

# Biochemical Results of Bioinfluence of Risimanski on Glucose Solution

Ignat Ignatov<sup>1\*</sup> Oleg Mosin<sup>2</sup>

1. DSc, professor, Scientific Research Center of Medical Biophysics (SRCMB),  
32 N. Kopernik St., Sofia 1111, Bulgaria

2. PhD, Biotechnology Department, Moscow State University of Applied Biotechnology,  
33 Talalikhina St., Moscow 109316, Russian Federation

## Abstract

This paper presents the results of evaluation of possible biophysical methods and approaches for registering of various non-ionizing radiation (NIR) wave types of the human body in the electromagnetic and optical range. Many types of NIR (electromagnetic waves, infrared radiation, thermo radiation, bioluminescence) emitted from the human body were reviewed. In particular the results on of spontaneous biophoton emission and delayed luminescence from the human body are submitted along with infrared thermography (IRT) results. It was shown that 1 cm<sup>2</sup> of skin generally emits ~85 photons for 1s. The intensity of biophoton emission ranges from 10<sup>-19</sup> to 10<sup>-16</sup> W/cm<sup>2</sup> (approx. ~1–1000 photons·cm<sup>-2</sup>·s<sup>-1</sup>). The specific photon emission from part of the human thumb was detected as a spectrum of various colors with the method of Colour coronal spectral analysis (Ignatov, 2007) on a device with an electrode made of polyethylene terephthalate (PET hostafan) with applied electric voltage 15 kV, electric impulse duration 10 μs, and electric current frequency 15 kHz. It was established that photons corresponding to a red color emission of visible electromagnetic spectrum have energy at 1.82 eV. The orange color of visible electromagnetic spectrum has energy at 2.05, yellow – 2.14, blue-green (cyan) – 2.43, blue – 2.64, and violet – 3.03 eV. The reliable result measurement norm was at E ≥ 2.53 eV, while the spectral range of the emission was within 380–495 nm and 570–750 nm±5 nm. Also were estimated some important physical characteristics (energy of hydrogen bonds, wetting angle, surface tension) of water by the methods of non-equilibrium energy (NES) and differential non-equilibrium energy (DNES) spectrum of water, that helps understand in general how electromagnetic radiation interacts with water and establish the structural characteristics of water.

**Keywords:** electromagnetic waves, 1% glucose solution, water spectrum, NES, DNES

## 1. Introduction

All living organisms have a cellular therefore, a molecular organized structure. The living processes inside of them run on a cellular and a molecular level. Bioelectrical activity is one of the very important physical parameters of living organisms (Ignatov et al., 1998). Bioelectric potentials generated by various cells are widely used in medical diagnostics (Rubik, 2002) and are recorded as electrocardiogram, electromyogram, electroencephalogram, etc. It was proved that the human body and tissues emanate weak electromagnetic waves, the electric voltage of which is denoted as resting potential, action potential, omega-potential etc. (Dobrin *et al.*, 1979; Adey, 1981). Between the outer surface of the cell membrane and the inner contents of the cell there is always the electric potential difference which is created because of different concentrations of K<sup>+</sup>, Na<sup>+</sup> and Cl<sup>-</sup> inside and outside of the cell and their different permeability through the cell membrane (Kiang *et al.*, 2005). Their value in the human body varies ~50–80 mV and is defined by the galvanic contact of a voltmeter input with an object that indicates on the galvanic type of their source (Cleary, 1993). When being excited a living cell changes the membrane electric potential due to changes in membrane permeability and active ion movement through the membrane. In cells of excitable tissues (muscle, nervous), these processes can occur within a very short time intervals (milliseconds) and are called “current action” potential. Its magnitude makes up ~120 mV. Electromagnetic fields refer to non-ionizing radiation (NIR), i.g. the radiative energy that, instead of producing charged ions when passing through matter, has sufficient energy only for excitation. Nevertheless it is known to cause biological effects (Kwan-Hoong, 2003). The NIR spectrum is divided into two main regions, optical radiations and electromagnetic fields. The optical spectrum can be further sub-divided into ultraviolet, visible, and infra-red. The electromagnetic fields are further divided into radiofrequency (microwave, very high frequency and low frequency radio wave). NIR encompass the long wavelength (> 100 nm) and low photon energy (<12.4 eV) portion of the electromagnetic spectrum, from 1 Hz to 3·10<sup>15</sup> Hz. As a result of research carried out in the 1990-s and subsequent years, it was established the property of animal and plant tissues to generate relatively strong transient NIR electric fields due to mechanical stresses and temperature changes in biological structure (Anderson, 1993). These electric fields are mainly due to the piezoelectric and pyroelectric voltage electric polarization of natural biological structures. Owing to cell metabolism, electric dipoles (polar and ionized molecules) involved in polarization of biostructures are continuously destroyed and restored, i.e. this

