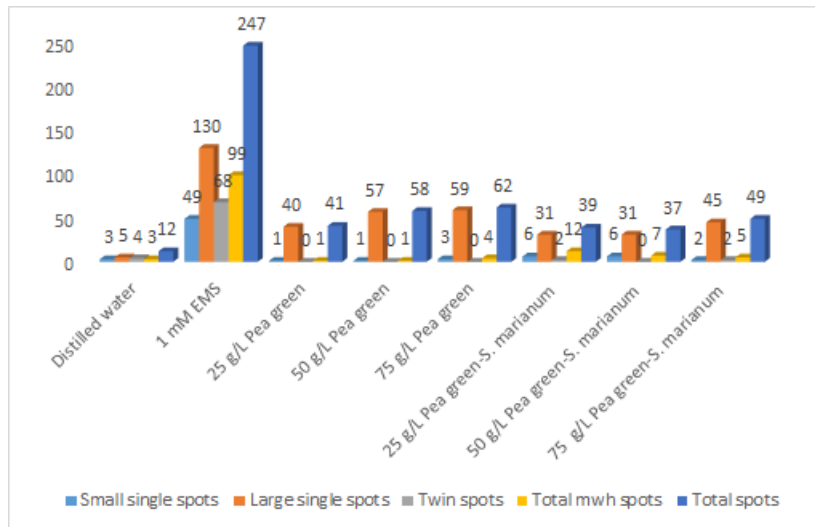


Figure 3. shows classification of mutant clones with the normal wings of the *Drosophila* flies in the experimental group, treatment group with *Silybum marianum* (L.) Gaertn. and the positive with negative control groups.

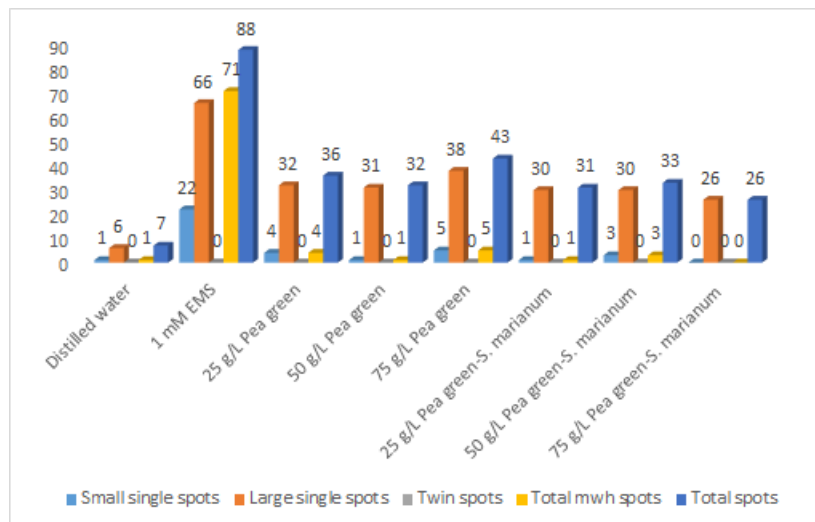


Figure 4. shows classification of mutant clones with the serrate wings of the *Drosophila* flies in the experimental group, treatment group with *Silybum marianum* (L.) Gaertn. and the positive with negative control groups.

The total numbers of mutant spots in the normal and serrate wings of the *Drosophila* flies grown in the mediums having three different concentrations of Pea green with *La-vandula stoechas* L. at 100 g/L concentration are higher than that of the flies grown in the medium prepared with distilled water (normal wing in Table 1: $X^2=37.33$, $df=3$, $P<0.001$; serrate wing in Table 2: $X^2=17.92$, $df=3$, $P<0.001$) but it was found to be less than those grown in medium with EMS (normal wing in Table 1: $X^2=322.31$, $df=3$, $P<0.001$; serrate wing in Table 2: $X^2=62.18$, $df=3$, $P<0.001$).