is a non-equilibrium polarization (Barnes & Greenebaum, 2006). Such type of non-equilibrium electric polarization is known as a main characteristic of electrets (Gubkin, 1978). Electrets include dielectric insulators and semiconductors, which under certain conditions, i.g. under the influence of a strong electrostatic field or ionizing radiation, light and other factors acquire property to generate an external electric field, existing for a long time (days, years) and slowly diminishes because the destruction of their substance by polarization (Sessler & Gerhard-Multhaupt, 1998). Along with the electromagnetic field electrets generate specific electric currents produced by heating – thermally stimulated current (TSC) (Gross, 1964). Electrets belong to the non-galvanic type of electrical sources, which tend to a strong electric field (up to  $10^6$  V/m) and the infinitesimal electric current ( $\sim 10^{-14}$  A/mm<sup>2</sup>). By analogy with the physical fields the electric field emitted from the human body on its physical characteristics resembles the electric field generated by electrets. The electrets play an important role in functioning of many biological structures as they themselves possess electret properties. The bioelectret field registered on the surface of the human body basically are generated by the basal cells of the epidermis (Marino, 1988). Dermis cells adjacent to the bottom layer of basal cells are surrounded by a conductive interstitial fluid, which electric voltage while grounding of the human body is close to zero (so called ground potential). This interstitial fluid screens off electromagnetic fields of underlying tissues. With the average thickness of the epidermis ( $\sim 0.1$  mm) and the maximum value of electric voltage ( $\sim 30.0$  V), the electric field strength can reach significant values at  $\sim 300000$  V/m (Seto et al., 1992). The strength of the electric field is quiet sufficient for its influence on the biological processes in cells and surrounding tissues, including the synthesis of proteins and nucleic acids (Liboff *et al.*, 1984; Frey, 1993; Shimizu *et al.*, 1995). This electric field along with the field of transmembrane assymetry of ions concentrated at inside and outside of the membrane ( $\sim 10^5$  V/cm<sup>2</sup>) can participate in the cooperative effects in cell membrane structures (Holzel & Lamprecht, 1994; Miller, 1986). Thus, owing to the bioelectret condition of certain subcellular structures in the cell and its surroundings is generated slowly oscillating electric field that is strong enough to influence the biological processes. This field and the electric field due to the piezoelectric voltage and intramembrane electric field formes the total electromagnetic field of the cell and its supracellular structures. It is known that the human skin emanates electromagnetic waves in close ultraviolet range, optic range and also in close infrared range. Infrared thermal bioradiation is found in the middle infrared range at wavelengths from 8 to 14  $\mu\text{m}$ . At wavelength of 9.7  $\mu\text{m}$  infrared bioradiation has its maximum value at  $t = 36.6$  °C. At this temperature the skin emission is closest to the emission of absolute black body (ABB) being at the same temperature. Infrared emission penetrates the skin surface at a depth of  $\sim 0.1$  mm, and is reflected in accordance with the physical laws of reflection of the visible part of the electromagnetic spectrum. Evidently, radiation energy influences tissues while being absorbed by them. Yu.V. Gulyaev and E.E. Godik (Gulyaev & Godik, 1984) determined that the threshold of skin sensitivity for infrared radiation compiled  $\sim 10^{-14}$  W/cm<sup>2</sup>. When thermal influence is applied to the point of threshold skin sensitivity, there is developed a physiological reaction toward the thermal current. The intensity of the radiated thermal current generated by skin makes up  $\sim 2.6 \cdot 10^{-2}$  W/cm<sup>2</sup>. The second component of electromagnetic waves is bioluminescence (Young & Roper, 1976; Chang *et al.*, 1998). It is supposed that biophotons, or ultraweak photon emissions of biological objects, are weak electromagnetic waves in the optical range of the spectrum (Cohen & Popp, 1997). The typical observed emission of biological tissues in the visible and ultraviolet frequencies ranges from  $10^{-19}$  to  $10^{-16}$  W/cm<sup>2</sup> ( $\sim 1$ – $1000$  photons $\cdot\text{cm}^{-2}\cdot\text{sec}^{-1}$ ) (Edwards *et al.*, 1989; Choi *et al.*, 2002). This light intensity is much weaker than that one to be seen in the perceptually visible and well-studied spectrum of normal bioluminescence detectable above the background of thermal radiation emitted by tissues at their normal temperature (Niggli, 1993). Bioelectric emission from parts of the human body as thumbs can be easily detected with the method of Color coronal spectral analysis under applying gas electrical discharge of high voltage and frequency developed by I. Ignatov (Ignatov, 2005). This method has big scientific and practical prospects in biophysics and medical diagnostics (Chiang et al., 2005). Its advantages include safety, sterility, clarity and interpretability of the data obtained, ease of storage and subsequent computer data processing, the ability to monitor the development of processes in time, comparing the structural, functional and temporal processes etc. The purpose of this research was studying of possible biophysical methods and approaches for registering various NIR wave's types emitted from the human body (electromagnetic waves, infrared radiation, thermo radiation) and methods of their visualization by different technique including magnetography, infrared thermography, chemiluminescence and coronal gas discharge spectral analysis.

## 2. Materials and methods

### 2.1. Registration of electromagnetic fields

The registration of electromagnetic fields was used with super conductive detectors based on Joseffson junctions – device made by sandwiching a thin layer of insulating nonsuperconducting material between two layers of superconducting cooper pairs (S-I-S). This allows the registering of magnetic fields  $10^{10}$  times weaker than the Earth's magnetic field. The study of electric field nearly the human body was done using a standard Faraday

cage formed by conducting material (aluminium foil) blocks external static and non-static electric fields by channeling electricity through the conducting material, providing constant voltage on all sides of the enclosure.

## **2.2. NES and DNES experiments on interaction of electromagnetic field with water**

The research was made with the method of non-equilibrium spectrum (NES) and differential non-equilibrium spectrum (DNES). The device measures the angle of evaporation of water drops from  $72^\circ$  to  $0^\circ$ . As the main estimation criterion was used the average energy ( $\Delta E_{H...O}$ ) of hydrogen O...H-bonds between  $H_2O$  molecules in water's samples. The spectrum of water was measured in the range of energy of hydrogen bonds 0.08–0.1387 eV or 8.9–13.8  $\mu\text{m}$  with using a specially designed computer program.

## **3. Results and discussions**

### **3.1. Electric fields**

The electric field surrounding the human body with frequency  $\nu = 1 \cdot 10^3$  Hz is created by electrochemical processes in the organism and is modulated by the rhythm of internal organs (Ignatov et al., 1998). The spatial distribution of the electric field around the body reflects the teamwork of the different organs and systems in the organism. There are also electric fields, which are generated by accumulation of triboelectric (caused by friction) charge on the epidermis, which depends on epidermal electric resistance and varies from  $10^9$  to  $10^{11}$   $\Omega/\text{cm}^2$ . Radiothermal emission is being detected in the centimetre and decimetre range of the spectrum. This type of emission is connected with the temperature and the biorhythms of the internal organs, and is being absorbed by surface layer of skin at depth from 5 cm to 10 cm (Gulyaev & Godik, 1984). Long persistent electric field nearby the human body can be detected with using an electrometer voltmeter after neutralizing electric charges on the skin caused by triboelectric charges. The electric strength of this field is undergoing slow oscillations, and most patients exert its value within the range of 100–1000 V/m at a distance of 5–10 cm from the body. People in a state of clinical death usually have the electric field strength's value reduced to 10–20 V/m after 2–3 hours of cardiac arrest. Intensity vector of the detected electric field is found to be normal to the surface of the skin, and the electric voltage is inversely proportional to the distance. On the skin surface the electric voltage of the field (the difference of its electric potential with respect to ground potential) reaches essential values of ~10000 mV or more, i.e., is about 1000 times greater than the source electric voltage of the electric unit above the bioelectric potentials. This allows us to characterize the electric field detected nearby the human body as relatively strong electric field emitted from living tissues. Its electric voltage was measured by electrometric methods, indicating on non-galvanic type of its source.

If the physical basis of the generation of a relatively strong electric field in the human tissue is non-equilibrium electric polarization of the substance due to metabolic processes, the electric field strength should depend on these processes. As noted above, this dependence is actually observed: inhibition of tissue metabolism due to hypoxia during cardiac arrest was accompanied by drop in the electric field strength. This relationship is confirmed in experiments on animals (Gerald et al., 2008). For example, in rats inhibition of metabolism of the tissue due to cardiac arrest (death of the animal) or by general anesthesia is accompanied by a significant drop in the electric field strength (Bars & Andre, 1976).

Electric fields depend on the magnitude of the electric voltage and the distance from the source (Kwan-Hoong, 2003). Generally, the electric voltages are stable and remain the same; however electric fields are easily perturbed and distorted by many surrounding objects. Relatively strong electric field investigated in humans and animals is being formed evidently by skin's biostructures, since the electric fields of the underlying tissues are largely shielded by conductive interstitial fluid (Goodman *et al.*, 1995; Gulyaev & Godik, 1990). The greatest contribution to the detected electric field makes the basal cells of the epidermis – the top layer of the skin. Electric polarization vector of these cells is normal to the surface of the skin, i.e., coincides with the electric voltage's vector field, and yet it is inherent in the metabolism intensity, conditioning the generation of the electric field.

### **3.2. Magnetic fields**

Magnetic field of a living organism can be caused by three following reasons. First of all, it is ion channels arising from electrical activity of cell membranes (primarily muscle and nervous cells). Another source of magnetic fields are tiny ferromagnetic particles, trapped or specially introduced into the human body. These two sources create their own magnetic fields. In addition, at imposition of external magnetic field there appears inhomogeneity of the magnetic susceptibility of different organs and tissues distorting the external magnetic field (Wikswow & Barach, 1980). The magnetic field in the last two cases is not accompanied by the appearance of the electric field, so the study of the behavior of magnetic particles in the human body and the magnetic properties of various organs are applicable only with using of magnetometric methods. Biocurrents on the contrary except for the magnetic fields create the distribution of electric potentials on a body's surface. Registration of these electric potentials has long been used in research and clinical diagnostics – in electrocardiography,

electroencephalography etc (Cohen, 1968). It would seem that their magnetic counterparts, i.e. magnetocardiography and magnetoencephalography recording the signals from the same electrical processes in the body, will give almost the same information about the studied organs. However, as follows from the theory of electromagnetism, the structure of the electric current source in the electric conductive medium (the body) and the heterogeneity of the medium have significantly different impact on the distribution of magnetic and electric fields: some types of bioelectric activity manifest themselves primarily in the electric field, giving a weak magnetic signal, while the others – on the contrary create rather strong magnetic signal (Zhadin, 2001; Anosov & Trukhan, 2003). Therefore, there are many biophysical processes which observation is preferable by using of magnetographic methods. Magnetography does not require the direct contact with the investigated object, i.e., it allows to carry out measurements over a bandage or other obstructions. It is not only practically useful for diagnostics, but is fundamental advantage over electrical methods towards data recording, as the attachment of the electrodes on the skin can be a source of slowly varying contact electric potentials. There are no such spurious noises while using magnetographic methods, therefore, magnetography allows, in particular, reliably explore slowly occurring processes (with the characteristic time of tens of minutes). Magnetic fields rapidly diminish with distance from the source of the activity, as they are caused by relatively strong currents running in the body, while the surface potentials are determined mainly by the weaker and “smeared” electric currents in the skin. Therefore, magnetography is more convenient for accurate determination (localization) of bioelectric activity parts on the human body. And finally, the magnetic field vector is characterized as not only by the absolute value but also by the direction, which also may provide additional useful information. However, it should not be assumed that the electricity and magnetographic methods compete with each other. On the contrary, it is their combination that gives the most complete information about the processes being investigated. But for each of the individual methods, there are practical areas wherein the use of any one of them is preferable. Water is the main substance of all living organisms and the magnetic field exerts a certain influence on water. This influence is a complex multivariate influence, which the magnetic field exerts on dissolved in water metal cations ( $\text{Fe}^{2+}$ ,  $\text{Fe}^{3+}$ ) and the structure of the hydrates and water associates (Mosin, 2011). Experimentally was proved that the magnetic field acts much weaker on still unmoved water, because water has a conductivity; as water moves in the electromagnetic field it is generated a small electric current (Mosin, 2012). The research performed with superconductive detectors based on Josephson junctions shows that magnetic fields around the human body are in the range from 1 to 100 Hz. The magnetic activity of the brain for example makes up  $\sim 30 \cdot 10^{15}$  T/Hz<sup>1/2</sup>. The magnetometric system has a sensitivity of  $10 \cdot 10^{15}$  T/Hz<sup>1/2</sup> in the range of 1 to 100 Hz (Gulyaev & Godik, 1990).

### 3.3. Chemiluminescence

Chemiluminescence denotes luminescence accompanying chemical reactions, detected in the near infrared, the optical and the near UV-range of the electromagnetic spectrum. When chemiluminescence takes place in living organisms, the phenomenon is called bioluminescence. The term is generally used for higher luminance ATP-dependent luciferin/luciferase systems observed in *Lampyridae* lightning bugs (Rauhut, 1985). Although there bacterial, latia and dinoflagellate luciferin, and coelenterazine found in some bacteria, freshwater snails, dinoflagellates, radiolarians, shrimp, squid and deep-sea fish species (Hastings, 1983). Chemiluminescence is observed in reactions accompanied by allocation of large amounts of energy, such as the reaction of combining of two radicals, or in reactions involving peroxides, e.g. peroxide oxidation of lipids (Halliwell & Gutteridge, 1989). Peculiar (“ultra-weak”) glow of cells and tissues of animals and mammals is caused by free radical reactions: lipid radicals and oxygen, and nitrogen oxides – compounds that play an essential role in cell metabolism, and under certain conditions in the development of a number of pathological conditions. It is suggested that the major radicals responsible for the light emission are excited triplet carbonyl and excited singlet oxygen, and that these radicals arise through the decomposition of hydroperoxides formed in the process of lipid oxidation (Zlatkevich & Kamal-Eldin, 2005; Porter & Wujek, 1988). The process of lipid peroxidation (LPO) is an important cause of the accumulation of cellular defects caused by radicals. LPO main substrate is polyunsaturated fatty acids in composition of cell membranes and lipoproteins. Their attack by oxygen radicals leads to the formation of hydrophobic radicals, interacting with each other (Vladimirov, 1996). Many different mechanisms have been suggested for the oxidative lipid fragmentation that produce biologically active aldehydes as 4-hydroxynon-2-enal (HNE), oxononanoyl phosphatidylcholine (ON-PC) from linoleic acid (LA) esters, or HNE and oxovaleroyl phosphatidylcholine (OV-PC) (Esterbauer *et al.*, 1990). Processes of life are almost always accompanied by a very weak radiation, which is sometimes called ultra-low illumination or radiation of cells and tissues (Boveris *et al.*, 1980). Some organisms possess the ability to emit bright light at photon fluxes below about  $10^4$  photons·cm<sup>-2</sup>·s<sup>-1</sup>, visible to the naked eye, this phenomenon is denoted “bioluminescence”. In biochemical systems, i.e. in tissue’s homogenates, cell suspensions or cell organelles, mixtures of enzymes and substrates, chemiluminescence in most cases has an extremely low intensity, and requires particularly sensitive equipment for its detection and measurement (Popp *et al.*, 2002). Some substances – enhancers, have the ability