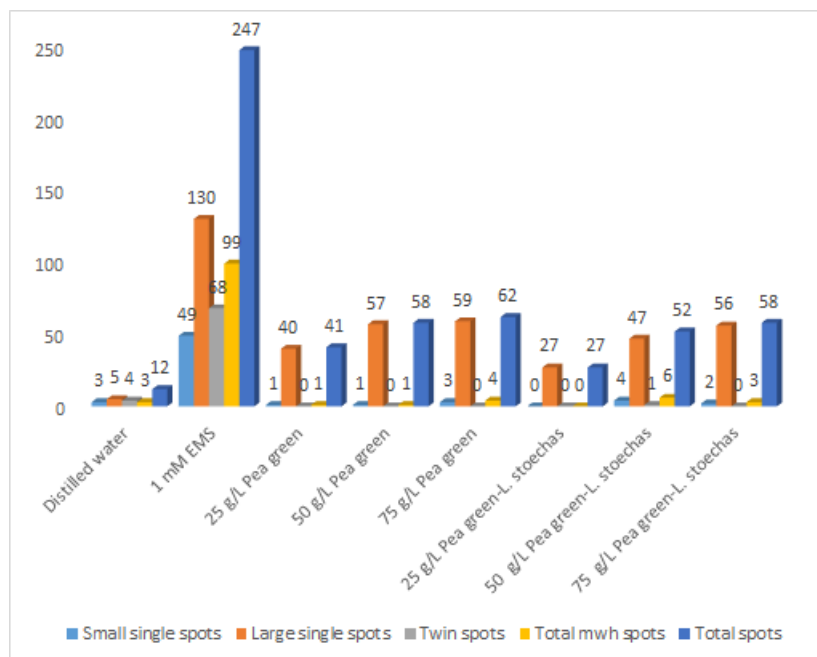


Figure 5. shows classification of mutant clones with the normal wings of the *Drosophila* flies in the experimental group, treatment group with *Lavandula stoechas* L. and the positive and negative control groups.

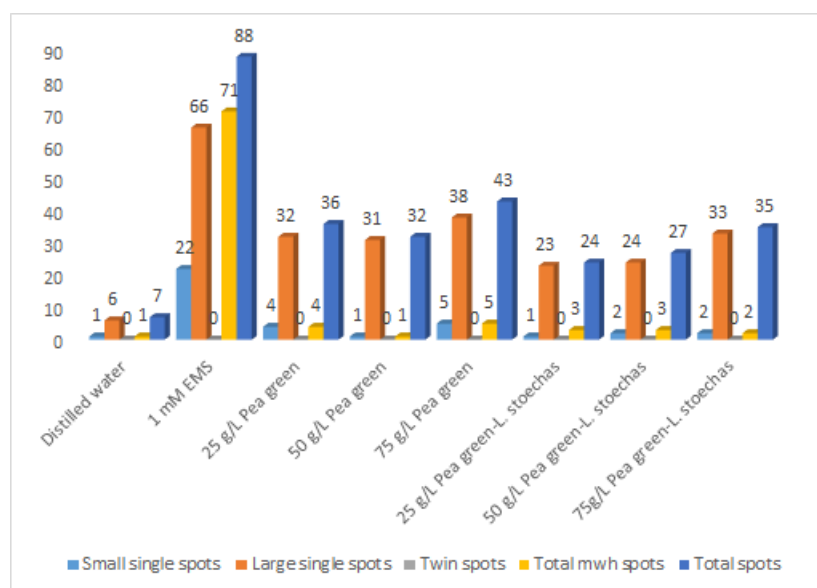


Figure 6. shows classification of mutant clones with the serrate wings of the *Drosophila* flies in the experimental group, treatment group with *Lavandula stoechas* L. and the positive and negative control groups.

Statistical analysis shows that total numbers of mutant spots in the wings of the flies in all experimental and treatment groups increased gradually depending on the concentration of Pea green.