to essentially enhance the chemiluminescence, sometimes many thousands of times (activated, or enhanced chemiluminescence). In addition, weak luminescence is accompanied by the formation of free radicals under the action of a number of physical factors on the object: at ionizing radiation is observed radiochemiluminescence, at ultraviolet or visible light illumination – photochemiluminescence, at passing an electric current – electroluminescence, with ultrasound – sonoluminescence, under the influence of friction forces – triboluminescence. Chemiluminescence differs from fluorescence in that the electronic excited state is derived from the product of a chemical reaction rather than the more typical way of creating electronic excited states, namely adsorption. In photomechanical reactions, in which light is used to drive an endothermic chemical reaction, light is generated from a chemically exothermic reaction. At present time it is known quite a lot of chemical reactions involving the formation of luminescence glow. In most cases they are generally quite a complex processes having many intermediate stages, but the basic processes leading to luminescence glowing in general are similar. They include the separation and transfer of charged particles (electrons and free radicals), the electron transfer (redox reactions) at one of the higher energy levels with the formation of the reaction product in an electron-excited state and further releasing of a photon in the transition of the molecule to the low excited electronic ground state with a lower energy level (luminescence). Theoretically, in this process on each molecule of the reactant should be allocated one photon. Chemiluminescence accompanies many chemical reactions (ozonation and fluorination reactions, the oxidation of phosphorus and complex organic substances, lipids) and has an impulse mode; the signals of this process usually are very weak. Thus, the human skin dissociates few photons per 1 sec. with emission power level  $\sim 10 \text{ mW/cm}^2$  (Gulyaev, Godik, 1991).

- Luminescence of cells and tissues are accompanied by three types of reactions:
- Reactions with active oxygen;
- Chain reactions of lipid peroxidation; Reactions involving nitric oxide (NO).

The more lipid radicals contains the system, e.g. the more energetically occurs the chain reaction of lipid oxidation, the higher the intensity of chemiluminescence accompanying the reaction of radicals. Substances reacting with free radicals and thereby inhibiting the chain lipid oxidation (so-called antioxidants) simultaneously inhibit chemiluminescence. That inhibition of chemiluminescence by cells and tissues by such antioxidants as tocopherol (vitamin E), indicating that chemiluminescence is stipulated by lipid oxidation chain reactions. On the other hand, studying the impact of various natural and synthetic compounds on the time (kinetics) of chemiluminescence, it can be judged on the ability of these substances to protect our body from the harmful effects of free radicals and thereby select candidates to certain medications. Chemiluminescent methods are used for recording of ultra-weak light waves accompanying from the chemical and biochemical reactions involving the formation of free radicals. They do not require special laboratory conditions and special material preparation for analysis and characterised by high sensitivity, reliability, meet the requirements for rapid methods of express research. Chemiluminescent methods are widely used in biomedical diagnostics for studying of the molecular basis of physiological processes in biological systems and general mechanisms of development of pathological conditions.

### **3.4. NES and DNES analysis of water and 1% solution of glucose**

Water seems to be a good model system for studying the interaction with electromagnetic field and structural research. The recent data indicated that water is a complex associated non-equilibrium liquid consisting of associative groups (clusters) containing from 3 to 50 individual  $\text{H}_2\text{O}$  molecules (Keutsch & Saykally, 2011). These associates can be described as unstable groups (dimers, trimers, tetramers, pentamers, hexamers etc.) in which individual  $\text{H}_2\text{O}$  molecules are linked by van der Waals forces, dipole-dipole and other charge-transfer interactions, including hydrogen bonding (Ignatov & Mosin, 2013c). At room temperature, the degree of association of  $\text{H}_2\text{O}$  molecules may vary from 2 to 21. The measurements were performed with using NES and DNES methods. It was established experimentally that the process of evaporation of water drops, the wetting angle  $\theta$  decreases discreetly to 0, and the diameter of water drop basis is only slightly altered, that is a new physical effect (Antonov & Yuskesseliya, 1983). Based on this effect, by means of measurement of the wetting angle within equal intervals of time is determined the function of distribution of  $\text{H}_2\text{O}$  molecules according to the value of  $f(\theta)$ . The distribution function is denoted as the energy spectrum of the water state. A theoretical research established the dependence between the surface tension of water and the energy of hydrogen bonds among individual  $\text{H}_2\text{O}$ -molecules (Antonov, 1995). The hydrogen bonding results from interaction between electron-deficient H-atom of one  $\text{H}_2\text{O}$  molecule (hydrogen donor) and unshared electron pair of an electronegative O-atom (hydrogen acceptor) on the neighboring  $\text{H}_2\text{O}$  molecule; the structure of hydrogen bonding may be defined as  $\text{O} \cdots \text{H}^+ - \text{O}^-$ .

For calculation of the function  $f(E)$  represented the energy spectrum of water, the experimental dependence between the wetting angle ( $\theta$ ) and the energy of hydrogen bonds ( $E$ ) is established:

$$f(E) = b \times f(\theta) / 1 - (1 + b \times E)^{1/2}, \quad (4)$$

$$\text{where } b = 14.33 \text{ eV}^{-1} \quad (5)$$

The relation between the wetting angle ( $\theta$ ) and the energy ( $E$ ) of the hydrogen bonds between  $H_2O$  molecules is calculated by the formula:

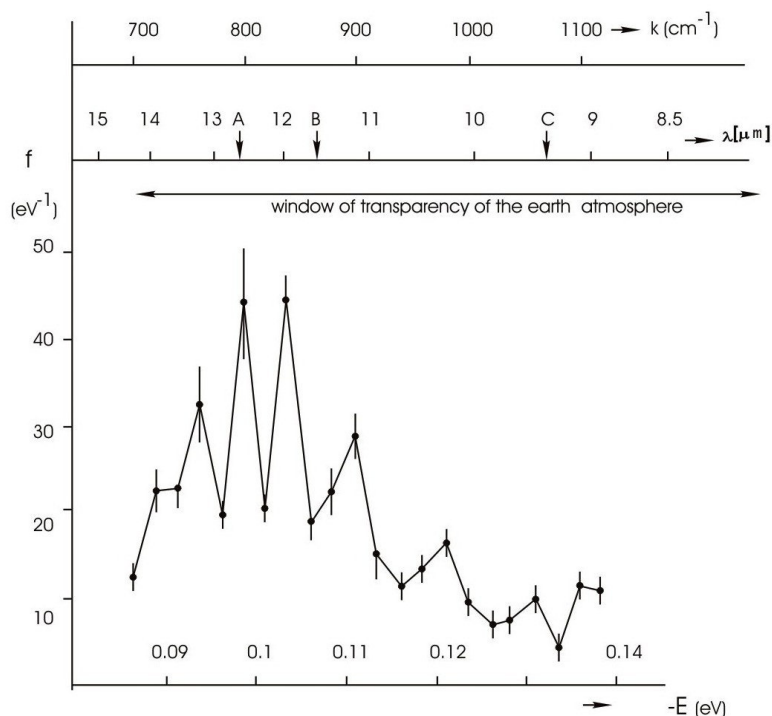
$$\theta = \arcsin(-1 - 14.33E) \quad (6)$$

The energy spectrum of water is characterized by a non-equilibrium process of water droplets evaporation, therefore, the term non-equilibrium spectrum (NES) of water is used. The energy of hydrogen bonds measured by NES is determined as  $\bar{E} = -0,1067 \pm 0,0011$  eV.

The difference  $\Delta f(E) = f(\text{samples of water}) - f(\text{control sample of water})$  – is called the “differential non-equilibrium energy spectrum of water” (DNES).

Thus, DNES spectrum is an indicator of structural changes of water as a result of various external factors. The cumulative effect of these factors is not the same for the control sample of water and the water sample being under the influence of this factor.

Figure 1 shows NES-spectrum of deionized water that was used as a model system for studying the interaction of electromagnetic field with water. On the X-axis are given three scales. The energies of hydrogen bonds among  $H_2O$  molecules are calculated in eV. On the Y-axis is shown the energy distribution function  $f(E)$  of  $H_2O$  molecules measured in  $eV^{-1}$ . It was shown that the window of transparency of the earth atmosphere for the electromagnetic radiation in the middle IR-range almost covers NES-spectrum of water. Arrows A and B designate the energy of hydrogen bonds among  $H_2O$  molecules. Arrow C designates the energy at which the human body behaves itself as absolute black body (ABB) at optimum temperature  $36.6^\circ C$  and adsorbs the thermal radiation. A horizontal arrow designates the window of transparency of the earth atmosphere for the electromagnetic radiation in the middle IR-range.

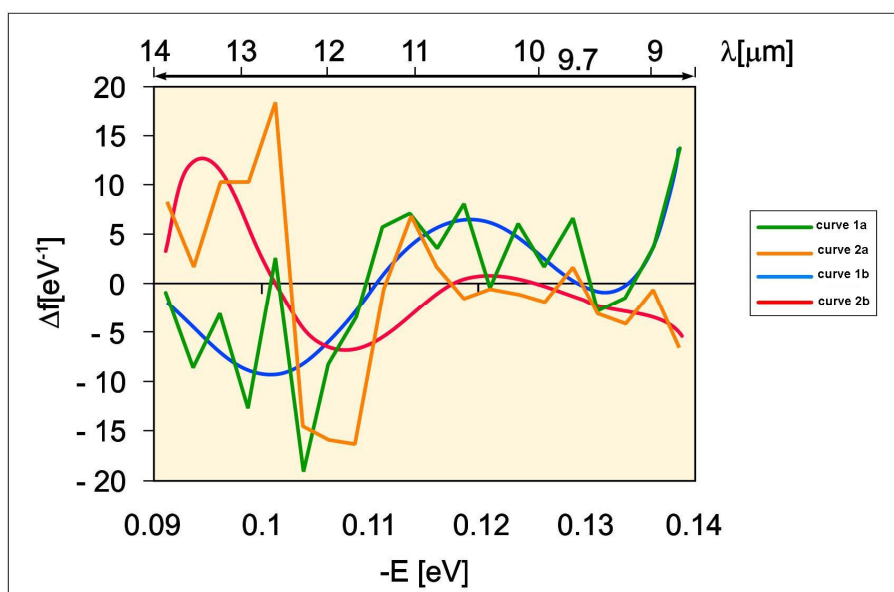


**Fig. 1.** Non-equilibrium energy spectrum (NES) of water as a result of measurement for 1 year:  $\lambda$  – wavelength,  $k$  – wave number.

Another important physical parameter was calculated with using NES and DNES methods – the average energy ( $\Delta E_{H...O}$ ) of H...O-bonds between  $H_2O$  compiled  $-0.1067 \pm 0.0011$  eV. The most remarkable peculiarity of H...O-bond consists in its relatively low strength; it is 5–10 times weaker than chemical covalent bond. In respect of energy hydrogen bond has an intermediate position between covalent bonds and intermolecular van der Waals forces, based on dipole-dipole interactions, holding the neutral molecules together in gasses or liquefied or solidified gasses. Hydrogen bonding produces interatomic distances shorter than the sum of van der Waals radii, and usually involves a limited number of interaction partners. These characteristics become more substantial when acceptors bind H atoms from more electronegative donors. Hydrogen bonds hold  $H_2O$  molecules on 15 % closer than if water was a simple liquid with van der Waals interactions. The hydrogen bond energy compiles 5–10 kcal/mole, while the energy of covalent O–H-bonds in  $H_2O$  molecule – 109 kcal/mole. With fluctuations of water temperature the average energy of hydrogen H...O-bonds in  $H_2O$  molecule associates changes. That is why hydrogen bonds in liquid state are relatively weak and unstable: it is thought that they can easily form and disappear as the result of temperature fluctuations. The next conclusion that can be drawn from

our research is that there is the distribution of energies among individual H<sub>2</sub>O molecules.

Further we performed two types of temperature-dependent experiments on heat exchange from the surface of the human body by DNES-method. In first experiment we studied heat exchange when the temperature of the human body was higher than the temperature of the surrounding environment (curve 1a and 1b on Fig. 5). In second experiment there was heat exchange when the temperature of the human body was lower than that of the surrounding environment (curve 2a and 2b on Fig. 5). In both experiments it was detected a local maximum at 9.7  $\mu\text{m}$  on curve 1 and curve 2 (Fig. 2). This local maximum corresponds to the maximal level of heat emission from the surface of the human body and lays within the “transparency window” of Earth atmosphere to electromagnetic radiation in the mid IR-range of the electromagnetic spectrum. In this range, the electromagnetic radiation emitted by the earth in the surrounding space is being absorbed by the Earth atmosphere. There is a statistical difference between the results of heat emission from the surface of the human body to the surrounding environment and back to the human body according to the *t*-criterion of Student at  $p < 0.01$ . The local maximum on curve 1a is detected at  $7.3 \text{ eV}^{-1}$ , while the local maximum on curve 2a – at  $2.4 \text{ eV}^{-1}$  (Fig. 2).