While numerical differences between the total numbers of mutant spots in the normal wings of the flies are evaluated in the experimental group with three different concentrations of Pea green 4R ($X^2=4.63$ $df=2$, $P>0.05$), in treatment group in which three different concentrations of Pea green and *Hypericum perforatum* L. ($X^2=1.62$ $df=2$, $P>0.05$) and in treatment group in which three different concentrations of Pea green and *Silybum marianum* (L.) Gaertn ($X^2=1.98$ $df=2$, $P>0.05$) have not statistically significant, in treatment group in which three different concentrations of Pea green and *Lavandula stoechas* L. ($X^2=11.83$, $df=2$, $P<0.01$) they have statistical significance. In other words, the total numbers of mutant wing spots within themselves of the treatment groups in which Pea green and the plant extracts does not significant differences. Furthermore, statistical differences of the total numbers of mutant spots in the serrate wings of the flies are not significant

de-pend on the concentration in the experimental group ($X^2=1.67$ $df=2$, $P>0.05$), in the treat-ment group in which Pea green and seperately Hypericum perforatum L. ($X^2=2.13$ $df=2$, $P>0.05$), in the treatment group with Pea green and Silybum marianum (L.) Gaertn ($X^2=0.86$ $df=2$, $P>0.05$) and in the treatment group with Pea green and Lavandula stoechas L. ($X^2=2.25$, $df=2$, $P>0.05$).

When average numbers of mutant spots in normal wings ($H=3.77$; $df=3$, $P>0.05$) and serrate wings ($H=3.85$; $df=3$, $P>0.05$) of flies in the experimental group with three different concentrations of Pea green and in the treatment groups in which three different concen-trations of Pea green were together with separately Hypericum perforatum L., Si-lybum.marianum (L.) Gaertn, Lavandula stoechas L. were compared with each other, it was observed that results found were not different.

Finally, when average of total numbers of mutant spots in the normal and serrate wings of the flies in the experimental and treatment groups were compared with each other, it was revealed to be statistically differences ($P<0.001$).

4. Discussion

Although the food additives play a significant role in the food industry, their effects on human health constitute an important issue. As a result of the increase in ready-made food consumption together with urbanization, potential health risks occur with the body's exposure to more additives(AKBULUT 2011).

Many studies on human diseases have shown that *D. melanogaster* Meigen, a down-organized eukaryotic organism, can be used instead of mammals. Because, the bio-logical properties of the imaginal disc cells in *Drosophila* larvae undergoing proliferation and differentiation to form body parts in the adult stage are similar to many can-cer-sensitive mammalian cells, and more than 60% of genes identified in human genetic diseases are common with the genes in *Drosophila* genome. *Drosophila* is frequently in-cluded as a model organism in Wing spot test.

Wing spot test assay principle is based on loss of heterozygosity in recessive marker genes; *mwh* and *flr3* on the third largest chromosome lead formation of imaginal disc cells in *Drosophila* heterozygous larvae into mutant wing spots cell by mutation or re-combination that are the results of genotoxic and anti-genotoxic effects induced by chem-icals (Graf et al. 1984). Moreover, this in vivo test method in *Drosophila* are associated with in vivo genotoxicity tests in mammals (Graf et al. 1984; Graf and Würgler 1996).

Wing spot test plays an important role in the present study for examining that geno-toxic damage caused by the food additive and anti-genotoxic effects of the plant extracts on the model organism, *D. melanogaster* Meigen.

When the present study data are evaluated using Wing spot test, it becomes clear that Pea green has a mutagenic effect in normal and serrate wings of *Drosophila* flies at stud-ied concentrations. Serrate wings of the fruit flies contain only clones resulting from muta-tion, while mutant clones resulting from both mutation and recombination can be seen in normal wings(Kaya, Yanikoglu, and Marcos 1999; Zordan et al. 1994).

The mutant spots induced in *Drosophila* flies with Pea green are a result of the geno-toxic activity of this genotoxic agent depend on its concentration. The food coloring agent has a genotoxic effect in the imaginal disc cells that will transform into the wing spot cells during the embryonic development of *Drosophila* heterozygous larvae and the genotypic changes caused by mutation or recombination in somatic cells also play a role in the for-mation of mutant spots in the wings. The present study result is consistent with the re-searches below, where the toxic effects of Pea green (Tartrazine - E 102 and Brilliant Blue FCF -E 133 combination) are examined. Uysal & Semerdöken(Uysal H. 2011) re-ported increased toxicity in Oregon R of *Drosophila* larvae (72 ± 4 hours) depending on the increase in concentration of Pea green, whereas Semerdoken (Semerdoken 2012) is-sued that Tartrazine increased the mortality rate in Oregon R wild and Vestigial mutant strains of *Drosophila* larvae at 72 ± 4 hours and Uysal et al. (Uysal, Genç, and Ayar 2017) reported that, according to larval mortality and life span, Tartrazine had the highest toxicity in Oregon R wild of *D. melanogaster* larvae at 72 ± 4 hours among other food dyes, Turkoglu et al. (Türkoğlu, Benli, and Şahin 2015) notified that Brilliant Blue FCF caused the biggest decreased in life span of *Drosophila melanogaster* among other food colorants and Kumar et al. (Parameswara Bushan Kumar 2020) revealed that on exposure to Brilliant Blue, larvae and pre-adult stages were prone to developmental toxicity.