**Fig. 2.** Differential non-equilibrium energy spectrum (DNES) reflecting the heat exchange of the human body with surrounding environment.

There are the following measurements of the results of bioinfluence of Risimanski on water spectrum and 1% solution with glucose. The biophysical result of Dimitar Risimanski from him to environment with measurement with deionized water is (-7.2 meV). The result of Dimitar Risimanski from environment to him is (7.1 meV). The difference is definite as effective energy is  $(-7.2) - (7.1) = (-14.3) \text{ meV}$ . The biochemical result of Dimitar Risimanski from him to 1% glucose solution is (-12.3 meV) and when he influences on bottle in metal screen is (-10.7 meV).

#### 4. Conclusions

In frames of this research many types of NIR radiation (electromagnetic waves, infrared radiation, thermo radiation, bioluminescence) emitted from the human body were studied and carefully scrutinized. The approaches and methods for detecting various types of radiation employed in this research as magnetography, chemiluminescence and coronal gas discharge spectral analysis can find further application in many branches of applied science and medical diagnostics, while other methods as NES and DNES may be applied for studying the interaction of electromagnetic fields with water and structural studies. In the report are presented the phenomenal biophysical results of influence of Dimitar Risimanski (Bulgaria). The results are base for additional biochemical, biological and medical research projects. The research of Risimanski's bioinfluence was performed with two model systems. First is biophysical analysis on water spectrum. Second is biochemical analysis with 1% solution of glucose.

#### References

- Adey, W.R. (1981) Tissue interaction with non-ionizing electromagnetic fields. *Physiol. Rev.*, 61: 435–514.
- Anderson, L.E. (1993) Biological effect of extremely low frequency electromagnetic fields: *in vivo* studies. *Am. Ind. Hig. Assoc. J.*, 54: 186–196.

- Anosov V.N. & Trukhan E.M. (2003) A new approach to the problem of weak magnetic fields: An effect on living objects. *Doklady Biochemistry and Biophysics*, 392(1-6): 274-278.
- Antonov, A. & Yuskesseliya, L. (1985) Selective high frequency discharge (Kirlian effect). *Acta Hydrophysica*, Berlin, 5: 29.
- Antonov, A. (1995) Research of the non-equilibrium processes in the area in allocated systems. Dissertation thesis "Doctor of physical sciences", Blagoevgrad, Sofia.
- Barnes, F.S. & Greenebaum, B. (eds.) (2006) CRC Handbook on biological effects of electromagnetic fields. 3d Edition, Boca Raton: CRC Press, November 2006, 2, 960 .
- Bars, Le. & Andre, G. (1976) Biological effects of electric fields on rats and rabbits. *Red. Gen. Elect.* (special issue), July 1976: 91-97.
- Belousov, L.V., Opitz, J.M. & Gilbert, SF (1997) Life of Alexander G. Gurwitsch and his relevant contribution to the theory of morphogenetic fields. *The International journal of developmental biology*, 41(6): 7-771.
- Belousov, L., Popp, F.A., Voeikov, V., van Wijk, R. (eds) (2000) Biophotonics and Coherent Systems. Moscow, Moscow University Press, 133.
- Boveris, A., Cadenas, E., Reiter, R., Filipkowski, M., Nakase, Y. & Chance, B. (1980) Organ chemiluminescence: Noninvasive assay for oxidative radical reactions. *Proc. Natl. Acad. Sci. USA*, 77: 347-351.
- Chang, J.J., Fisch, J. & Popp, F.A. (eds) (1998) Biophotons. Dordrecht, Kluwer Academic Publishers, 417 p.
- Chiang, L.H, Wah Khong, P. & Ghista, D. (2005) Bioenergy based medical diagnostic application based on gas discharge visualization. *Conf Proc IEEE Eng. Med. Biol. Soc.*, 2:1533.
- Choi, C., Woo, W.M., Lee, M.B. *et al.* (2002) Biophoton emission from the hands. *J. Korean Physical. Soc.*, 41:275-278.
- Cleary, S.F. (1993) A review of *in vitro* studies: low-frequency electromagnetic fields. *J. Am. Ind. Hyg. Assoc*, 54(4): 178-185.
- Cohen, D. (1968) Magnetoencephalography: evidence of magnetic fields produced bi alpha-rhythm currents. *Science*, 161(3843): 784-786.
- Cohen, S. & Popp, F.A. (1997) Biophoton emission of the human body. *Journal of Photochemistry and Photobiology B: Biology*, 40(2): 187-189.
- Devaraj, B., Usa, M. & Inaba, H. (1997) Biophotons: ultraweak light emission from living systems. *Curr. Opin. Solid State Mater Sci.*, 2:188.
- Dobrin, R., Kirsch, C., Kirsch, S. *et al.* (1979) Experimental measurements of the human energy field. In S. Krippner (ed.), *Psychoenergetic Systems: The Interface of Consciousness, Energy and Matter*. New York, Gordon & Breach, 230 .
- Edwards, R., Ibson, M.C., Jessel-Kenyon, J. & Taylor, R.B. (1989) Light emission from the human body. *Complement Med. Res.*, 3:16.
- Esterbauer, H., Zollner, H. & Schaur, R. J. (1990) Aldehydes formed by lipid peroxidation: mechanisms of formation, occurrence, and determination. In *Membrane Lipid Oxidation*. Boca Raton, CRC Press, 283 p.
- Frey, A.H. (1993) Electromagnetic field interactions with biological systems. *FASEB Journal*, 7(2): 272-281.
- Gerardi, G., De Ninno A., Prodocimi, M. *et al.* (2008) Effects of electromagnetic fields of low frequency and low intensity on rat metabolism. *Biomagnetic Research and Technology*, 6: 3.
- Gubkin, A.N. (1978) *Electrets*. Moscow, Nauka. 192.
- Gross, B. (1964) *Charge storage in solid dielectrics; a bibliographical review on the electret and related effects*. New York, Elsevier Pub. Co., 230.
- Goodman, R., Greenbaum, B. & Marron, M.T. (1995) Effects of electromagnetic fields on molecules and cells. *Int. Rev. Cytol.*, 158: 279-338.
- Gulyaev, Yu.V. & Godik, E.E. (1984) On the possibilities of the functional diagnostics of the biological subjects via their temporal dynamics of the infrared images. *USSR Academy Nauk Proceedings/Biophysics*, 277: 1486-1491.
- Gulyaev, Yu.V. & Godik, E.E. (1990) Human and animal physical fields. *Scientific American*, 5: 74-83.
- Gulyaev, Yu.V. & Godik, E.E. (1991) Functional Imaging of the Human Body. *IEEE Engineering in Medicine and Biology*, 10: 21-29.
- Gurwitsch, A.G. (1959) Die mitogenetische strahlung, ihre physikalische-chemischen grundlagen und ihre anwendung in biologie und medizin, Jena, Germany, Veb G. Fisher.
- Gurwitsch, A.G. (1988) A historical review of the problem of mitogenetic radiation. *Experientia*, 44: 545-550.
- Halliwell, B. & Gutteridge, J.M.C. (1989) *Free Radicals in Biology and Medicine* (2nd ed.), Oxford, Clarendon Press.
- Hastings, J.W. (1983). Biological diversity, chemical mechanisms, and the evolutionary origins of bioluminescent systems. *J. Mol. Evol.*, 19 (5): 309-321.



- Holzel, R. & Lamprecht, I. (1994) Wirkungen elektromagnetischer Felder auf biologische Systeme. *Nachrichtentech Elektron*, 44(2): 28–32.
- Ignatov, I., Antonov, A. & Galabova, T. (1998) *Medical Biophysics – Biophysical Fields of Man*. Gea Libris, Sofia: 1–71.
- Ignatov, I., Antonov, A. & Galabova, T., (2002) Scientific Research Studies with Christos Drossinakis (October 2001 – October 2002), *Int. Conference “Man and Nature”*, SRCMB, Sofia.
- Ignatov, I. (2005) *Energy Biomedicine*, Gea-Libris, Sofia, 1–88.
- Marinov, M., Ignatov, I. (2008) Color Kirlian Spectral Analysis. Color Observation with Visual Analyzer, *Euromedica*, Hanover, 57-59.
- Ignatov, I., Tsvetkova, V. (2011) Water for the Origin of Life and “Informationability” of Water, Kirlian (Electric Images) of Different Types of Water, *Euromedica*, Hanover: 62-65.
- Ignatov, I. & Mosin, O.V. (2012) Coronal Effect in Biomedical Diagnostics and Study of Bioenergetical Properties of Biological Objects and Water. *Biomedical Radio electronics, Biomedical Technologies and Radio Electronics*, 12: 13–21 [in Russian].
- Ignatov, I. & Mosin, O.V. (2013a) Method for Color coronal (Kirlian) Spectral Analysis. *Biomedical Radio electronics, Biomedical Technologies and Radio Electronics*, 1: 38–47 [in Russian].
- Ignatov, I. & Mosin O.V. (2013b) Color crown Spectral Kirlian Analysis in the Modeling of Non-equilibrium Conditions with a Gas electric Discharge that Simulates the Primary Atmosphere. *Nano engineering*, 12(30): 3–13 [in Russian].
- Ignatov, I. & Mosin, O.V. (2013c) Structural Mathematical Models Describing Water Clusters. *Journal of Mathematical Theory and Modeling*, 3(11): 72–87.
- Ignatov, I., Mosin, O. V., Niggli, H., Drossinakis, Ch. (2014) Evaluating of Possible Methods and Approaches for Registering of Electromagnetic Waves Emitted from the Human Body, *Advances in Physics Theories and Applications*, 30: 15-33
- Ignatov, I., Mosin, O.V.&Drossinakis, Ch. (2014) Infrared Thermal Field Emitted from Human Body. Thermovision, *Journal of Medicine, Physiology, Biophysics*, 1:1-12.
- Ignatov, I.&Mosin,O.V. (2014) Mathematical Models of Distribution of Water Molecules Regarding Energies of Hydrogen Bonds, *Journal of Medicine, Physiology and Biophysics*, 2: 71-94.
- Ignatov, I. &Mosin,O.V. (2014) Mathematical Models Describing Water Clusters as Interaction among Water Molecules. Distributions of Energies of Hydrogen Bonds, *Journal of Medicine, Physiology and Biophysics*, 3: 48-70.
- Ignatov, I., Mosin, O. V., Niggli, H., Drossinakis, Ch.&Stoyanov, Ch. (2014) Registration of Electromagnetic Waves Emitted the Human Body, *Journal of Medicine, Physiology and Biophysics*, 5: 1-22.
- Ignatov, I., Mosin, O. V. (2014) Coronal Gas Discharge Effect in Modeling of Non-Equilibrium Conditions with Gas Electric Discharge Simulating Primary Atmosphere and Hydrosphere for Origin of Life and Living Matter, *Journal of Medicine, Physiology and Biophysics*, 5: 47-70.
- Ignatov, I., Mosin, O.V. & Stoyanov, Ch. (2014) Biophysical Fields. Color Coronal Spectral Analysis. Registration with Water Spectral Analysis. Biophoton Emission, *Journal of Medicine, Physiology and Biophysics*, 6: 1-22.
- Ignatov, I., Mosin, O. V., Niggli, H., Drossinakis, Ch. (2014) Evaluating of Possible Methods and Approaches for Registering Electromagnetic Waves Emitted from the Human Body, *Nanotechnology Research and Practice*, 2 (2): 96-116.
- Ignatov, I., Mosin, O. V. (2014) The Methods for Studying the Structure of Water Clusters (H<sub>2</sub>O), where n=3-20. Water Clusters as Nano-structures, *Journal of Health, Medicine and Nursing*, 8: 29-58
- Ignatov, I., Mosin, O. V. (2014) The Methods for Studying the Structure of Water Clusters (H<sub>2</sub>O), where n=3-20, Application in Medicine, *Journal of Health, Medicine and Nursing*, 7: 23-52.
- Ignatov, I., Mosin, O. V., Stoyanov, Ch. (2014) Fields in Electromagnetic Spectrum Emitted from Human Body. Application in Medicine, *Journal of Health, Medicine and Nursing*, 7 (1-22).
- Inaba, H. (1988). Super-high sensitivity systems for detection and spectral analysis of ultraweak photon emission from biological cells and tissues. *Experientia*, 44: 550–559.
- Ignatov, I.& Mosin, O.V. (2016) Results of Bioinfluence of Dimitar Risimanski with Biophysical Model Systems, *Journal of Medicine, Physiology and Biophysics*: 24: 5-17.
- Ignatov, I., Mosin, O.V. (2016) Biophysical Results of Bioinfluence of Dimitar Risimanski as Base of Medical Effects, *Journal of Health, Medicine and Nursing*: 27: 24-35.
- Ignatov, I., Mosin, O.V. (2016) Structural Alterations among Water Molecules after Bioinfluence of Dimitar Risimanski, *Advances in Physics Theories and Applications*, Vol. 57, pp.20-44.
- Kim, T.J (2002) Biophoton emission from fingernails and fingerprints of living human subjects. *Acupuncture Electrother Res.*, 27:85.
- Kirlian, S.D. (1949) Method for receiving photographic pictures of different types of objects. USSR Patent №