Hypericum perforatum L., *Silybum marianum* (L.) Gaertn. and *Lavandula stoechas* L. didn't show reducing or preventive effects on mutations caused by Pea green. When the average of the total numbers of mutant spots on wings of the flies in Pea green treatment and experiment groups were compared with each other, it didn't have any difference.

When the literature is examined, it is seen that there are a few studies about antigen-otoxicity of the plant extracts in *D. melanogaster* Meigen. It was also given below that similar antigenotoxicity studies in various model organisms about the plant extracts evaluated with different test systems.

(Neo A. 2011) reported that *Hypericum perforatum* L. at 2.5%, 12.5% and 50% concentrations slowed cell death in *Drosophila* cells by suppressing the expression of denatured proteins. (Handan Uysal 2007) showed that when *Hypericum perforatum* L. in different concentrations didn't cause elongation of *D. melanogaster* Meigen F1 progeny transformation time to adult, increased numbers of off-spring due to increase in concentrations and had highest numbers of offspring at concentration of 10 ml/100 ml. (Branda et al. 2006) published that *Hypericum perforatum* L. taken with diet was ineffective on leukopenia caused by chemotherapeutic drugs on experimental cancer model induced in rats. (Martarelli et al. 2004) notified that while *Hypericum perforatum* L. prepared in methanol 80% reduced rate of growth and development of prostate adenocarcinoma cells in humans, 70% experimental cancer induced in mice.

(CAN 2014) reported that *Silybum marianum* (L.) taken with diet at 10% concentration reduced DNA damage and apoptosis in rats which liver damage induced by carbon tetrachloride. (Zhou et al. 2006) showed that active substances of *Silybum marianum* (L.) was effective in reducing DNA damage and regulation of cell cycle in rats. (García-Maceira and Mateo 2009) published that cervical and liver cancers in humans are inhibited by *Silybum marianum* (L.).

(Kulcanay 1998) notified that when oil with leaf and flower extracts of *Lavandula stoechas* L. evaluated with Salmonella/microsome test, they were mutagenic. (Ayril 1997) reported that while some of essential oil compounds in above ground parts of *Lavandula stoechas* L. had strong cytotoxic and anti-bacterial effects in different cancer cells. (Magro et al. 2006) showed that when one of the antigenotoxic effects of *Lavandula stoechas* L. tested for fungistatic (delaying or inhibiting fungal growth) activity in various mushrooms, the plant extract was ineffective.

Results of the researches above mentioned and these study data are not parallel to each other. Accordingly, the study data show that these plant extracts were ineffective at 100 g/L concentration studied as a single dose in protecting *Drosophila* flies against the genotoxicity of the synthetic food coloring. More basic and clinical studies are need to be determined for effectiveness of the plant extracts. Furthermore, the plant extracts can be used in different concentrations and for longer periods of time. Determining the efficacy and safety of the plant extracts is closely related to the health of today and in the future generations.

It is thought that this study can contribute to scientific researches and similar experiments will do later. More scientific studies are required to explain molecular mechanisms and distributions of the genotoxicity of the food coloring used in the study and the anti-genotoxicity of the plant extracts by using in vivo and in vitro test systems in/on different model organisms.

Nutrition is one of the most important factors ensuring the continuity of life. Nowa-days, it is seen that people's eating habits significantly differ as a result of the changes in lifestyle and economic development. (AKBULUT 2011). With controlling the synthetic food coloring and the plants extracts consumption public health should be protected.

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