- 106401.
- Inaba, H. (2000) Measurement of biophotons from human body. *J. Int. Soc. Life Inf. Sci.*, 18:448.
- Liboff, A.R., Williams, T., Strong, D.M. & Wistar, R. (1984) Time-varying magnetic fields: effect on DNA synthesis. *Science*, 223: 818–820.
- Kiang, J.G., Ives J.A. & Jonas, W.B. (2005) External bioenergy-induced increases in intracellular free calcium concentrations are mediated by  $\text{Na}^+/\text{Ca}^{2+}$  exchanger and L-type calcium channel. *Mol. Cell Biochem.*, 271:51.
- Kwan-Hoong, Ng. (2003) Non-ionizing radiations – sources, biological effects, emissions and exposures. *Proceedings of the International Conference on Non-Ionizing Radiation at UNITEN (ICNIR2003). Electromagnetic Fields and Our Health.* 20–22 October 2003.
- Lin, S., Chevalier, G., Lin, H., Ross, T. & Lin, T. (2006). Measurement of biophoton emission with a single photon counting system. *J. Altern. Complement. Med.*, 12:210.
- Marino, A.A (Ed.) (1988) *Modern Bioelectricity*. Mariele Dekker, New York, Basel, ISBN 0-8247-7788-3.
- Marinov, M. & Ignatov, I. (2008) Color Kirlian spectral analysis. Color observation with visual analyzer. *Euromedica*, Hanover, 57–59.
- Miller, M.W. (1986) Extremely low frequency (ELF) electric fields: experimental work on biological effects. *CRC Handbook of biological effects of electromagnetic fields*, 138–168.
- Mosin, O.V. (2011) Magnetic devices for water treatment. *C.O.K. Publishing House "Media Technology" (Moscow)*, 6: 24–27 [in Russian].
- Mosin, O.V. (2012) Advanced technologies and equipment for magnetic water treatment (review). *Water supply and sanitary technique*, 8: 12–32 [in Russian].
- Motohiro, T. (2004). Biophoton detection as a novel technique for cancer imaging. *Cancer Science*, 95(8): 656–661.
- Niggli, H. (1993). Artificial sunlight irradiation induces ultra weak photon emission in human skin fibroblasts, *Journal of Photochemistry and Photobiology B: Biology*, 18 (2–3): 281–285.
- Nikolaev, Y.A. (2000) Distant Interactions in Bacteria. *Microbiology*, 69(5): 497–503.
- Pehek, J.O., Kyler, H.J & Faust, D.L. (1976). Image modulating Corona discharge photography, *Science*, 194(4262): 263–270.
- Popp, F.A., Li, K. & Gu, Q. (1992) Recent advances in biophoton research and its application, *World scientific*, 1–18.
- Popp, F.A., Quao, G. & Ke-Hsuen, L. (1994) Biophoton emission: experimental background and theoretical approaches, *Modern physics Letters B.*, 8: 21–22.
- Popp, F.A., Chang, J.J., Herzog, A., Yan, Z. & Yan, Y. (2002) Evidence of non-classical (squeezed) light in biological systems. *Physics Letters A.*, 293(1–2): 98–102.
- Popp, F.A. (2005) Essential differences between coherent and non-coherent effects of photon emission from living organisms. In: X. Shen, R. van Wijk (eds). *Biophotonics*. New York: Springer, 124 p.
- Porter, N.A. & Wujek, D.G. (1988) *Reactive Oxygen Species in Chemistry, Biology, and Medicine*. In A. Quintanilha, Ed. New York, Plenum Press, pp. 55–79.
- Rauhut, M.M. (1985) Chemiluminescence. In: M. Grayson (Ed). *Kirk-Othmer Concise Encyclopedia of Chemical Technology* (3rd ed). New York, John Wiley and Sons, ISBN 0-471-51700-3, 247 p.
- Rattemeyer, M., Popp, F.A. & Nagl, W. (1981) Evidence of photon emission from DNA in living systems, *Nature Wissenshanften*, 68(11): 572–573.
- Ring, E.F.J. & Hughes, H. (1986) Real time video thermography in recent developments in medical and physiological imaging. *Suppl. Journal of Medical Engineering and Technology*, 86–89.
- Rubik, B. (2002) The biofield hypothesis: its biophysical basis and role in medicine. *J. Altern. Complement Med.*, 8( 6 ):703-717.
- Sessler, G.M. & Gerhard-Multhaupt, R. (eds) (1998) *Electrets*. Laplacian Press, Morgan Hill, California, USA, ISBN 1-885540-07-8.
- Seto, A., Kusaka, C., Nakazato, S. et al. (1992) Detection of extraordinary large bio-magnetic field strength from human hand. *Acupuncture Electrother Res. Int. J.*, 17:75.
- Shimizu H., Suzuki, Y. & Okonogi, H. (1995) Biological effects of electromagnetic fields. *Nippon Eiseigaki Zasshi.*, 50(6): 919–931.
- Sisodia, M.L. (2007). *Microwaves: introduction to circuits, devices and antennas*. New Delhy, New Age International Ltd., ISBN 8122413382, 602 p.
- Vladimirov, Y.A. (1996) Studies of antioxidants with chemiluminescence. In: *Proceedings of the International Symposium on Natural Antioxidants. Molecular Mechanisms and Health Effects.* L. Packer, M.G. Traber & W. Xin (eds.). 125-144.
- Wikswow, J. & Barach, J. (1980) An estimation of the steady magnetic field strength required to influence nerve condition. *IEEE Trans. Bio-Med. Eng.*, 27: 722–723.

- Young, R.E. & Roper, C.F. (1976) Bioluminescent countershading in midwater animals: evidence from living squid, *Science*, 191(4231): 1046–1048.
- Zhadin, M.N. (2001) Review of russian literature on biological action of DC and low-frequency AC magnetic fields. *Bioelectromagnetics*, 22: 27– 45.
- Zlatkevich, L. & Kamal-Eldin, A. (2005) *Analysis of Lipid Oxidation*. In: A. Kamal-Eldin & J. Pokorn (Eds.). New York, AOCS Publishing, 281.